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(71) Applicant:
**Kyowa Hakko Kogyo Co., Ltd.
Chiyoda-ku, Tokyo 100 (JP)**

(72) Inventors:
• **Kazuyasu, Nakamura
Machida-shi, Tokyo (JP)**

• **Nobuo, Hanai
Sagamihara-shi, Kanagawa (JP)**

(74) Representative:
**Kinzebach, Werner, Dr. et al
Patentanwälte
Reitstötter, Kinzebach und Partner
Postfach 86 06 49
81633 München (DE)**

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(54) **Human complementarity determining region (CDR)-grafted antibody to ganglioside gm2**

(57) A human CDR-grafted antibody which specifically reacts with ganglioside GM₂, wherein said antibody comprises CDR 1, CDR 2 and CDR 3 of heavy chain (H chain) variable region (V region) comprising amino acid sequences described in SEQ ID NO:1, SEQ ID NO:2 and SEQ ID NO:3, and CDR 1, CDR 2 and CDR 3 of light chain (L chain) V region comprising amino acid sequences described in SEQ ID NO:4, SEQ ID NO:5 and SEQ ID NO:6, and wherein at least one of the frameworks (FR) of said H chain and L chain V regions comprises an amino acid sequence selected from common sequences (HMHCS; human most homologous consensus sequence) derived from human antibody subgroups.

EP 0 882 794 A2

Description

FIELD OF THE INVENTION

5 This invention relates to a human complementarity determining region (referred to as "CDR" hereinafter) grafted antibody to ganglioside GM₂ (referred to as "GM₂" hereinafter). This invention also relates to a DNA fragment encoding the above-described antibody, particularly its variable region (referred to as "V region" hereinafter). This invention relates to an expression vector which contains the DNA fragment and to a host transformed with the expression vector. This invention further relates to a method for the production of the human CDR-grafted antibody specific for GM₂ and 10 to its therapeutic and diagnostic use.

BACKGROUND OF THE INVENTION

It is known in general that, when a mouse antibody is administered to human, the mouse antibody is recognized as 15 foreign matter in the human body and thus induces a human antibody to a mouse antibody (human anti-mouse antibody, referred to as "HAMA" hereinafter) which reacts with the administered mouse antibody to produce adverse effects (Dillman, R.O. *et al.*, *J. Clin. Oncol.*, 2, 881 (1984); Meeker, T.C. *et al.*, *Blood*, 65, 1349 (1985); LoBuglio, A.F. *et al.*, *J. Natl. Cancer Inst.*, 80, 932 (1988); Houghton, A.N. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 82, 1242 (1985)), and the administered mouse antibody is quickly cleared (Pimm, M.V. *et al.*, *J. Nucl. Med.*, 26, 1011 (1985); Meeker, T.C. *et al.*, 20 *Blood*, 65, 1349 (1985); Khazaeli, M.B. *et al.*, *J. Natl. Cancer Inst.*, 80, 937 (1988)) to reduce effects of the antibody (Shawler, D.L. *et al.*, *J. Immunol.*, 135, 1530 (1985); Courtenay-Luck, N.S. *et al.*, *Cancer Res.*, 46, 6489 (1986)).

In order to solve these problems, attempts have been made to convert a mouse antibody into a humanized antibody such as a human chimeric antibody or a human CDR-grafted antibody. The human chimeric antibody is an antibody in which its V region is derived from an antibody of nonhuman animal and its constant region (referred to as "C region" 25 hereinafter) is derived from a human antibody (Morrison, S.L. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 81, 6851 (1984)). Furthermore, it is reported that, when this type of antibody is administered to human, HAMA is hardly induced and its half-life in blood increases six times (LoBuglio, A.F. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 86, 4220 (1989)). The human CDR-grafted antibody is an antibody in which the CDR of human antibody is replaced by other CDR derived from nonhuman animal (Jones, P.T. *et al.*, *Nature*, 321, 522 (1986)), which is also called a reshaped human antibody. It is reported that, 30 in a test of a human CDR-grafted antibody in monkeys, its immunogenicity is reduced and its half-life in blood is increased four to five times, in comparison with a mouse antibody (Hakimi, J. *et al.*, *J. Immunol.*, 147, 1352 (1991)).

Also, with regard to the cytotoxicity of antibodies, it is reported that the Fc region of a human antibody activates human complement and human effector cells more effectively than the Fc region of mouse antibody. For example, it is reported that human effector cell-mediated anti-tumor effects of a mouse antibody to GD₂ is increased when the antibody is converted into a human chimeric antibody having human antibody Fc region (Mueller, B.M. *et al.*, *J. Immunol.*, 35 144, 1382 (1990)), and similar results are reported on a human CDR-grafted antibody to CAMPATH-1 antigen (Reichmann, L. *et al.*, *Nature*, 332, 323 (1988)). These results indicate that humanized antibodies are more desirable than mouse antibodies as antibodies to be clinically used in human.

Ganglioside as a glycolipid having sialic acid is a molecule which constitutes an animal cell membrane, and comprises a carbohydrate chain as a hydrophilic side chain and sphingosine and fatty acid as hydrophobic side chains. It 40 is known that types and expression quantities of ganglioside vary depending on the cell species, organ species, animal species and the like. It is known also that the expression of ganglioside changes quantitatively and qualitatively in the process of cancer development of cells (Hakomori, S. *et al.*, *Cancer Res.*, 45, 2405 (1985)). For example, it is reported that gangliosides GD₂, GD₃, GM₂ and the like which are hardly observed in normal cells are expressed in nerve ecto- 45 derm system tumors considered to have high malignancy, such as neuroblastoma, pulmonary small cell carcinoma and melanoma (Pukel, C.S. *et al.*, *J. Exp. Med.*, 155, 1133 (1982); Nudelman, E. *et al.*, *J. Biol. Chem.*, 257, 12752 (1982); Werkmeister, J.A. *et al.*, *Cancer Res.*, 47, 225 (1987); Mujoo, K. *et al.*, *Cancer Res.*, 47, 1098 (1987); Cheung, N.V. *et al.*, *Cancer Res.*, 45, 2642 (1985); Tai, T. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 80, 5392 (1983)), and antibodies to these gangliosides are considered to be useful for diagnosis and treatment of various cancers in human.

50 It is indicated that human antibodies to GM₂ are useful for treatment of human melanoma (Irie, R.F. *et al.*, *Lancet*, 1, 786 (1989)). However, the antibodies to GM₂ so far reported are either those which are derived from nonhuman animal or a human antibody belonging to the IgM class (Natoli, E.J. *et al.*, *Cancer Res.*, 46, 4116 (1986); Miyake, M. *et al.*, *Cancer Res.*, 48, 6154 (1988); Cahan, L.D. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 79, 7629 (1982); Fredman, P. *et al.*, *J. Biol. Chem.*, 264, 12122 (1989)). The antibody of the IgM class, however, is unsuitable for applying to human, because 55 it has a pentameric structure having a large molecular weight (about 900,000) in comparison with the antibody of IgG class which has a molecular weight of about 150,000, thus posing a problem in carrying out its purification, in addition to other problems such as its short half-life in blood and weak anti-tumor effect (Bernstein, I.D. *et al.*, *Monoclonal Antibodies*, Plenum Press, p.275 (1980)).

Because of the above, it is desirable to develop a humanized antibody to GM₂ of the IgG class which, when applied to human, does not induce HAMA in the human body, causes less adverse effects, shows prolonged half-life in blood and has improved anti-tumor effect, so that its high diagnostic and therapeutic effects on human cancers can be expected.

- 5 The inventors of the present invention disclose in JP-A-6-205694 (the term "JP-A" as used herein means an "unexamined published Japanese patent application") (corresponding to EP-A-0 598 998) a method for producing an IgG class human chimeric antibody and a human CDR-grafted antibody, which can specifically reacts with GM₂ and are useful for diagnosis and treatment of human cancers. However, there are no reports on a human CDR-grafted antibody which, when compared with a human chimeric antibody, has similar levels of binding activity and binding specificity for GM₂ and anti-tumor effects upon GM₂-positive cells.

SUMMARY OF THE INVENTION

- As described in the foregoing, it is considered that human CDR-grafted antibodies are useful for diagnosis and treatment of human cancers and the like. However, the antibody activity is reduced when the CDRs of the heavy chain (referred to as "H chain" hereinafter) V region and light chain (referred to as "L chain" hereinafter) V region of an antibody of nonhuman animal are replaced only with the CDRs of the H chain V region and L chain V region of a human antibody, so that great concern has been directed toward the establishment of a method for the production of a human CDR-grafted antibody to GM₂ belonging to the IgG class (referred to as "human CDR-grafted anti-GM₂ antibody" hereinafter) which, when compared with a human chimeric antibody, has similar levels of binding activity and binding specificity for GM₂ and anti-tumor effects upon GM₂-positive cells, as well as a method for producing a human CDR-grafted antibody, which can be applied to all antibodies.

- This invention relates to a human CDR-grafted antibody which specifically reacts with ganglioside GM₂, wherein said antibody comprises CDR 1, CDR 2 and CDR 3 of H chain V region comprising amino acid sequences of SEQ ID NO:1, SEQ ID NO:2 and SEQ ID NO:3 or functional equivalents thereof, and CDR 1, CDR 2 and CDR 3 of L chain V region comprising amino acid sequences of SEQ ID NO:4, SEQ ID NO:5 and SEQ ID NO:6 or functional equivalents thereof, and wherein at least one of the frameworks (referred to as "FR" hereinafter) of said H chain and L chain V regions comprises an amino acid sequence selected from common sequences (human most homologous consensus sequence (referred to as "HMHCS" hereinafter) derived from human antibody subgroups.

- 30 Furthermore, the present invention relates to the above human CDR-grafted antibody, wherein said FR of H chain or L chain V region of the human CDR-grafted antibody comprises an amino acid sequence in which at least one amino acid is replaced by an other amino acid, and wherein said antibody has antigen-binding activity, binding specificity, antibody dependent cell mediated cytotoxicity (ADCC), and complement dependent cytotoxicity (CDC) comparable to those of a human chimeric antibody having a V region of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂.

Moreover, the present invention relates to the above human CDR-grafted antibody, wherein said H chain C region of the antibody is derived from an antibody belonging to the human antibody IgG class.

BRIEF EXPLANATION OF THE DRAWINGS

- 40 Fig. 1 shows a construction scheme for a plasmid named pBSA.
 Fig. 2 shows a construction scheme for a plasmid named pBSAE.
 Fig. 3 shows a construction scheme for a plasmid named pBSH-S.
 Fig. 4 shows a construction scheme for a plasmid named pBSK-H.
 45 Fig. 5 shows a construction scheme for plasmids named pBSH-SA and pBSK-HA.
 Fig. 6 shows a construction scheme for plasmids named pBSH-SAE and pBSK-HAE.
 Fig. 7 shows a construction scheme for plasmids named pBSH-SAE and pBSK-HAE.
 Fig. 8 shows a construction scheme for a plasmid named pBSK-HAEESal.
 Fig. 9 shows a construction scheme for a plasmid named pBSX-S.
 50 Fig. 10 shows a construction scheme for a plasmid named pBSX-SA.
 Fig. 11 shows a construction scheme for a plasmid named pBSSC.
 Fig. 12 shows a construction scheme for a plasmid named pBSMo.
 Fig. 13 shows a construction scheme for a plasmid named pBSMoS.
 Fig. 14 shows a construction scheme for a plasmid named pChilgLA1S.
 55 Fig. 15 shows a construction scheme for a plasmid named pMohCκ.
 Fig. 16 shows a construction scheme for a plasmid named pBSMoSal.
 Fig. 17 shows a construction scheme for a plasmid named pBSMoSalS.
 Fig. 18 shows a construction scheme for a plasmid named pBSHCγ1.

Fig. 19 shows a construction scheme for a plasmid named pMohCy1.

Fig. 20 shows a construction scheme for a plasmid named pMoy1SP.

Fig. 21 shows a construction scheme for a plasmid named pMox1SP.

Fig. 22 shows a construction scheme for a plasmid named pKANTEX93.

5 Fig. 23 shows a construction scheme for a plasmid named pBSNA.

Fig. 24 shows a construction scheme for a plasmid named pBSH3.

Fig. 25 shows a construction scheme for a plasmid named pBSES.

Fig. 26 shows a construction scheme for a plasmid named pBSL3.

Fig. 27 shows a construction scheme for a plasmid named pKANTEX796H.

10 Fig. 28 shows a construction scheme for a plasmid named pKANTEX796.

Fig. 29 shows a construction scheme for a plasmid named pT796.

Fig. 30 is a graphic representation of transient mouse-human chimeric anti-GM₂ antibody expression by the plasmids pKANTEX796 and pT796. The ordinate denotes the antibody concentration that showed GM₂-binding activity, and the abscissa denotes the time after introduction of the plasmid.

15 Fig. 31 shows a construction scheme for a plasmid named pBSH10.

Fig. 32 shows a construction scheme for a plasmid named pBSL16.

Fig. 33 illustrates a process for mutagenesis by PCR and a process for cloning DNA fragments mutated.

Fig. 34 shows a construction scheme for a plasmid named pBSLV1+2.

Fig. 35 shows a construction scheme for a plasmid named pBSLm-28.

20 Fig. 36 shows a construction scheme for a plasmid named pBSHSGL.

Fig. 37 shows a construction scheme for a plasmid named pT796LCDR.

Fig. 38 shows a construction scheme for plasmids named pT796HLCDR, pT796HLCDRHV2 and pT796HLCDRHV4.

Fig. 39 shows a construction scheme for a plasmid named pT796HLCDRH10.

25 Fig. 40 shows construction scheme for plasmids named pT796HCDR, pT796HCDRHV2, pT796HCDRHV4 and pT796HCDRH10.

Fig. 41 is a graphic representation of the results of human CDR-grafted anti-GM₂ antibody activity evaluation in terms of transient expression as obtained using the plasmids pT796, pT796HCDR, pT796HCDRHV2, pT796HCDRHV4 and pT796HCDRH10. The ordinate denotes the plasmid used, and the abscissa denotes the relative activity value with the activity obtained with the chimera antibody being taken as 100%.

30 Fig. 42 shows a construction scheme for plasmids named pT796HLCDRLV1, pT796HLCDRLV2, pT796HLCDRLV3, pT796HLCDRLV4, pT796HLCDRLV8, pT796HLCDRLVm-2, pT796HLCDRLVm-8, pT796HLCDRLVm-28 and pT796HLCDRHSGL.

Fig. 43 is a graphic representation of the results of human CDR-grafted anti-GM₂ antibody activity evaluation in terms of transient expression as obtained using the plasmids pT796, pT796HLCDR, pT796HLCDRLV1, pT796HLCDRLV2, pT796HLCDRLV3, pT796HLCDRLV4, pT796HLCDRLV8, pT796HLCDRLVm-2, pT796HLCDRLVm-8, pT796HLCDRLVm-28 and pT796HLCDRHSGL. The ordinate denotes the plasmid used, and the abscissa denotes the relative activity value with the activity obtained with the chimera antibody being taken as 100%.

40 Fig. 44 shows a construction scheme for plasmids named pKANTEX796HLCDRLVm-28 and pKANTEX796HLCDRHSGL.

Fig. 45 shows electrophoretic patterns obtained for mouse-human chimeric anti-GM₂ antibody KM966 and purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 by SDS-PAGE (4 to 15% gradient gels used). The patterns shown on the left side are those obtained under reducing conditions, and those on the right under nonreducing conditions. From the left of each lane, the electrophoretic patterns for high-molecular-weight marker, KM966, KM8966, KM8967, low-molecular-weight marker, KM966, KM8966 and KM8967 are shown in that order.

Fig. 46 is a graphic representation of the GM₂-binding activities of mouse-human chimeric anti-GM₂ antibody KM966 and purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967. The ordinate denotes the GM₂-binding activity, and the abscissa the antibody concentration.

50 Fig. 47 is a graphic representation of the reactivities of mouse-human chimeric anti-GM₂ antibody KM966 and purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 against various gangliosides. The ordinate denotes the ganglioside species, and the abscissa the binding activity. AcGM₂ stands for N-acetyl-GM₂, GcGM₂ for N-glycolyl-GM₂, AcGM₃ for N-acetyl-GM₃ and GcGM₃ for N-glycolyl-GM₃.

Fig. 48 is a graphic representation of the reactivities of mouse-human chimeric anti-GM₂ antibody KM966 and purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 against the human lung small cell carcinoma cell line SBC-3. The ordinate denotes the number of cells, and the abscissa the fluorescence intensity. From the lowermost graph, the reactivities of control, KM8967, KM8966 and KM966 are shown in that order.

Fig. 49 graphically shows the CDC activities of mouse-human chimeric anti-GM₂ antibody KM966 and purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 against the human lung small cell carcinoma cell line

SBC-3. The ordinate indicates the cytotoxic activity and the abscissa the concentration of the antibody.

Fig. 50 graphically shows the ADCC activities of mouse-human chimeric anti-GM₂ antibody KM966 and purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 against the human lung small cell carcinoma cell line SBC-3. The ordinate indicates the cytotoxicity and the abscissa the concentration of the antibody.

Fig. 51 shows a construction scheme for plasmids, pKANTEX796HM1Lm-28, pKANTEX796HM2Lm-28, pKANTEX796HM3Lm-28, pKANTEX796HM31Lm-28 and pKANTEX796HM32Lm-28.

Fig. 52 shows the electrophoretic patterns in SDS-PAGE (using 4-15% gradient gels) of mouse-human chimeric anti-GM₂ antibody KM966, human CDR-grafted anti-GM₂ antibody KM8966 and human CDR-grafted anti-GM₂ antibodies each having various types of substitution. The pattern obtained under nonreducing conditions is shown on the left side and that obtained under reducing conditions on the right side. M stands for molecular weight markers (from the top, the arrows indicate the molecular weight of 205 Kd, 140 Kd, 83 Kd, 45 Kd, 32.6 Kd, 18 Kd and 7.5 Kd in that order) and 1, 2, 3, 4, 5, 6 and 7 stand for the electrophoretic patterns of KM966, KM8966, M1-28, M2-28, M3-28, M31-28 and M32-28, respectively.

Fig. 53 graphically shows the CDC activities of mouse-human chimeric anti-GM₂ antibody KM966, human CDR-grafted anti-GM₂ antibody KM8966 and human CDR-grafted anti-GM₂ antibodies each having various types of substitution against the human lung small cell carcinoma cell line SBC-3. The ordinate indicates the cytotoxic activity and the abscissa the concentration of the antibody.

Fig. 54 shows a construction scheme for plasmids, pKANTEX796HLM-28 No.1, pKANTEX796HM1Lm-28 No.1, pKANTEX796HM2Lm-28 No.1 and pKANTEX796HM3Lm-28 No.1.

Fig. 55 shows the electrophoretic patterns in SDS-PAGE (using 4-15% gradient gels) of mouse-human chimeric anti-GM₂ antibody KM966 and human CDR-grafted anti-GM₂ antibodies each having various types of substitution. The pattern obtained under nonreducing conditions is shown on the left side and that obtained under reducing conditions on the right side. M stands for molecular weight markers (from the top, the arrows indicate the molecular weight of 205 Kd, 140 Kd, 83 Kd, 45 Kd, 32.6 Kd, 18 Kd and 7.5 Kd in that order) and 1, 2, 3, 4 and 5 stand for the electrophoretic patterns of KM966, h796H-No.1, M1-No.1, M2-No.1 and M3-No.1, respectively.

Fig. 56 graphically shows the CDC activities of mouse-human chimeric anti-GM₂ antibody KM966, human CDR-grafted anti-GM₂ antibodies KM8966 and KM8970 and human CDR-grafted anti-GM₂ antibodies each having various types of substitution against the human lung small cell carcinoma cell line SBC-3. The ordinate indicates the cytotoxic activity and the abscissa the concentration of the antibody.

Fig. 57 graphically shows the GM₂-binding activities of mouse-human chimeric anti-GM₂ antibody KM966 and human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970. The ordinate indicates the GM₂-binding activity and the abscissa the concentration of the antibody.

Fig. 58 graphically shows the reactivities of mouse-human chimeric anti-GM₂ antibody KM966 and human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 against various gangliosides. The ordinate indicates the ganglioside species and the abscissa the binding activity. AcGM₂ stands for N-acetyl-GM₂, GcGM₂ for N-glycolyl-GM₂, AcGM₃ for N-acetyl-GM₃ and GcGM₃ for N-glycolyl-GM₃.

Fig. 59 graphically shows the reactivities of mouse-human chimeric anti-GM₂ antibody KM966 and human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 against the human lung small cell carcinoma cell line SBC-3. The ordinate indicates the number of cells and the abscissa the fluorescence intensity. From the lowermost graph, the reactivities of control, KM966, KM8970 and KM8969 are shown in that order.

Fig. 60 graphically shows the ADCC activities of mouse-human chimeric anti-GM₂ antibody KM966 and human CDR-grafted anti-GM₂ antibodies KM8966, KM8969 and KM8970 against the human lung small cell carcinoma cell line SBC-3. The ordinate indicates the cytotoxicity and the abscissa the concentration of the antibody.

Fig. 61 graphically shows the CDC activities of mouse-human chimeric anti-GM₂ antibody KM966 and human CDR-grafted anti-GM₂ antibodies KM8966, KM8969 and KM8970 against the human lung small cell carcinoma cell line SBC-3 obtained when the reaction was carried out for 1 hour and 4 hours after the addition of the human complement. The ordinate indicates the cytotoxicity and the abscissa the concentration of the antibody.

DETAILED DESCRIPTION OF THE INVENTION

In the human CDR-grafted antibody, only CDRs of the H chain and L chain V regions comprise amino acid sequences of an antibody derived from nonhuman animal, and FRs of the H and L chain V regions and the C region comprise of amino acid sequences of a human antibody. Examples of the nonhuman animal include mouse, rat, hamster, rabbit and the like, as long as a hybridoma can be prepared therefrom.

With regard to the FR of the V regions of H chain and L chain, any amino acid sequence of known human antibodies can be used, such as an amino acid sequence selected from human antibody amino acid sequences, HMHCS, registered at the Protein Data Bank. Preferably, an amino acid sequence of the FR of HMHCS, which has a high homology with the FR of a monoclonal antibody of nonhuman animal, may be used.

As described in the foregoing, the antibody activity is reduced when the CDRs of the H chain V region and L chain V region of an antibody of nonhuman animal are replaced only with the CDRs of the H chain V region and L chain V region of a human antibody. In consequence, the present invention relates to a human CDR-grafted antibody wherein at least one amino acid in the FR of H chain and L chain V regions of the human CDR-grafted antibody is replaced by an other amino acid, so that it can show certain levels of antigen-binding activity, binding specificity and antibody dependent cell mediated cytotoxicity (ADCC), as well as complement dependent cytotoxicity (CDC), which are comparable to those of a human chimeric antibody having the V region of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂, and to a method for producing the same.

The replacement of at least one amino acid in the FR of H chain and L chain V regions of the human CDR-grafted antibody of the present invention means that amino acid residues desired to be replaced in the FR of H chain and L chain V regions of the human CDR-grafted antibody having a human antibody amino acid sequence are replaced by an other amino acid residues at corresponding positions in the FR of H chain and L chain V regions of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂. For example, at least one amino acid of positions 38, 40, 67, 72, 84 and 98 in the FR of H chain V region and positions 4, 11, 15, 35, 42, 46, 59, 69, 70, 71, 72, 76, 77 and 103 in the FR of L chain V region is replaced by an other amino acid.

Mouse anti-GM₂ monoclonal antibody KM796 (FERM BP-3340, JP-A-4-311385) can be cited as an example of the monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂. A chimeric anti-GM₂ antibody KM966 (FERM BP-3931, JP-A-6-205694) can be cited as an example of the human chimeric antibody having the V region of a monoclonal antibody which is derived from nonhuman animal which specifically reacts with ganglioside GM₂.

Examples of the antibody having certain levels of antigen-binding activity, binding specificity and antibody dependent cell mediated cytotoxicity (ADCC), which are comparable to those of a human chimeric antibody having a V region of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂ include KM8966 produced by a transformant cell line KM8966 (FERM BP-5105), KM8967 produced by a transformant cell line KM8967 (FERM BP-5106) and KM8970 produced by a transformant cell line KM8970 (FERM BP-5528).

KM8969 produced by a transformant cell line KM8969 (FERM BP-5527) can be cited as an example of the antibody having certain levels of antigen-binding activity, binding specificity, antibody dependent cell mediated cytotoxicity (ADCC) and complement dependent cytotoxicity (CDC), which are comparable to those of a human chimeric antibody having a V region of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂.

A method for producing the human CDR-grafted anti-GM₂ antibody is discussed below.

1. Construction of humanized antibody expression vector

The humanized antibody expression vector is an expression vector for use in animal cells, in which cDNA molecules encoding the C regions of H chain and L chain of a human antibody are integrated, and can be constructed by inserting the cDNA molecules encoding the C regions of H chain and L chain of a human antibody into respective expression vectors for animal cell use or by inserting the cDNA molecules which encode the C regions of H chain and L chain of a human antibody into a single expression vector for animal cell use (such a vector is called a tandem cassette vector). The C regions of human antibody can be any of C regions of human antibody H chain and L chain, and examples thereof include γ 1 type C region (referred to as " γ 1" hereinafter) and γ 4 type C region (referred to as " γ 4" hereinafter) of the human antibody H chain and κ type C region (referred to as " κ " hereinafter) of the human antibody L chain. Any expression vector for animal cell use can be used, as long as the cDNA encoding the human antibody C region can be integrated and expressed. Examples thereof include pAGE107 (Miyaji, H. *et al.*, *Cytotechnology*, **3**, 133 (1990)), pAGE103 (Mizukami, T. *et al.*, *J. Biochem.*, **101**, 1307 (1987)), pHSG274 (Brady, G. *et al.*, *Gene*, **27**, 223 (1984)), pKCR (O'Hare, K. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, **78**, 1527 (1981)), and pSG1 β d2-4 (Miyaji, H. *et al.*, *Cytotechnology*, **4**, 173 (1990)). Examples of the promoter and enhancer to be used in the expression vector for animal cell use include early promoter and enhancer of SV40 (Mizukami, T. *et al.*, *J. Biochem.*, **101**, 1397 (1987)), LTR promoter and enhancer of Moloney mouse leukemia virus (Kuwana, Y. *et al.*, *Biochem. Biophys. Res. Comm.*, **149**, 960 (1987)) and promoter (Mason, J.O. *et al.*, *Cell*, **41**, 479 (1985)) and enhancer (Gillies, S.D. *et al.*, *Cell*, **33**, 717 (1983)) of immunoglobulin H chain. The thus constructed humanized antibody expression vector can be used for expressing the human chimeric antibody and human CDR-grafted antibody in animal cells.

2. Preparation of cDNA encoding the V region of antibody of nonhuman animal

The cDNA encoding the H chain V region and L chain V region of the antibody of nonhuman animal to GM₂ is obtained in the following manner.

cDNA molecules are synthesized by extracting mRNA from cells of a hybridoma which produces the anti-GM₂ mon-

oclonal antibody. A library is prepared from the thus synthesized cDNA using a phage or a plasmid. Using cDNA corresponding to the C region moiety or cDNA corresponding to the V region moiety of each chain of a mouse antibody as a probe, a recombinant phage or recombinant plasmid having a cDNA which encodes the V region of H chain or a recombinant phage or recombinant plasmid having a cDNA encoding the V region of L chain is isolated from the library, and complete nucleotide sequences of the intended H chain V region and L chain V region of the antibody on the recombinant phage or recombinant plasmid are determined. Complete amino acid sequences of the H chain V region and L chain V region are deduced from the thus determined nucleotide sequences.

KM796 (FERM BP-3340, JP-A-4-311385) can be cited as an example of the hybridoma cells which produce the anti-GM₂ monoclonal antibody.

The guanidine thiocyanate-caesium trifluoroacetate method [*Methods in Enzymol.*, 154, 3 (1987)] can be exemplified as a method for preparing total RNA from hybridoma cells KM796, and the oligo (dT) immobilized cellulose column method [*Molecular Cloning; A Laboratory Manual* (2nd ed.)] can be exemplified as a method for preparing poly(A)⁺ RNA from the total RNA. As a kit for use in the preparation of mRNA from the hybridoma KM796 cells, Fast Track mRNA Isolation Kit; manufactured by Invitrogen), Quick Prep mRNA Purification Kit; manufactured by Pharmacia) or the like can be exemplified.

With regard to the method for synthesizing cDNA and preparing cDNA library, the methods described in *Molecular Cloning; A Laboratory Manual* (2nd ed.) and *Current Protocols in Molecular Biology*, supplements 1 - 34 and the like, or a method which uses a commercially available kit such as Super Script™ Plasmid System for cDNA Synthesis and Plasmid Cloning (manufactured by Life Technologies) or Zap-cDNA Synthesis Kit (manufactured by Stratagene) can be exemplified. In preparing a cDNA library, any vector can be used as the vector into which the cDNA synthesized using the mRNA extracted from the hybridoma cells KM796 is to be integrated, as long as the cDNA can be integrated therein. Examples of such vectors include ZAP Express [*Strategies*, 5, 58 (1992)], pBluescript II SK(+) [*Nucleic Acids Research*, 17, 9494 (1989)], λzap II (manufactured by Stratagene), λgt10, λgt11 [*DNA Cloning, A Practical Approach*, Vol. 1, 49 (1985)], Lambda BlueMid (manufactured by Clontech), λExCell, pT7T3 18U (manufactured by Pharmacia), pcD2 [*Mol. Cell. Biol.*, 3, 280 (1983)] and pUC18 [*Gene*, 33, 103 (1985)].

As *Escherichia coli* into which a cDNA library constructed by the vector is to be introduced, any strain can be used, as long as the cDNA library can be introduced, expressed and maintained. Examples of such strains include XL1-Blue NRF [*Strategies*, 5, 81 (1992)], C600 [*Genetics*, 39, 440 (1954)], Y1088, Y1090 [*Science*, 222, 778 (1983)], NM522 [*J. Mol. Biol.*, 166, 1 (1983)], K802 [*J. Mol. Biol.*, 16, 118 (1966)] and JM105 [*Gene*, 38, 275 (1985)]. Selection of cDNA clones encoding the V regions of H chain and L chain of the antibody of nonhuman animal from the cDNA library can be carried out by a colony hybridization or plaque hybridization method in which a probe labeled with an isotope or a fluorescence is used [*Molecular Cloning; A Laboratory Manual* (2nd ed.)]. Also, a DNA fragment encoding the V regions of H chain and L chain can be prepared by preparing primers and carrying out the polymerase chain reaction (referred to as "PCR" hereinafter) method [*Molecular Cloning; A Laboratory Manual* (2nd ed.), *Current Protocols in Molecular Biology*, supplements 1 - 34] using cDNA or cDNA library synthesized from poly(A)⁺ RNA or mRNA as the template.

Nucleotide sequence of the DNA can be determined by digesting the cDNA clone selected by the aforementioned method with appropriate restriction enzymes, cloning the digests into a plasmid such as pBluescript SK(-) (manufactured by Stratagene) and then analyzing the resulting clones by a generally used nucleotide sequence analyzing method such as the dideoxy method of Sanger *et al.* [*Proc. Natl. Acad. Sci., U.S.A.*, 74, 5463 (1977)]. Analysis of the nucleotide sequence can be carried out using an automatic nucleotide sequence analyzer such as 373A DNA Sequencer (manufactured by Applied Biosystems).

3. Identification of CDR of the antibody of nonhuman animal

Each V region of H chain and L chain of the antibody forms an antigen binding site. Each of the V regions of H chain and L chain comprises four FRs whose sequences are relatively stable and three CDRs which connect them and are rich in sequence changes (Kabat, E.A. *et al.*, "Sequences of Proteins of Immunological Interest", US Dept. Health and Human Services, 1991). Each CDR can be found by comparing it with the V region amino acid sequences of known antibodies (Kabat, E.A. *et al.*, *Sequences of Proteins of Immunological Interest*, US Dept. Health and Human Services, 1991).

4. Construction of CDR of the antibody of nonhuman animal

The DNA sequences encoding the H chain V region and L chain V region of the human CDR-grafted anti-GM₂ antibody are obtained in the following manner.

First, an amino acid sequence of the V region of each of the H chain and L chain of the human antibody is selected for grafting the CDR of the V region of the anti-GM₂ antibody of nonhuman animal. As the amino acid sequence of the human antibody V region, any of the known V region amino acid sequences derived from human antibodies can be

used. For example, an amino acid sequence selected from human antibody V region amino acid sequences, HMHCS, registered at the Protein Data Bank may be used. However, in order to create a human CDR-grafted antibody having activities of interest such as binding activity and binding specificity for GM₂ or anti-tumor effect on GM₂-positive cells, it is desirable that the sequence has a high homology with the amino acid sequence of the V region of monoclonal antibody derived from nonhuman animal. Next, the DNA sequence encoding the FR in the selected V region amino acid sequence of human antibody is connected with the DNA sequence which encodes the amino acid sequence of the CDR, that becomes the source of the creation, of the V region of monoclonal antibody originated from nonhuman animal, thereby designing a DNA sequence which encodes the amino acid sequence of the V region of each of the H chain and L chain. A total of 6 synthetic DNA fragments are designed for each chain in such a manner that they can cover the thus designed DNA sequence, and PCR is carried out using them. Alternatively, 6 or 7 of each of anti-sense and sense DNA sequences, each comprising 35 to 84 bases, are synthesized in such a manner that they can cover the thus designed DNA sequence, and they are annealed to form double-stranded DNA fragments which are then subjected to the linking reaction. Thereafter, the amplification reaction product or the linking reaction product is subcloned into an appropriate vector and then its nucleotide sequence is determined, thereby obtaining a plasmid which contains the DNA sequence that encodes the amino acid sequence of the V region of each chain of the human CDR-grafted antibody of interest.

5. Modification of amino acid sequence of the V region of human CDR-grafted antibody.

Modification of amino acid sequence of the V region of human CDR-grafted antibody is carried out by a mutation introducing method using PCR. Illustratively, a sense mutation primer and an anti-sense mutation primer, comprising 20 to 40 bases and containing a DNA sequence which encodes amino acid residues after the modification, are synthesized and PCR is carried out using, as the template, a plasmid containing a DNA sequence which encodes the amino acid sequence of the V region to be modified. The amplified fragments are subcloned into an appropriate vector and then their nucleotide sequences are determined to obtain a plasmid which contains a DNA sequence in which the mutation of interest is introduced.

6. Construction of human CDR-grafted antibody expression vector

The human CDR-grafted antibody expression vector can be constructed by inserting the DNA sequences obtained in the above paragraphs 4 and 5, encoding V regions of H chain and L chain of the human CDR-grafted antibody, into upstream of the cDNA, corresponding to the C regions of H chain and L chain of human antibody, of the humanized antibody expression vector prepared in the above paragraph 1. For example, they are inserted into upstream of the cDNA of desired human antibody C regions so that they are properly expressed, by introducing appropriate restriction enzyme recognition sequences into the 5'- and 3'-termini of a synthetic DNA when PCR is carried out in order to construct a DNA sequence which encodes amino acid sequences of the V regions of H chain and L chain of the human CDR-grafted antibody.

7. Expression of the human CDR-grafted antibody and its activity evaluation

A transformant cell line capable of producing the human CDR-grafted antibody can be obtained by introducing the human CDR-grafted antibody expression vector prepared in the above paragraph 6.

Electroporation (JP-A-2-257891; Miyaji, H. *et al.*, *Cytotechnology*, **3**, 133 (1990)) or the like can be used as the introduction method of the expression vector into host cells.

With regard to the host cells into which the human CDR-grafted antibody expression vector is introduced, any type of host cells can be used with the proviso that the human CDR-grafted antibody can be expressed therein. Examples of such cells include mouse SP2/0-Ag14 cells (ATCC CRL1581, referred to as "SP2/0 cells" hereinafter), mouse P3X63-Ag8.653 cells (ATCC CRL1580), dihydrofolate reductase gene (referred to as "DHFR gene" hereinafter)-deficient CHO cells (Urlaub, G. *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, **77**, 4216 (1980)), rat YB2/3HLP2.G11.16Ag.20 cells (ATCC CRL1662, referred to as "YB2/0 cells" hereinafter) and the like.

After introduction of the vector, a transformant cell line capable of producing the human CDR-grafted antibody is selected in accordance with the method disclosed in JP-A-2-257891, using the RPMI 1640 medium containing geneticin (manufactured by Gibco, referred to as "G418" hereinafter) and fetal calf serum (referred to as "FCS" hereinafter). By culturing the thus obtained transformant cell line in a medium, the human CDR-grafted antibody can be produced and accumulated in the culture supernatant. Activity of the human CDR-grafted antibody in the culture supernatant is measured, for example, by the enzyme-linked immunosorbent assay (referred to as "ELISA method" hereinafter; Harlow, E. *et al.*, *Antibodies, A laboratory Manual*, Cold Spring Harbor Laboratory, Chapter 14 (1988)). In addition, production of the human CDR-grafted antibody by the transformant cell line can be improved in accordance

with the method disclosed in JP-A-2-257891 making use of a DHFR gene amplifying system and the like.

The human CDR-grafted antibody can be purified from the aforementioned culture supernatant using a protein A column (Harlow, E. *et al.*, *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory, Chapter 8 (1988)). Alternatively, other purification methods usually used for proteins can be employed. For example, it can be purified by carrying out gel filtration, ion exchange chromatography, ultrafiltration and the like techniques in an appropriate combination. Molecular weight of the H chain, L chain or entire antibody molecule of the thus purified human CDR-grafted antibody is measured for example by polyacrylamide gel electrophoresis (referred to as "SDS-PAGE" hereinafter; Laemmli, U.K. *et al.*, *Nature*, **227**, 680 (1970) or western blot technique (Harlow, E. *et al.*, *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory, Chapter 12 (1988)).

Reactivity of the purified human CDR-grafted antibody with antigens and its binding activity to cultured cancer cell lines are measured by ELISA method, fluorescent antibody technique and the like means. Its complement dependent cytotoxicity (referred to as "CDC" hereinafter) activity and antibody dependent cell mediated cytotoxicity (referred to as "ADCC" hereinafter) activity upon cultured cancer cell lines are measured by the method of Shitara, K. *et al.* (*Cancer Immunol. Immunother.*, **36**, 373 (1993)).

Since the human CDR-grafted antibody of the present invention binds to cultured cancer cell lines of human origin in a specific fashion and shows cytotoxic activities such as CDC activity and ADCC activity, it is useful in the diagnosis and treatment of human cancers and the like. In addition, since most portions of said antibody are originated from the amino acid sequence of a human antibody, when compared with monoclonal antibodies of animal origins excluding human, it is expected that it will exert strong anti-tumor effect without showing immunogenicity and that the effect will be maintained for a prolonged period of time.

The human CDR-grafted antibody of the present invention can be used as an anti-tumor composition, alone or together with at least one pharmaceutically acceptable auxiliary (carrier). For example, the human CDR-grafted antibody is made into an appropriate pharmaceutical composition by dissolving it in physiological saline or an aqueous solution of glucose, lactose, mannitol or the like. Alternatively, the human CDR-grafted antibody is freeze-dried in the usual way and then mixed with sodium chloride to prepare powder injections. As occasion demands, the pharmaceutical composition may contain pharmaceutically acceptable salts and the like additives commonly known in the field of pharmaceutical preparations.

Though the dosage of the pharmaceutical preparation varies depending on the age, symptoms and the like of each patient, the human CDR-grafted antibody is administered to animals including human at a dose of from 0.2 to 20 mg/kg/day. The administration is carried out once a day (single administration or every day administration) or 1 to 3 times a week or once in 2 to 3 weeks, by intravenous injection.

The present invention will be illustrated by the following Examples; however, the present invention is not limited thereto.

EXAMPLE 1

Construction of tandem cassette humanized antibody expression vector, pKANTEX93:

A tandem cassette humanized antibody expression vector, pKANTEX93, for the expression of a human CDR-grafted antibody in animal cells was constructed based on the plasmid pSE1UK1SEd1-3 described in JP-A-2-257891 by inserting a cDNA fragment coding for a human CDR-grafted antibody H chain V region and a cDNA fragment coding for a human CDR-grafted antibody L chain V region into said plasmid upstream of the human antibody C γ 1 cDNA and human antibody C κ cDNA, respectively, in the following manner. The humanized antibody expression vector thus constructed can be also used for expressing a mouse-human chimeric antibody.

1. Modification of *Apal* and *EcoRI* restriction enzyme sites occurring in rabbit β -globin gene splicing and poly A signals

For making it possible to construct a human CDR-grafted antibody expression vector by inserting human CDR-grafted antibody V regions cassette-wise in the form of *NotI*-*Apal* (H chain) and *EcoRI*-*SpI* (L chain) restriction fragments into a vector for humanized antibody expression, the *Apal* and *EcoRI* restriction sites occurring in the rabbit β -globin gene splicing and poly A signals of the plasmid pSE1UK1SEd1-3 were modified in the following manner.

Three μ g of the plasmid pBluescript SK(-) (Stratagene) was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the digestion reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, and the 3' cohesive ends resulting from *Apal* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo), followed by ligation using DNA Ligation Kit (Takara Shuzo). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101. Thus was obtained a plasmid, pBSA, shown in Fig. 1.

Furthermore, 3 μ g of the plasmid pBSA thus obtained was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH

7.5) containing 10 mM magnesium chloride, 100 mM sodium chloride and 1 mM DTT, 10 units of the restriction enzyme *EcoRI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, and the 5' cohesive ends resulting from *EcoRI* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo), followed by ligation using DNA Ligation Kit (Takara Shuzo). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101. Thus was obtained the plasmid pBSAE shown in Fig. 2.

Then, 3 µg of the thus-obtained plasmid pBSAE was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride, 50 mM sodium chloride and 1 mM DTT, 10 units of the restriction enzyme *HindIII* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 20 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, and the solution was divided into two 10-µl portions. To one portion, 10 units of the restriction enzyme *SacII* (Toyobo) was further added and, to the other, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. Both the reaction mixtures were fractionated by agarose gel electrophoresis, whereby about 0.3 µg each of a *HindIII*-*SacII* fragment (about 2.96 kb) and a *KpnI*-*HindIII* fragment (about 2.96 kb) were recovered.

Then, 3 µg of the plasmid pSE1UK1SEd1-3 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SacII* (Toyobo) and 10 units of the restriction enzyme *KpnI* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride, 50 mM sodium chloride and 1 mM DTT, 10 units of the restriction enzyme *HindIII* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.2 µg each of a *HindIII*-*SacII* fragment (about 2.42 kb) and a *KpnI*-*HindIII* fragment (about 1.98 kb) were recovered.

Then, 0.1 µg of the thus-obtained *HindIII*-*SacII* fragment of pSE1UK1SEd1-3 and 0.1 µg of the above *HindIII*-*SacII* fragment of pBSAE were dissolved in a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101 and, as a result, a plasmid, pBSH-S, shown in Fig. 3 was obtained. Furthermore, 0.1 µg of the above-mentioned *KpnI*-*HindIII* fragment of pSE1UK1SEd1-3 and 0.1 µg of the above-mentioned *KpnI*-*HindIII* fragment of pBSAE were dissolved in a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSK-H shown in Fig. 4 was obtained.

Then, 3 µg each of the thus-obtained plasmids pBSH-S and pBSK-H were respectively added to 10-µl portions of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added to each mixture, and the reaction was allowed to proceed at 37°C for 1 hour. Both the reaction mixtures were subjected to ethanol precipitation. With each precipitate, the 3' cohesive ends resulting from *Apal* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo), followed by ligation using DNA Ligation Kit (Takara Shuzo). The thus-obtained recombinant DNA solution were used to transform *Escherichia coli* HB101, and the plasmids pBSH-SA and pBSK-HA shown in Fig. 5 were obtained.

Then, 5 µg each of the thus-obtained plasmids pBSH-SA and pBSK-HA were respectively added to 10-µl portions of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride, 100 mM sodium chloride and 1 mM DTT, 1 unit of the restriction enzyme *EcoRI* (Takara Shuzo) was further added to each mixture, and the reaction was allowed to proceed at 37°C for 10 minutes for partial digestion. Both the reaction mixtures were subjected to ethanol precipitation. With each precipitate, the 5' cohesive ends resulting from *EcoRI* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo), followed by fractionation by agarose gel electrophoresis, whereby about 0.5 µg each of a fragment about 5.38 kb in length and a fragment about 4.94 kb in length were recovered. The thus-recovered fragments (0.1 µg each) were each dissolved in a total of 20 µl of sterilized water and subjected to ligation treatment using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant DNA solutions were respectively used to transform *Escherichia coli* HB101, and the plasmids pBSH-SAE and pBSK-HAE shown in Fig. 6 were obtained.

Then, 3 µg each of the thus-obtained plasmids pBSH-SAE and pBSK-HAE were respectively added to 10-µl portions of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride, 100 mM sodium chloride and 1 mM DTT, 10 units of the restriction enzyme *EcoRI* (Takara Shuzo) was further added to each mixture, and the reaction was allowed to proceed at 37°C for 1 hour. Both the reaction mixtures were subjected to ethanol precipitation. With each precipitate, the 5' cohesive ends resulting from *EcoRI* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo), followed by ligation using DNA Ligation Kit (Takara Shuzo). The thus-obtained recombinant plasmid DNA solutions were each used to transform *Escherichia coli* HB101, and two plasmids, pBSH-SAE and pBSK-HAE, shown in Fig. 7 were obtained. Ten µg each of the thus-obtained plasmids were subjected to sequencing reaction according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by base sequence determination by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech), whereby it was confirmed that both

the *Apal* and *EcoRI* sites had disappeared as a result of the above modification.

(2) *SaI* restriction site introduction downstream from rabbit β -globin gene splicing and poly A signals and SV40 early gene poly A signal

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For making it possible to exchange the antibody H chain and L chain expression promoters of the humanized antibody expression vector each for an arbitrary promoter, a *SaI* restriction site was introduced into the plasmid pSE1UK1SEd1-3 downstream from the rabbit β -globin gene splicing and poly A signals and from the SV40 early gene poly A signal in the following manner.

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Three μ g of the plasmid pBSK-HAEE obtained in Paragraph 1 of Example 1 was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *NaeI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 20 μ l of 50 mM Tris-hydrochloride buffer (pH 9.0) containing 1 mM magnesium chloride, 1 unit of alkaline phosphatase (*E. coli* C75, Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour for dephosphorylation at the 5' termini. The reaction mixture was further subjected to phenol-chloroform extraction and then to ethanol precipitation, and the precipitate was dissolved in 20 μ l of 10 mM Tris-hydrochloride buffer (pH 8.0) containing 1 mM disodium ethylenediaminetetraacetate (hereinafter briefly referred to as "TE buffer"). One μ l of said reaction solution and 0.1 μ g of a phosphorylated *SaI* linker (Takara Shuzo) were added to sterilized water to make a total volume of 20 μ l, followed by ligation treatment using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and a plasmid, pBSK-HAEE*SaI*, shown in Fig. 8 was obtained. Ten μ g of the plasmid thus obtained was subjected to sequencing reaction according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech) for base sequence determination, whereby it was confirmed that one *SaI* restriction site had been introduced downstream from the rabbit β -globin gene splicing and poly A signals and from the SV40 early gene poly A signal.

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3. Modification of *Apal* restriction site occurring in poly A signal of Herpes simplex virus thymidine kinase (hereinafter referred to as "HSVtk") gene

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The *Apal* restriction site occurring in the HSVtk gene poly A signal downstream from the Tn5 kanamycin phosphotransferase gene of the plasmid pSE1UK1SEd1-3 was modified in the following manner.

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Three μ g of the plasmid pBSA obtained in Paragraph 1 of Example 1 was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SacII* (Toyobo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 μ g of a *SacII-XhoI* fragment (about 2.96 kb) was recovered.

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Then, 5 μ g of the plasmid pSE1UK1SEd1-3 was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SacII* (Toyobo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was fractionated by agarose gel electrophoresis, whereby about 1 μ g of a *SacII-XhoI* fragment (about 4.25 kb) was recovered.

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Then, 0.1 μ g of the above *SacII-XhoI* fragment of pBSA and the above *SacII-XhoI* fragment of pSE1UK1SEd1-3 were added to a total of 20 μ l of sterilized water, followed by ligation using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSX-S shown in Fig. 9 was obtained.

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Then, 3 μ g of the thus-obtained plasmid pBSX-S was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the 3' cohesive ends resulting from *Apal* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo) and then ligation was carried out using DNA Ligation Kit (Takara Shuzo). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and a plasmid, pBSX-SA, shown in Fig. 10 was obtained. Ten μ g of the thus-obtained plasmid was subjected to sequencing reaction according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer

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(Pharmacia Biotech) for base sequence determination, whereby it was confirmed that the *Apal* restriction site in the HSVtk gene poly A signal had disappeared.

4. Construction of humanized antibody L chain expression unit

A plasmid, pMohCκ, containing a human antibody Cκ cDNA downstream from the promoter/enhancer of the Moloney mouse leukemia virus long terminal repeat and having a humanized antibody L chain expression unit allowing cassette-wise insertion therein of a humanized antibody L chain V region was constructed in the following manner.

Three μg of the plasmid pBluescript SK(-) (Stratagene) was added to 10 μl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SacI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 μl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *ClaI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, and the cohesive ends resulting from *SacI* and *ClaI* digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo), followed by fractionation by agarose gel electrophoresis, whereby about 1 μg of a DNA fragment about 2.96 kb in length was recovered. A 0.1-μg portion of the DNA fragment recovered was added to a total of 20 μl of sterilized water and subjected to ligation reaction using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSSC shown in Fig. 11 was obtained.

Then, 3 μg of the thus-obtained plasmid pBSSC was added to 10 μl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 μl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 μg of a *KpnI*-*XhoI* fragment (about 2.96 kb) was recovered.

Then, 5 μg of the plasmid pAGE147 described in JP-A-6-205694 was added to 10 μl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 μl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was fractionated by agarose gel electrophoresis, whereby about 0.3 μg of a *KpnI*-*XhoI* fragment (about 0.66 kb) containing the Moloney mouse leukemia virus long terminal repeat promoter/enhancer was recovered.

Then, 0.1 μg of the *KpnI*-*XhoI* fragment of pBSSC and 0.1 μg of the *KpnI*-*XhoI* fragment of pAGE147 each obtained as mentioned above were dissolved in a total of 20 μl of sterilized water and subjected to ligation using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSMo shown in Fig. 12 was obtained.

Then, 3 μg of the above plasmid pBSMo was added to 10 μl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 μl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *HindIII* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 μg of a *KpnI*-*HindIII* fragment (about 3.62 kb) was recovered.

Then, synthetic DNAs respectively having the base sequences shown in SEQ ID NO:12 and SEQ ID NO:13 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). To 15 μl of sterilized water were added 0.3 μg each of the thus-obtained synthetic DNAs, and the mixture was heated at 65°C for 5 minutes. The reaction mixture was allowed to stand at room temperature for 30 minutes and then 2 μl of 10-fold concentrated buffer [500 mM Tris-hydrochloride (pH 7.6), 100 mM magnesium chloride, 50 mM DTT] and 2 μl of 10 mM ATP were added, 10 units of T4 polynucleotide kinase was further added, and the reaction was allowed to proceed at 37°C for 30 minutes for phosphorylation of the 5' termini. To a total of 20 μl of sterilized water were added 0.1 μg of the above *KpnI*-*HindIII* fragment (3.66 kb) derived from the plasmid pBSMo and 0.05 μg of the phosphorylated synthetic DNA pair, and ligation was effected using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSMoS shown in Fig. 13 was obtained. Ten μg of the plasmid thus obtained was subjected to sequencing reaction according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Bio-

tech) for base sequence determination, whereby it was confirmed that the synthetic DNA pair had been introduced as desired.

Then, 3 µg of the plasmid pChilgLA1 described in JP-A-5-304989 was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *EcoRV* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of an *EcoRI-EcoRV* fragment (about 9.70 kb) was recovered.

Then, synthetic DNAs respectively having the base sequences shown in SEQ ID NO:14 and SEQ ID NO:15 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). To 15 µl of sterilized water were added 0.3 µg each of the thus-obtained synthetic DNAs, and the mixture was heated at 65°C for 5 minutes. The reaction mixture was allowed to stand at room temperature for 30 minutes. Then, 2 µl of 10-fold concentrated buffer [500 mM Tris-hydrochloride (pH 7.6), 100 mM magnesium chloride, 50 mM DTT] and 2 µl of 10 mM ATP were added, 10 units of T4 polynucleotide kinase was further added, and the reaction was allowed to proceed at 37°C for 30 minutes for phosphorylation of the 5' termini. To a total of 20 µl of sterilized water were added 0.1 µg of the above *EcoRI-EcoRV* fragment (9.70 kb) derived from the plasmid pChilgLA1 and 0.05 µg of the phosphorylated synthetic DNA, and ligation was effected using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pChilgLA1S shown in Fig. 14 was obtained.

Then, 3 µg of the plasmid pBSMoS obtained in the above manner was dissolved in 10 µl of 20 mM Tris-hydrochloride buffer (pH 8.5) containing 100 mM potassium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *HpaI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *EcoRI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of an *HpaI-EcoRI* fragment (about 3.66 kb) was recovered.

Then, 10 µg of the plasmid pChilgLA1S obtained as mentioned above was dissolved in 10 µl of 20 mM Tris-acetate buffer (pH 7.9) containing 50 mM potassium acetate, 10 mM magnesium acetate, 1 mM DTT and 100 µg/ml BSA, 10 units of the restriction enzyme *NlaIV* (New England Biolabs) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *EcoRI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.3 µg of an *NlaIV-EcoRI* fragment (about 0.41 kb) was recovered.

Then, 0.1 µg of the above *HpaI-EcoRI* fragment of pBSMoS and 0.1 µg of the above *NlaIV-EcoRI* fragment of pChilgLA1S were added to a total of 20 µl of sterilized water, and ligation was effected using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pMohCk shown in Fig. 15 was obtained.

5. Construction of humanized antibody H chain expression unit

A plasmid, pMohCγ1, containing a human antibody Cγ1 cDNA downstream from the promoter/enhancer of the Moloney mouse leukemia virus long terminal repeat and having a humanized antibody H chain expression unit allowing cassette-wise insertion therein of a humanized antibody H chain V region was constructed in the following manner.

Three µg of the plasmid pBSMo obtained in Paragraph 4 of Example 1 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 30 mM sodium acetate buffer (pH 5.0) containing 100 mM sodium chloride, 1 mM zinc acetate and 10% glycerol, 10 units of Mung bean nuclease (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 10 minutes. The reaction mixture was subjected to phenol-chloroform extraction and then to ethanol precipitation, the cohesive ends of the precipitate were rendered blunt using DNA Blunting Kit (Takara Shuzo) and ligation was effected using DNA Ligation Kit (Takara Shuzo). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSMoSal shown in Fig. 16 was obtained. A 10-µg portion of the plasmid obtained was subjected to sequencing reaction according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech) for base sequence determination, whereby it was confirmed that the *XhoI* restriction site upstream of the Moloney mouse leukemia virus long terminal repeat promoter/enhancer had disappeared.

Then, 3 µg of the plasmid pBSMoSal obtained as mentioned above was added to 10 µl of 10 mM Tris-hydrochloride

buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *HindIII* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of a *KpnI-HindIII* fragment (about 3.66 kb) was recovered.

Then, synthetic DNAs respectively having the base sequences shown in SEQ ID NO:16 and SEQ ID NO:17 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). To 15 µl of sterilized water were added 0.3 µg each of the thus-obtained synthetic DNAs, and the mixture was heated at 65°C for 5 minutes. The reaction mixture was allowed to stand at room temperature for 30 minutes. Then, 2 µl of 10-fold concentrated buffer [500 mM Tris-hydrochloride (pH 7.6), 100 mM magnesium chloride, 50 mM DTT] and 2 µl of 10 mM ATP were added, 10 units of T4 polynucleotide kinase was further added, and the reaction was allowed to proceed at 37°C for 30 minutes for phosphorylation of the 5' termini. To a total of 20 µl of sterilized water were added 0.1 µg of the above *KpnI-HindIII* fragment (3.66 kb) derived from the plasmid pBSMoSa1 and 0.05 µg of the phosphorylated synthetic DNA, and ligation was effected using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSMoSa1S shown in Fig. 17 was obtained. A 10-µg portion of the thus-obtained plasmid was subjected to sequencing reaction according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech), for base sequence determination whereby it was confirmed that the synthetic DNA had been introduced as desired.

Then, 10 µg of the plasmid pChilgHB2 described in JP-A-5-304989 was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Eco52I* (Toyobo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 30 mM sodium acetate buffer (pH 5.0) containing 100 mM sodium chloride, 1 mM zinc acetate and 10% glycerol, 10 units of Mung bean nuclease (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 10 minutes. The reaction mixture was subjected to phenol-chloroform extraction and then to ethanol precipitation, and the cohesive ends were rendered blunt using DNA Blunting Kit (Takara Shuzo). After ethanol precipitation, the precipitate was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.7 µg of *Apal*-blunt end fragment (about 0.99 kb) was recovered.

Then, 3 µg of the plasmid pBluescript SK(-) (Stratagene) was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (2U Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 33 mM Tris-acetate buffer (pH 7.9) containing 10 mM magnesium acetate, 66 mM potassium acetate, 0.5 mM DTT and 100 µg/ml BSA, 10 units of the restriction enzyme *SmaI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of an *Apal-SmaI* fragment (about 3.0 kb) was recovered.

Then, 0.1 µg of the *Apal*-blunt end fragment of pChilgHB2 and 0.1 µg of the *Apal-SmaI* fragment of pBluescript SK(-), each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSHCy1 shown in Fig. 18 was obtained.

Then, 5 µg of the above plasmid pBSHCy1 was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SpeI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of an *Apal-SpeI* fragment (about 1.0 kb) was recovered.

Then, 3 µg of the plasmid pBSMoSa1S obtained as mentioned above was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SpeI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture

was fractionated by agarose gel electrophoresis, whereby about 1 µg of an *Apal-SpeI* fragment (about 3.66 kb) was recovered.

Then, 0.1 µg of the *Apal-SpeI* fragment of pBSHCy1 and 0.1 µg of the *Apal-SpeI* fragment of pBSMoSa1S, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pMohCy1 shown in Fig. 19 was obtained.

6. Construction of tandem cassette humanized antibody expression vector, pKANTEX93

A tandem cassette humanized antibody expression vector, pKANTEX93, was constructed using the various plasmids obtained in Paragraphs 1 through 5 of Example 1 in the following manner.

Three µg of the plasmid pBSH-SAE obtained in Paragraph 1 of Example 1 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *HindIII* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Sall* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of a *HindIII-Sall* fragment (about 5.42 kb) was recovered.

Then, 5 µg of the plasmid pBSK-HAEE obtained in Paragraph 1 of Example 1 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *HindIII* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.8 µg of a *KpnI-HindIII* fragment (about 1.98 kb) containing the rabbit β-globin gene splicing and poly A signals, the SV40 early gene poly A signal and the SV40 early gene promoter was recovered.

Then, 5 µg of the plasmid pMohCy1 obtained in Paragraph 5 of Example 1 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Sall* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.8 µg of a human CDR-grafted antibody H chain expression unit-containing *KpnI-Sall* fragment (about 1.66 kb) was recovered.

Then, 0.1 µg of the *HindIII-Sall* fragment of pBSH-SAE, 0.1 µg of the *KpnI-HindIII* fragment of pBSK-HAEE and 0.1 µg of the *KpnI-Sall* fragment of pMohCy1, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated together using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pMoy1SP shown in Fig. 20 was obtained.

Then, 3 µg of the above plasmid pMoy1SP was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Sall* (Takara Shuzo) and 10 units of the restriction enzyme *XhoI* were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of a *Sall-XhoI* fragment (about 9.06 kb) was recovered.

Then, 5 µg of the plasmid pBSK-HAEEsal obtained in Paragraph 2 of Example 1 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Sall* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.7 µg of a *KpnI-Sall* fragment (about 1.37 kb) containing the rabbit β-globin gene splicing and poly A signals and the SV40 early gene poly A signal was recovered.

Then, 5 µg of the plasmid pMohCx obtained in Paragraph 4 of Example 1 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5)

containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 0.7 µg of a human CDR-grafted antibody L chain expression unit-containing *KpnI-XhoI* fragment (about 1.06 kb) was recovered.

Then, 0.1 µg of the *Sall-XhoI* fragment of pMoy1SP, 0.1 µg of the *KpnI-Sall* fragment of pBSK-HAEEsal and 0.1 µg of the *KpnI-XhoI* fragment of pMohCk, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated together using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plaid pMoy1SP shown in Fig. 21 was obtained.

Then, 3 µg of the above plasmid pMoy1SP was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 1 unit of the restriction enzyme *SacII* (Toyobo) was further added, and the reaction was allowed to proceed at 37°C for 10 minutes for partial digestion. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.2 µg of a *SacII-XhoI* fragment (about 8.49 kb) was recovered.

Then, 3 µg of the plaid pBSX-SA obtained in Paragraph 3 of Example 1 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SacII* (Toyobo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *XhoI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of a *SacII-XhoI* fragment (about 4.25 kb) was recovered.

Then, 0.1 µg of the *SacII-XhoI* fragment of pMoy1SP and 0.1 µg of the *SacII-XhoI* fragment of pBSX-SA, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pKANTE93 shown in Fig. 22 was obtained.

EXAMPLE 2

1. Expression of mouse-human chimeric anti-GM₂ antibody

Mouse-human chimeric anti-GM₂ antibody expression was effected using the humanized antibody expression vector pKANTE93 mentioned above in Example 1 in the following manner.

(1) Construction of plasmid pBSH3 containing mouse anti-GM₂ antibody KM796 H chain V region cDNA

Three µg of the plasmid pBluescript SK(-) (Stratagene) was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units each of the restriction enzymes *SacII* (Toyobo) and *KpnI* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, and the precipitate was subjected to blunting treatment for rendering blunt the 3' cohesive ends resulting from the restriction enzyme digestion using DNA Blunting Kit (Takara Shuzo). The resulting reaction was precipitated with ethanol, the precipitate thus obtained was dissolved in 20 µl of a buffer containing 50 mM Tris-hydrochloride buffer (pH 9.0) and 1 mM magnesium chloride, and the mixture thus obtained was allowed to react by adding one unit of alkali phosphatase (*E. coli* C75, Takara Shuzo) at 37°C for 1 hour for dephosphorylation of the 5' termini. Then, fractionation by agarose gel electrophoresis was carried out, and about 1 µg of a DNA fragment about 2.95 kb in size was recovered.

Then, synthetic DNAs respectively having the base sequences shown in SEQ ID NO:18 and SEQ ID NO:19 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). To 15 µl of sterilized water were added 0.3 µg each of the synthetic DNAs obtained, and the mixture was heated at 65°C for 5 minutes. The reaction mixture was allowed to stand at room temperature for 30 minutes and then 2 µl of 10-fold concentrated buffer [500 mM Tris-hydrochloride (pH 7.6), 100 mM magnesium chloride, 50 mM DTT] and 2 µl of 10 mM ATP were added, 10 units of T4 polynucleotide kinase was further added, and the reaction was allowed to proceed at 37°C for 30 minutes for phosphorylating the 5' termini. To a total of 20 µl of sterilized water were added 0.1 µg of the DNA fragment (2.95 kb) derived from the plasmid pBluescript SK(-) and 0.05 µg of the phosphorylated synthetic DNA, each obtained as mentioned above, followed by ligation to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plaid pBSNA shown in Fig.

23 was obtained. Ten µg of the plasmid obtained was subjected to sequencing reaction treatment according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech) for base sequence determination, whereby it was confirmed that the synthetic DNA had been introduced as desired.

5 Then, 3 µg of the plasmid pBSNA obtained as mentioned above was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 µg/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *NotI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of a DNA fragment about 2.95 kb in size was recovered.

10 Then, 10 µg of the plasmid pChi796HM1 described in JP-A-6-205964 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 µg/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *NotI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 µg of a DNA fragment about 0.45 kb in size was recovered.

20 Then, 0.1 µg of the *Apal-NotI* fragment of pBSNA and 0.1 µg of the *Apal-NotI* fragment of pChi796HM1, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSH3 shown in Fig. 24 was obtained.

25 (2) Construction of plasmid pBSL3 containing mouse anti-GM₂ antibody KM796 L chain V region cDNA

Three µg of the plasmid pBluescript SK(-) (Stratagene) was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *KpnI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, and the precipitate was subjected to blunting treatment for rendering blunt the 3' cohesive ends resulting from *KpnI* digestion using DNA Blunting Kit (Takara Shuzo) and then to ethanol precipitation, the precipitate was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *SacI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of a DNA fragment about 2.95 kb in size was recovered.

40 Then, synthetic DNAs respectively having the base sequences shown in SEQ ID NO:20 and SEQ ID NO:21 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). To 15 µl of sterilized water were added 0.3 µg each of the synthetic DNAs obtained, and the mixture was heated at 65°C for 5 minutes. The reaction mixture was allowed to stand at room temperature for 30 minutes. Then, 2 µl of 10-fold concentrated buffer [500 mM Tris-hydrochloride (pH 7.5), 100 mM magnesium chloride, 50 mM DTT] and 2 µl of 10 mM ATP were added, 10 units of T4 polynucleotide kinase was further added, and the reaction was allowed to proceed at 37°C for 30 minutes for phosphorylating the 5' termini. The, 0.1 µg of the DNA fragment (2.95 kb) derived from the plasmid pBluescript SK(-) and 0.05 µg of the phosphorylated synthetic DNA, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSES shown in Fig. 25 was obtained. Ten µg of the plasmid obtained was subjected to sequencing reaction treatment according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech) for base sequence determination, whereby it was confirmed that the synthetic DNA had been introduced as desired.

50 Then, 3 µg of the plasmid pBSES obtained as mentioned above was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *SpI* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of a DNA fragment about 2.95 kb in size was recovered.

55 Then, 5 µg of the plasmid pKM796L1 described in JP-A-6-205964 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *AflIII* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1

hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 µg of an *EcoRI*-*A*/III fragment about 0.39 kb in size was recovered. Then, synthetic DNAs respectively having the base sequences shown in SEQ ID NO:22 and SEQ ID NO:23 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). To 15 µl of sterilized water were added 0.3 µg each of the synthetic DNAs obtained, and the mixture was heated at 65°C for 5 minutes. The reaction mixture was allowed to stand at room temperature for 30 minutes. Then, 2 µl of 10-fold concentrated buffer [500 mM Tris-hydrochloride (pH 7.6), 100 mM magnesium chloride, 50 mM DTT] and 2 µl of 10 mM ATP were added, 10 units of T4 polynucleotide kinase was further added, and the reaction was allowed to proceed at 37°C for 30 minutes for phosphorylating the 5' termini.

Then, 0.1 µg of the pBSES-derived *EcoRI*-*Sp*II fragment (2.95 kb), 0.1 µg of the pKM796LI-derived *EcoRI*-*A*/III fragment and 0.05 µg of the phosphorylated synthetic DNA, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated together using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSL3 shown in Fig. 26 was obtained. Ten µg of the plasmid obtained was subjected to sequencing reaction treatment according to the instructions attached to AutoRead Sequencing Kit (Pharmacia Biotech), followed by electrophoresis on A.L.F. DNA Sequencer (Pharmacia Biotech) for base sequence determination, whereby it was confirmed that the synthetic DNA had been introduced as desired.

3. Construction of mouse-human chimeric anti-GM₂ antibody expression vector, pKANTEX796

A mouse-human chimeric anti-GM₂ antibody expression vector, pKANTEX796, was constructed using the plasmid pKANTEX93 obtained in Example 1 and the plasmids pBSH3 and pBSL3 respectively obtained in Paragraph 1 (1) and (2) of Example 2, in the following manner.

Three µg of the plasmid pBSH3 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Ap*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 µg/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *Not*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 µg of an *Ap*I-*Not*I fragment about 0.46 kb in size was recovered.

Then, 3 µg of the plasmid pKANTEX93 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Ap*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 µg/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *Not*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, whereby about 1 µg of an *Ap*I-*Not*I fragment about 12.75 kb in size was recovered.

Then, 0.1 µg of the pBSH3-derived *Ap*I-*Not*I fragment and 0.1 µg of the pKANTEX93-derived *Ap*I-*Not*I fragment, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pKANTEX796H shown in Fig. 27 was obtained.

Then, 3 µg of the plasmid pBSL3 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 µg of an *EcoRI*-*Sp*II fragment about 0.4 kb in size was recovered.

Then, 3 µg of the plasmid pKANTEX796H was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *EcoRI*-*Sp*II fragment about 13.20 kb in size was recovered.

Then, 0.1 µg of the pBSL3-derived *EcoRI*-*Sp*II fragment and 0.1 µg of the pKANTEX796H-derived *EcoRI*-*Sp*II fragment, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pKANTEX796 shown in Fig. 28 was obtained.

(4) Expression of mouse-human chimeric anti-GM₂ antibody in YB2/0 cells using pKANTEX796

Introduction of the plasmid into YB2/0 cells (ATCC CRL1662) was carried out by the electroporation method (Miyaji, H. *et al.*, *Cytotechnology*, 3, 133 (1990)). A 4 µg portion of pKANTEX796 obtained in Paragraph 1 (3) of Example 2 was introduced into 4×10^6 cells of YB2/0 cells, and the resulting cells were suspended in 40 ml of RPMI1640-FCS (10) medium [RPMI1640 medium (manufactured by Nissui Pharmaceutical) supplemented with 10% of FCS, an appropriate amount of 7.5% sodium bicarbonate solution, 3% of 200 mM L-glutamine solution (manufactured by Gibco) and 0.5% of penicillin-streptomycin solution (manufactured by Gibco, contains 5,000 U/ml of penicillin and 5 mg/ml of streptomycin)] and dispensed in 200 µl portions into wells of a 96 well microplate. After 24 hours of culturing at 37°C in a 5% CO₂ incubator, G418 was added to each well to a final concentration of 0.5 mg/ml, and the cells were cultured for 1 to 2 weeks. Culture supernatants were recovered from wells in which colonies of transformant cell lines have been formed, and the activity of the mouse-human chimeric anti-GM₂ antibody in the culture supernatants was measured by the ELISA method described in the following paragraph (5). Cells in wells in which the activity was found were subjected to gene amplification in the following manner with an attempt to increase expression quantity of the chimera antibody. Firstly, the cells were suspended in the RPMI1640-FCS (10) medium supplemented with 0.5 mg/ml of G418 and 50 nM of methotrexate (manufactured by Sigma, to be referred to as "MTX" hereinafter), to a density of $1-2 \times 10^5$ cells/ml, and the suspension was dispensed in 2 ml portions in wells of a 24 well plate. The cells were cultured at 37°C for 1 to 2 weeks in a 5% CO₂ incubator to induce resistant cells to 50 nM MTX. In wells in which the cells resistant to 50 nM MTX have been formed, the final concentration of MTX was increased to 100 nM and then to 200 nM and the expression quantity was evaluated by the ELISA method to select cells having the highest expression quantity. The thus selected cells were subjected twice to cloning by the limiting dilution analysis and then established as the final chimera antibody stable expression cells. The thus established mouse-human chimeric anti-GM₂ antibody stable expression cells showed an expression quantity of about 1 to 2 µg/ml, so that it was confirmed that efficient and stable expression of the humanized antibody can be effected by the use of pKANTEX93.

(5) ELISA method

A 2 ng portion of ganglioside was dissolved in 2 ml of ethanol solution containing 5 ng of phosphatidylcholine (manufactured by Sigma) and 2.5 ng of cholesterol (manufactured by Sigma). This solution or a diluted solution thereof was dispensed in 20 µl portions in wells of a 96 well microplate (manufactured by Greiner), air-dried and then subjected to blowing with a phosphate buffer containing 1% BSA (to be referred to as "PBS" hereinafter). To the resulting plate was added culture supernatant of a transformant cell line, a purified mouse monoclonal antibody a purified mouse-human chimeric antibody or a purified humanized antibody in an amount of from 50 to 100 µl, subsequently carrying out 1 to 2 hours of reaction at room temperature. After the reaction and subsequent washing of each well with PBS, 50 to 100 µl of a peroxidase-labeled rabbit anti-mouse IgG antibody (manufactured by Dako, used by 400 times dilution) or a peroxidase-labeled goat anti-human γ chain antibody (manufactured by Kiyukegard & Perry Laboratory, used by 1,000 times dilution) was added thereto, and 1 to 2 hours of reaction was carried out at room temperature. After washing with PBS, 50 to 100 µl of an ABTS substrate solution [a solution prepared by dissolving 550 mg of 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) in 1 liter of 0.1 M citrate buffer (pH 4.2) and adding 1 µl/ml of hydrogen peroxide to the solution just before its use] was added to each well to effect development of color which was then measured at OD₄₁₅.

2. Transient mouse-human chimeric antibody expression in COS-7 (ATCC CRL 1651) cells

For enabling more rapid activity evaluation of various versions of human CDR-grafted anti-GM₂ antibody, transient expression of mouse-human chimeric anti-GM₂ antibody expression was caused in COS-7 cells by the Lipofectamine method using pKANTEX796 and a variant thereof in the following manner.

(1) Construction of variant of pKANTEX796

Since transient antibody expression in animal cells is dependent on the copy number of an expression vector introduced, it was supposed that an expression vector smaller in size would show a higher expression efficiency. Therefore, a smaller humanized antibody expression vector, pT796, was constructed by deleting a region supposedly having no effect on humanized antibody expression from pKANTEX796 in the following manner.

Thus, 3 µg of the plasmid pKANTEX796 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Hind*III (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Mlu*I

(Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, and the 5' cohesive ends resulting from the restriction enzyme digestion were rendered blunt using DNA Blunting Kit (Takara Shuzo). The reaction mixture was fractionated by agarose gel electrophoresis and about 1 µg of a DNA fragment about 9.60 kb in size was recovered. A 0.1-µg portion of the thus-recovered DNA fragment was added to a total of 20 µl of sterilized water and subjected to ligation treatment using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pT796 shown in Fig. 29 was obtained.

(2) Transient expression of mouse-human chimeric anti-GM₂ antibody using pKANTEX796 and pT796

A 1×10^5 cells/ml suspension of COS-7 cells was distributed in 2-ml portions into wells of a 6-well plate (Falcon) and cultured overnight at 37°C. Two µg of pKANTEX796 or pT796 was added to 100 µl of OPTI-MEM medium (Gibco), a solution prepared by adding 10 µl of LIPOFECTAMINE reagent (Gibco) to 100 µl of OPTI-MEM medium (Gibco) was further added, and the reaction was allowed to proceed at room temperature for 40 minutes to cause DNA-liposome complex formation. The COS-7 cells cultured overnight were washed twice with 2 ml of OPTI-MEM medium (Gibco), the complex-containing solution was added, and the cells were cultured at 37°C for 7 hours. Then, the solution was removed, 2 ml of DMEM medium (Gibco) containing 10% FCS was added to each well, and the cells were cultured at 37°C. After 24 hours, 48 hours, 72 hours, 96 hours and 120 hours of cultivation, the culture supernatant was recovered and, after concentration procedure as necessary, evaluated for mouse-human chimeric anti-GM₂ antibody activity in the culture supernatant by the ELISA method described in Paragraph 1 (5) of Example 2. The results are shown in Fig. 30. As shown in Fig. 30, higher levels of transient mouse-human chimeric anti-GM₂ antibody expression was observed with pT796 as compared with pKANTEX796. For pT796, the level of expression was highest at 72 to 96 hours, the concentration being about 30 ng/ml (in terms of GM₂ binding activity). The above results indicate that construction of a pKANTEX93-derived vector having a reduced size and introduction thereof into COS-7 cells make it possible to make activity evaluation of expression vector-derived humanized antibodies in a transient expression system. Furthermore, for close activity comparison of various versions of human CDR-grafted anti-GM₂ antibody as mentioned hereinafter, the ELISA method described below under (3) was used to determine antibody concentrations in transient expression culture supernatants.

(3) Determination by sandwich ELISA of humanized antibody concentrations in various culture supernatants

A solution prepared by 400-fold dilution of goat anti-human γ chain antibody (Igaku Seibutugaku Kenkyusho) with PBS was distributed in 50-µl portions into wells of a 96-well microtiter plate and allowed to stand overnight at 4°C for binding to the wells. After removing the antibody solution, blowing was effected with 100 µl of PBS containing 1% BSA at 37°C for 1 hour. Fifty µl of a transient expression culture supernatant or purified mouse-human chimeric anti-GM₂ antibody was added thereto and allowed to react at room temperature for 1 hour. Thereafter, the solution was removed, the wells were washed with PBS, and 50 µl of a solution prepared by 500-fold dilution of peroxidase-labeled mouse anti-human κ chain antibody (Zymet) with PBS was added and allowed to react at room temperature for 1 hour. After washing with PBS, 50 µl of an ABTS substrate solution was added for causing color development, and the OD₄₁₅ was measured.

EXAMPLE 3

Production of human CDR-grafted anti-GM₂ antibody I

A human CDR-grafted anti-GM₂ antibody higher in GM₂-binding activity than the human CDR-grafted anti-GM₂ antibody described in Example 2 of JP-A-6-105694 was produced in the following manner.

(1) Modification of human CDR-grafted anti-GM₂ antibody H chain V region described in Paragraph 1 (1) of Example 2 of JP-A-6-205694

DNAs coding for some versions of the human CDR-grafted anti-GM₂ antibody H chain V region described in Example 2 as derived by replacing several amino acids in the FR with original mouse antibody amino acids were constructed in the following manner. Based on a computer model for the V region of mouse antibody KM796, those amino acid residues that were expected to contribute to restoration of antigen-binding activity as a result of replacement were selected as the amino acid residues to be replacement. First, DNAs respectively having the base sequences of SEQ ID NO:24 and SEQ ID NO:25 were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A).

Then, a version (version 2) of human CDR-grafted antibody H chain V region shown in SEQ ID NO:26 and having

replacement in positions 78 (threonine in lieu of glutamine), 79 (alanine in lieu of phenylalanine) and 80 (tyrosine in lieu of serine) was constructed in the same manner as in Paragraph 1 (1) of Example 2 of JP-A-6-205964 using a synthetic DNA of SEQ ID NO:24 in lieu of the synthetic DNA of SEQ ID NO:27 of JP-A-6-205964.

Then, another version (version 4) of human CDR-grafted antibody H chain V region shown in SEQ ID NO:27 and having replacements in positions 27 (tyrosine in lieu of phenylalanine), 30 (threonine in lieu of serine), 40 (serine in lieu of proline) and 41 (histidine in lieu of proline) was constructed in the same manner as in Paragraph 1 (1) of Example 2 of JP-A-6-205964 using a synthetic DNA of SEQ ID NO:25 in lieu of the synthetic DNA of SEQ ID NO:25 of JP-A-6-205964.

(2) Construction of human CDR-grafted anti-GM₂ antibody H chain V region using known HMHCS of human antibody H chain V region

According to Kabat *et al.* (Kabat E. A. *et al.*, *Sequences of Proteins of Immunological Interest*, US Dept. of Health and Human Services, 1991), known human antibody H chain V regions are classifiable into subgroups I to III (Human Sub Groups (HSG) I to III) based on the homology of their FR regions, and coon sequences have been identified for respective subgroups. The present inventors identified HMHCS as one meaning from the common sequences, a human CDR-grafted anti-GM₂ antibody H chain V region was constructed based on the HMHCS. First, for selecting HMHCS to serve as the base, the homology was examined between the FR, of the mouse antibody KM796 H chain V region and the FR of the HMHCS of the human antibody H chain V region of each subgroup (Table 1).

TABLE 1

Homology (%) between mouse antibody KM796 H chain V region FR and human antibody H chain V region common sequence FR		
HSG I	HSG II	HSG III
72.1	52.9	58.6

As a result, it was confirmed that subgroup I shows the greatest similarity. Thus, based on the HMHCS of subgroup I, a human CDR-grafted anti-GM₂ antibody H chain V region was constructed by the PCR method in the following manner.

Synthetic DNAs respectively having the base sequences of SEQ ID NO:28 through SEQ ID NO:33 were synthesized using an automatic DNA synthesizer (Applied Systems model 380A). The DNAs synthesized were added, each to a final concentration of 0.1 μ M, to 50 μ l of 10 mM Tris-hydrochloride buffer (pH 8.3) containing 50 mM potassium chloride, 1.5 mM magnesium chloride, 0.001% gelatin, 200 μ M dNTP, 0.5 μ M M13 primer RV (Takara Shuzo), 0.5 μ M M13 primer M4 (Takara Shuzo) and 2 units of TaKaRa Taq DNA polymerase, the mixture was covered with 50 μ l of mineral oil, a DNA thermal cycler (Perkin Elmer model PJ480) was loaded with the mixture, and 30 PCR cycles (2 minutes at 94°C, 2 minutes at 55°C and 2 minutes at 72°C per cycle) were conducted. The reaction mixture was purified using QIAquick PCR Purification Kit (Qiagen) and then made into a solution in 30 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 μ l of 50 mM Tris-hydrochloride (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 μ g/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *NotI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.2 μ g of an *Apal-NotI* fragment about 0.44 kb in size was recovered.

Then, 3 μ g of the plasmid pBSH3 obtained in Paragraph 1 (1) of Example 2 was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 μ g/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *NotI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 μ g of an *Apal-NotI* fragment about 2.95 kb in size was recovered.

Then, 0.1 μ g of the *Apal-NotI* fragment of the human CDR-grafted anti-GM₂ antibody H chain V region and 0.1 μ g of the *Apal-NotI* fragment of pBSH3, each obtained as mentioned above, were added to a total of 20 μ l of sterilized

water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101. Plasmid DNAs were prepared from 10 transformant clones and their base sequences were determined. As a result, a plasmid, pBSB10, shown in Fig. 31 and having the desired base sequence was obtained. The amino acid sequence and base sequence of the human CDR-grafted anti-GM₂ antibody H chain V region contained in pBSH10 are shown in SEQ ID NO:7. In the amino acid sequence of the thus-constructed human CDR-grafted anti-GM₂ antibody H chain V region, arginine in position 67, alanine in position 72, serine in position 84 and arginine in position 98 in the FR as selected based on a computer model for the V region are replaced by lysine, valine, histidine and threonine, respectively, that are found in the mouse antibody KM796 H chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

(3) Modification of human CDR-grafted anti-GM₂ antibody L chain V region described in Paragraph 1 (2) of Example 2 of JP-A-6-205694

First, a DNA having the base sequence of SEQ ID NO:34 was synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A), and a human CDR-grafted anti-GM₂ antibody L chain V region cDNA with a 3' terminus capable of pairing with the restriction enzyme *Sp*I was constructed by following the same reaction procedure as in Paragraph 1 (2) of Example 2 of JP-A-6-205694 using the synthetic DNA in lieu of the synthetic DNA of SEQ ID NO:35 of JP-A-6-205964.

Then, 3 µg of the plasmid pBSL3 obtained in Paragraph 1 (2) of Example 2 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *Eco*RI (Takara Shuzo) and *Sp*I (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *Eco*RI-*Sp*I fragment about 2.95 kb in size was recovered.

Then, 0.1 µg of the *Eco*RI-*Sp*I fragment of the human CDR-grafted anti-GM₂ antibody L chain V region obtained as mentioned above and 0.1 µg of the above *Eco*RI-*Sp*I fragment of pBSL3 were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSL16 shown in Fig. 32 was obtained.

Then, DNAs coding for certain versions of the human CDR-grafted anti-GM₂ antibody L chain V region contained in the above plasmid pBSL16 were constructed by replacing a certain number of amino acids in the FR with original mouse antibody amino acids by mutagenesis by means of PCR in the following manner (Fig. 33). Based on a computer model for the V region of mouse antibody KM796, those amino acid residues that were expected to contribute to restoration of antigen-binding activity as a result of replacement were selected as the amino acid residues to be replaced.

Antisense and sense DNA primers for introducing mutations were synthesized using an automatic DNA synthesizer (Applied Biosystems model 380A). A first PCR reaction was conducted in the same manner as in Paragraph 1 (2) of Example 3 using a final concentration each of 0.5 µM of M13 primer RV (Takara Shuzo) and the antisense DNA primer and of M13 primer M4 (Takara Shuzo) and the sense DNA primer, with 1 ng of pBSL16 as the template. Each reaction mixture was purified using QIAquick PCR Purification Kit (Qiagen) with elution with 20 µl of 10 mM Tris-hydrochloride (pH 8.0). Using 5 µl of each elute, a second PCR reaction was conducted in the same manner as in Paragraph 1 (2) of Example 3. The reaction mixture was purified using QIAquick PCR Purification Kit (Qiagen) and then made into a solution in 30 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *Eco*RI (Takara Shuzo) and *Sp*I (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.2 µg of an *Eco*RI-*Sp*I fragment (about 0.39 kb) of each replacement version of the human CDR-grafted anti-GM₂ antibody L chain V region was recovered.

Then, 0.1 µg of the above *Eco*RI-*Sp*I fragment of each replacement version of the human CDR-grafted anti-GM₂ antibody L chain V region and 0.1 µg of the *Eco*RI-*Sp*I fragment of pBSL3 were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and a plasmid DNA was prepared from a transformant clone, and the base sequence of said plasmid was determined. In this way, plasmids respectively containing a base sequence having a desired mutation or mutations were obtained.

Thus, a plasmid, pBSLV1, containing version 1, shown in SEQ ID NO:37, of the human CDR-grafted anti-GM₂ antibody L chain V region was obtained following the above procedure using the synthetic DNA of SEQ ID NO:35 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:36 as the mutant sense primer. In the amino acid sequence of the version 1 human CDR-grafted anti-GM₂ antibody L chain V region, the amino acid valine in position 15 in the FR is replaced by proline that is found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

A plasmid, pBSLV2, containing version 2, shown in SEQ ID NO:40, of the human CDR-grafted anti-GM₂ antibody

L chain V region was obtained following the above procedure using the synthetic DNA of SEQ ID NO:38 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:39 as the mutant sense primer. In the amino acid sequence of the version 2 human CDR-grafted anti-GM₂ antibody L chain V region, the amino acid leucine in positions 46 in the FR is replaced by tryptophan that is found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

A plasmid, pBSLV3, containing version 3, shown in SEQ ID NO:43, of the human CDR-grafted anti-GM₂ antibody L chain V region was obtained following the above procedure using the synthetic DNA of SEQ ID NO:41 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:42 as the mutant sense primer. In the amino acid sequence of the version 3 human CDR-grafted anti-GM₂ antibody L chain V region, proline in position 79 and isoleucine in position 82 in the FR are both replaced by alanine that is found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

Then, a plasmid, pBSLV1+2, containing a human CDR-grafted anti-GM₂ antibody L chain V region having both the version 1 and version 2 replacements was constructed in the following manner.

Three µg of the plasmid pBSLV1 obtained as mentioned above was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *HindIII* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.2 µg of an *EcoRI-HindIII* fragment about 0.20 kb in size was recovered.

Then, 3 µg of the plasmid pBSLV2 obtained as mentioned above was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *HindIII* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *EcoRI-HindIII* fragment about 3.2 kb in size was recovered.

Then, 0.1 µg of the *EcoRI-HindIII* fragment of pBSLV1 and 0.1 µg of the *EcoRI-HindIII* fragment of pBSLV2, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSLV1+2 shown in Fig. 34 was obtained.

Then, the PCR reaction procedure mentioned above was followed using 1 ng of the plasmid pBSLV1+2 obtained as mentioned above as the template, a synthetic DNA having the base sequence of SEQ ID NO:44 as the mutant antisense primer and a synthetic DNA having the base sequence of SEQ ID NO:45 as the mutant sense primer, whereby a plasmid, pBSLV4, containing a version 4 human CDR-grafted anti-GM₂ antibody L chain V region set forth in SEQ ID NO:46 was obtained. In the amino acid sequence of the version 4 human CDR-grafted anti-GM₂ antibody L chain V region, valine in position 15, leucine in position 46, aspartic acid in position 69, phenylalanine in position 70 and threonine in position 71 in the FR are replaced by proline, tryptophan, serine, tyrosine and serine, respectively, that are found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

Then, the PCR reaction procedure mentioned above was followed using 1 ng of the plasmid pBSLV1+2 obtained as mentioned above as the template, a synthetic DNA having the base sequence of SEQ ID NO:47 as the mutant antisense primer and a synthetic DNA having the base sequence of SEQ ID NO:48 as the mutant sense primer, whereby a plasmid, pBSLV8, containing a version 8 human CDR-grafted anti-GM₂ antibody L chain V region set forth in SEQ ID NO:49 was obtained. In the amino acid sequence of the version 8 human CDR-grafted anti-GM₂ antibody L chain V region, valine in position 15, leucine in position 46, aspartic acid in position 69, phenylalanine in position 70, threonine in position 71, serine in position 76, leucine in position 77 and glutamine in position 78 in the FR are replaced by proline, tryptophan, serine, tyrosine, serine, arginine, methionine and glutamic acid, respectively, that are found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

Then, the PCR reaction procedure mentioned above was followed using 1 ng of the plasmid pBSLV4 obtained as mentioned above as the template, a synthetic DNA having the base sequence of SEQ ID NO:50 as the mutant antisense primer and a synthetic DNA having the base sequence of SEQ ID NO:51 as the mutant sense primer, whereby a plasmid, pBSLm-2, containing a version Lm-2 human CDR-grafted anti-GM₂ antibody L chain V region set forth in SEQ ID NO:52 was obtained. In the amino acid sequence of the version Lm-2 human CDR-grafted anti-GM₂ antibody L chain V region, valine in position 15, tyrosine in position 35, leucine in position 46, aspartic acid in position 69, phenylalanine in position 70 and threonine in position 71 in the FR are replaced by proline, phenylalanine, tryptophan, serine, tyrosine and serine, respectively, that are found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

Then, the PCR reaction procedure mentioned above was followed using 1 ng of the plasmid pBSLV4 obtained as mentioned above as the template, a synthetic DNA having the base sequence of SEQ ID NO:53 as the mutant antisense primer and a synthetic DNA having the base sequence of SEQ ID NO:54 as the mutant sense primer, whereby

a plasmid, pBSLm-8, containing a version Lm-8 human CDR-grafted anti-GM₂ antibody L chain V region set forth in SEQ ID NO:55 was obtained. In the amino acid sequence of the version Lm-8 human CDR-grafted anti-GM₂ antibody L chain V region, valine in position 15, leucine in position 46, aspartic acid in position 69, phenylalanine in position 70, threonine in position 71, phenylalanine in position 72 and serine in position 76 in the FR are replaced by proline, tryptophan, serine, tyrosine, serine, leucine and arginine, respectively, that are found in the mouse antibody KM796 L chain V region. This is for the purpose of retaining the antigen-binding capacity of mouse antibody KM796.

Then, a plasmid, pBSLm-28, containing a human CDR-grafted anti-GM₂ antibody L chain V region having both the version Lm-2 and version Lm-8 replacements was constructed in the following manner.

Three µg of the plasmid pBSLm-2 obtained as mentioned above was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *EcoRI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units of the restriction enzyme *XbaI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.2 µg of an *EcoRI*-*XbaI* fragment about 0.24 kb in size was recovered.

Then, 3 µg of the plasmid pBSLm-8 obtained as mentioned above was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *EcoRI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 50 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units of the restriction enzyme *XbaI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *EcoRI*-*XbaI* fragment about 3.16 kb in size was recovered.

Then, 0.1 µg of the *EcoRI*-*XbaI* fragment of pBSLm-2 and 0.1 µg of the *EcoRI*-*XbaI* fragment of pBSLm-8, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pBSLm-28 shown in Fig. 35 was obtained. The version Lm-28 human CDR-grafted anti-GM₂ antibody L chain V region contained in the plasmid pBSLm-28 is shown in SEQ ID NO:8. In the amino acid sequence of the version Lm-28 human CDR-grafted anti-GM₂ antibody L chain V region thus constructed, valine in position 15, tyrosine in position 35, leucine in position 46, aspartic acid in position 69, phenylalanine in position 70, threonine in position 71, phenylalanine in position 72 and serine in position 76 are replaced by proline, phenylalanine, tryptophan, serine, tyrosine, serine, leucine and arginine, respectively, that are found in the mouse antibody KM796 L chain V region. This is for the intended purpose of retaining the antigen-binding capacity of mouse antibody KM796.

(4) Construction of human CDR-grafted anti-GM₂ antibody L chain V region using known HMHCS of human antibody L chain V region

According to Kabat *et al.* (Kabat E. A. *et al.*, "Sequences of Proteins of Immunological Interest", US Dept. of Health and Human Services, 1991), known human antibody L chain V regions are classifiable into subgroups I to IV based on the homology of their FR regions, and common sequences have been identified for respective subgroups. The present inventors identified HMHCS as one meaning from the common sequences, a human CDR-grafted anti-GM₂ antibody L chain V region was constructed based on the HMHCS. First, for selecting common sequences to serve as the base, the homology was examined between the FR of the mouse antibody KM796 L chain V region and the FR of the HMHCS of the human antibody L chain V region of each subgroup (Table 2).

TABLE 2

Homology (%) between mouse antibody KM796 L chain V region FR and human antibody L chain V region common sequence FR			
HSG I	HSG II	HSG III	HSG IV
70.0	65.0	68.8	67.5

As a result, it was confirmed that subgroup I shows the greatest similarity. Thus based on the common sequence of subgroup I, a human CDR-grafted anti-GM₂ antibody L chain V region was constructed by the PCR method in the

following manner.

Synthetic DNAs respectively having the base sequences of SEQ ID NO:56 through SEQ ID NO:61 were synthesized using an automatic DNA synthesizer (Applied Systems model 380A). The DNAs synthesized were added, each to a final concentration of 0.1 μ M, to 50 μ l of 10 mM Tris-hydrochloride buffer (pH 8.3) containing 50 mM potassium chloride, 1.5 mM magnesium chloride, 0.001% gelatin, 200 μ M dNTP, 0.5 μ M M13 primer RV (Takara Shuzo), 0.5 μ M M13 primer M4 (Takara Shuzo) and 2 units of TaKaRa Taq DNA polymerase. The mixture was covered with 50 μ l of mineral oil, a DNA thermal cycler (Perkin Elmer model PJ480) was loaded with the mixture, and 30 PCR cycles (2 minutes at 94°C, 2 minutes at 55°C and 2 minutes at 72°C per cycle) were conducted. The reaction mixture was purified using QIAquick PCR Purification Kit (Qiagen) and then made into a solution in 30 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 μ g/ml BSA, 10 units each of the restriction enzymes *Eco*RI (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.2 μ g of an *Eco*RI-*Sp*II fragment about 0.39 kb in size was recovered.

Then, 0.1 μ g of the above *Eco*RI-*Sp*II fragment of the human CDR-grafted anti-GM₂ antibody L chain V region and 0.1 μ g of the *Eco*RI-*Sp*II fragment of pBSL3 were added to a total of 20 μ l of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101. Plasmid DNAs were prepared from 10 transformant clones and their base sequences were determined. As a result, a plasmid, pBSHSGSL, shown in Fig. 36 and having the desired base sequence was obtained. The amino acid sequence and base sequence of the human CDR-grafted anti-GM₂ antibody L chain V region contained in pBSHSGSL are shown in SEQ ID NO:9. In the amino acid sequence of the thus-constructed human CDR-grafted anti-GM₂ antibody L chain V region, methionine in position 4, leucine in position 11, valine in position 15, tyrosine in position 35, alanine in position 42, leucine in position 46, aspartic acid in position 69, phenylalanine in position 70, threonine in position 71, leucine in position 77 and valine in position 103 in the FR as selected based on a computer model for the V region are replaced by leucine, methionine, proline, phenylalanine, serine, tryptophan, serine, tyrosine, serine, methionine and leucine, respectively, that are found in the mouse antibody KM796 L chain V region. This is for the intended purpose of retaining the antigen-binding capacity of mouse antibody KM796.

2. Activity evaluation of replacement versions of human CDR-grafted anti-GM₂ antibody in terms of transient expression

Various replacement version human CDR-grafted anti-GM₂ antibodies composed of the human CDR-grafted anti-GM₂ antibody H chain and L chain V regions constructed in Paragraphs 3 (1) through (4) of Example 3 and having various replacements were evaluated for activity in terms of transient expression in the following manner.

First, for evaluating the human CDR-grafted anti-GM₂ antibody H chain V regions having various replacements, expression vectors, pT796HCDRHV2, pT796HCDRHV4 and pT796HCDRH10, were constructed by replacing the mouse H chain V region of the mouse-human chimeric anti-GM₂ antibody transient expression vector pT796 obtained in Paragraph 1 (1) of Example 2 of JP-A-6-205694 with the human CDR-grafted anti-GM₂ antibody H chain V regions having various replacements, in the following manner. For comparison, an expression vector, pT796HCDR was constructed by replacing the mouse H chain V region of pT796 with the human CDR-grafted anti-GM₂ antibody H chain V region obtained in Paragraph 1 (1) of Example 2.

Three μ g of the plasmid pT796 was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 μ g/ml BSA, 10 units each of the restriction enzymes *Eco*RI (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 μ g of an *Eco*RI-*Sp*II fragment about 9.20 kb in size was recovered. Then, 3 μ g of the plasmid pBSL16 obtained in Paragraph 1 (3) of Example 3 was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 μ g/ml BSA, 10 units each of the restriction enzymes *Eco*RI (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 μ g of an *Eco*RI-*Sp*II fragment about 0.39 kb in size was recovered.

Then, 0.1 μ g of the *Eco*RI-*Sp*II fragment of pT796 and 0.1 μ g of the *Eco*RI-*Sp*II fragment of pBSL16, each obtained as mentioned above, were added to a total of 20 μ l of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pT796LCDR shown in Fig. 37 was obtained.

Then, 3 μ g of the above plasmid pT796LCDR was added to 10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Ap*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 μ g/ml BSA and 0.01% Triton X-100, 10 units of the restriction

enzyme *NotI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *Apal-NotI* fragment about 9.11 kb in size was recovered.

Then, 0.1 µg of the human CDR-grafted anti-GM₂ antibody H chain V region obtained in Paragraph 1 (1) of Example 2 of JP-A-6-205694 or the replacement version 2 or 4 human CDR-grafted anti-GM₂ antibody H chain V region obtained in Paragraph 1 (1) of Example 3 and 0.1 µg of the *Apal-NotI* fragment of pT796LCDR were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). Each recombinant plasmid DNA solution thus obtained was used to transform *Escherichia coli* HB101. The plasmids pT796HLCDDR, pT796HLCDDRHV2 and pT796HLCDDRHV4 shown in Fig. 38 were obtained.

Then, 3 µg of the plasmid pBSH10 obtained in Paragraph 1 (2) of Example 3 was added to 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Apal* (Takara Shuzo) was further added, and the restriction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 µg/ml BSA and 0.01% Triton X-100, 10 units of the restriction enzyme *NotI* (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 µg of an *Apal-NotI* fragment about 0.44 kb in size was recovered.

Then, 0.1 µg of the *Apal-NotI* fragment of pBSM10 and 0.1 µg of the *Apal-NotI* fragment of pT796LCDR were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). The thus-obtained recombinant plasmid DNA solution was used to transform *Escherichia coli* HB101, and the plasmid pT796HLCDDRH10 shown in Fig. 39 was obtained.

Then, 3 µg each of the plasmids pT796HLCDDR, pT796HLCDDRHV2, pT796HLCDDRHV4 and pT796HLCDDRH10 were respectively added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *SpI* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. Each reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *EcoRI-SpI* fragment about 9.15 kb in size was recovered.

Then, 5 µg of the plasmid pBSL3 obtained in Paragraph 1 (2) of Example 2 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *EcoRI* (Takara Shuzo) and *SpI* (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 0.4 µg of an *EcoRI-SpI* fragment about 0.39 kb in size was recovered.

Then, 0.1 µg of the *EcoRI-SpI* fragment of each of pT796HLCDDR, pT796HLCDDRHV2, pT796HLCDDRHV4 and pT796HLCDDRH10 and 0.1 µg of the *EcoRI-SpI* fragment of pBSL3 were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go DNA Ligase (Pharmacia Biotech). Each recombinant plasmid DNA solution thus obtained was used to transform *Escherichia coli* HB101. In this way, the plasmids pT796HLCDDR, pT796HLCDDRHV2, pT796HLCDDRHV4 and pT796HLCDDRH10 shown in Fig. 40 were obtained.

Then, 2 µg each of the plasmids pT796HLCDDR, pT796HLCDDRHV2, pT796HLCDDRHV4 and pT796HLCDDRH10 thus obtained were used for transient human CDR-grafted anti-GM₂ antibody expression and for culture supernatant human CDR-grafted anti-GM₂ antibody activity evaluation by the procedures described in Paragraphs 1 (5), 2 (2) and (3) of Example 2. After introduction of each plasmid, the culture supernatant was recovered at 72 hours, and the GM₂-binding activity and antibody concentration in the culture supernatant were determined by ELISA and the relative activity was calculated with the activity of the positive control chimera antibody taken as 100%. The results are shown in Fig. 41.

The results revealed that the amino acid residue replacements alone in replacement versions 2 and 4 have little influence on the restoration of the antigen-binding activity of the human CDR-grafted anti-GM₂ antibody but that the use of the pBSH10-derived human CDR-grafted antibody H chain V region constructed based on the known HMHCS of the human antibody H chain V region, contributes to the restoration of the antigen-binding activity.

In view of the above results, the human CDR-grafted anti-GM₂ antibody H chain V region constructed based on the known HMHCS of the human antibody H chain V region as shown in SEQ ID NO:7 was selected as a novel human CDR-grafted anti-GM₂ antibody H chain V region.

Then, for evaluating the human CDR-grafted anti-GM₂ antibody L chain V regions having various replacements, expression vectors, pT796HLCDDL1, pT796HLCDDL2, pT796HLCDDL3, pT796HLCDDL4, pT796HLCDDL8, pT796HLCDDLm-2, pT796HLCDDLm-8, pT796HLCDDLm-28, and pT796HLCDDHSG, were constructed in the following manner by replacing the mouse L chain V region of the vector pT796HLCDDRH10 for transient human CDR-grafted anti-GM₂ antibody expression obtained as mentioned above with the human CDR-grafted anti-GM₂ antibody L chain V regions having various replacements.

Thus, 3 µg of the plasmid pT796HLCDDRH10 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restric-

tion enzymes *Eco*RI (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of an *Eco*RI-*Sp*II fragment about 9.15 kb in size was recovered.

Then, 3 µg of the plasmid pBSLV1, pBSLV2, pBSLV3, pBSLV4, pBSLV8, pBSLm-2, pBSLm-8, pBSLm-28 or pBSHSGSL obtained in Paragraph 1 (3) or (4) of Example 3 was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA, 10 units each of the restriction enzymes *Eco*RI (Takara Shuzo) and *Sp*II (Takara Shuzo) were further added, and the reaction was allowed to proceed at 37°C for 1 hour. Each reaction mixture was fractionated by agarose gel electrophoresis, and about 0.3 µg of an *Eco*RI-*Sp*II fragment about 0.39 kb in size was recovered.

Then, 0.1 µg of the *Eco*RI-*Sp*II fragment of the pT796HCDRLH10, and 0.1 µg of the *Eco*RI-*Sp*II fragment of each replacement version human CDR-grafted anti-GM₂ antibody L chain V region were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). Each recombinant plasmid DNA solution thus obtained was used to transform *Escherichia coli* HB101. In this way, the plasmids pT796HLCDRLV1, pT796HLCDRLV2, pT796HLCDRLV3, pT796HLCDRLV4, pT796HLCDRLV8, pT796HLCDRLm-2, pT796HLCDRLm-8, pT796HLCDRLm-28 and pT796HLCDRHSGSL were obtained as shown in Fig. 42.

Then, 2 µg each of the thus-obtained plasmids pT796HLCDRLV1, pT796HLCDRLV2, pT796HLCDRLV3, pT796HLCDRLV4, pT796HLCDRLV8, pT796HLCDRLm-2, pT796HLCDRLm-8, pT796HLCDRLm-28 and pT796HLCDRHSGSL and of the plasmid pT796HLCDR described in Example 2 of JP-A-6-205694 and capable of expressing human CDR-grafted anti-GM₂ antibody were used for transient human CDR-grafted anti-GM₂ antibody expression and for culture supernatant human CDR-grafted anti-GM₂ antibody activity evaluation by the procedures described in Paragraphs 1 (5) and 2 (2) and (3) of Example 2. After introduction of each plasmid, the culture supernatant was recovered at 72 hours, and the GM₂-binding activity and antibody concentration in the culture supernatant were determined by ELISA and the relative activity was calculated with the activity of the positive control chimera antibody taken as 100%. The results are shown in Fig. 43.

The results revealed that the amino acid residue replacements alone in replacement versions 1, 2, 3, 4 and 8 have little influence on the restoration of the antigen-binding activity of the human CDR-grafted anti-GM₂ antibody but that the amino acid residue replacements in replacement versions Lm-2 and Lm-8 contributes to the restoration of the antigen-binding activity. Furthermore, version Lm-28 having both the amino acid residue replacements of Lm-2 and Lm-8 showed a high level of antigen-binding activity almost comparable to that of the chimera antibody, revealing that those amino acid residues replaced in producing Lm-28 were very important from the antigen-binding activity viewpoint.

In view of the above results, the version Lm-28 human CDR-grafted anti-GM₂ antibody L chain V region shown in SEQ ID NO:8 was selected as a first novel human CDR-grafted anti-GM₂ antibody L chain V region.

It was further revealed that the antigen-binding activity can be restored when the pBSHSGSL-derived human CDR-grafted anti-GM₂ antibody L chain V region, namely the human CDR-grafted anti-GM₂ antibody L chain V region constructed based on the known HMHCS of the human antibody L chain V region, is used.

In view of the above result, the human CDR-grafted anti-GM₂ antibody L chain V region constructed based on the known HMHCS of the human antibody L chain V region as set forth in SEQ ID NO:9 was selected as a second novel human CDR-grafted anti-GM₂ antibody an L chain V region.

It is to be noted that in those human CDR-grafted anti-GM₂ antibody L chain V regions that showed high binding activity against GM₂, certain amino acid residues which cannot be specified by deduction from known human CDR-grafted antibody production examples have been replaced by amino acid residues found in the mouse L chain V region. Thus, obviously, it was very important, in human CDR-grafted anti-GM₂ antibody production, to identify these amino acid residues.

Furthermore, the fact that the human CDR-grafted anti-GM₂ antibodies having those human CDR-grafted anti-GM₂ antibody H chain and L chain V regions based on the known HMHCS of the human antibody V region showed high antigen binding activity is proof of the usefulness of the present process in human CDR-grafted antibody production.

3. Acquisition of cell lines for stable production of human CDR-grafted anti-GM₂ antibodies

Based on the results of Paragraph 2 (5) of Example 3, two cell lines, KM8966 and KM8967, capable of stably expressing KM8966, which has the amino acid sequence set forth in SEQ ID NO:7 as the H chain V region and the amino acid sequence set forth in SEQ ID NO:8 as the L chain V region, and KM8967, which has the amino acid sequence set forth in SEQ ID NO:7 as the H chain V region and the amino acid sequence set forth in SEQ ID NO:9 as the L chain V region, respectively as human CDR-grafted anti-GM₂ antibodies having higher antigen-binding activity than the human CDR-grafted anti-GM₂ antibody described in Example 2 of JP-A-6-205694 were obtained in the following manner.

Three µg each of the plasmids pT796HLCDRLm-28 and pT796HLCDRHSGSL obtained in Paragraph 2 (5) of Example 3 were respectively added to 10 µl of 20 mM Tris-hydrochloride buffer (pH 8.5) containing 100 mM potassium chlo-

ride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Bam*HI (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. Each reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Xho*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. Each reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of a *Bam*HI-*Xho*I fragment about 4.93 kb in size was recovered.

Then, 3 µg of the plasmid pKANTEX93 obtained in Example 1 was added to 10 µl of 20 mM Tris-hydrochloride buffer (pH 8.5) containing 100 mM potassium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Bam*HI (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was subjected to ethanol precipitation, the precipitate was added to 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT, 10 units of the restriction enzyme *Xho*I (Takara Shuzo) was further added, and the reaction was allowed to proceed at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis, and about 1 µg of a *Bam*HI-*Xho*I fragment about 8.68 kb in size was recovered.

Then, 0.1 µg of the *Bam*HI-*Xho*I fragment of pT796HLCDRIm-28 or pT796HLCDRHSGL and 0.1 µg of the *Bam*HI-*Xho*I fragment of pKANTEX93, each obtained as mentioned above, were added to a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). Each recombinant plasmid DNA solution thus obtained was used to transform *Escherichia coli* HB101. In this way, the plasmids pKANTEX796HLCDRIm-28 and pKANTEX796HLCDRHSGL shown in Fig. 44 were obtained.

Then, 4 µg each of the above plasmids pKANTEX796HLCDRIm-28 and pKANTEX796HLCDRHSGL were respectively used to transform YB2/0 (ATCC CRL 1581) cells according to the procedure described in Paragraph 1 (4) of Example 2 and, after final selection using G418 (0.5 mg/ml) and MTX (200 nM), a transformant cell line, KM8966, capable of producing about 40 µg/ml of KM8966, i.e. the pKANTEX796HLCDRIm-28-derived human CDR-grafted anti-GM₂ antibody, and a transformant cell line, KM8967, capable of producing about 30 µg/ml of KM8967, i.e. the pKANTEX796HLCDRHSGL-derived human CDR-grafted anti-GM₂ antibody, were obtained.

The transformants KM8966 and KM8967 have been deposited with National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology (Higashi 1-1-3, Tsukuba, Ibaraki, Japan; hereinafter the address is the same as this) on May 23, 1995 under the deposit numbers FERM BP-5105, and FERM BP-5106, respectively.

4. Purification of human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967

The transformant cell lines KM8966 and 8967 obtained in Paragraph 3 of Example 3 were respectively suspended in GIT medium (Nippon Pharmaceutical) containing 0.5 mg/ml G418 and 200 nM MTX and, according to the procedure of Paragraph 11 of Example 1 of JP-A-6-205694, 18 mg of purified human CDR-grafted anti-GM₂ antibody KM8966 and 12 mg of purified KM8967 were obtained each from about 0.5 liter of culture fluid. Three µg each of the purified human CDR-grafted anti-GM₂ antibodies obtained and the mouse-human chimeric anti-GM₂ antibody KM966 were subjected to electrophoresis by the known method [Laemli, U.K., *Nature*, **227**, 680 (1979)] for molecular weight determination. The results are shown in Fig. 45. As shown in Fig. 45, under reducing conditions, both antibody H chains showed a molecular weight of about 50 kilodaltons and both antibody L chains showed a molecular weight of about 25 kilodaltons. Expression of H and L chains of correct molecular weights was thus confirmed. Under nonreducing conditions, both human CDR-grafted anti-GM₂ antibodies showed a molecular weight of about 150 kilodaltons and it was thus confirmed that antibodies each composed of two H chains and two L chains and having a correct size had been expressed. Furthermore, the H and L chains of each human CDR-grafted anti-GM₂ antibody were analyzed for N-terminal amino acid sequence by automatic Edman degradation using a protein sequencer (Applied Biosystems model 470A), whereby an amino acid sequence deducible from the base sequence of the V region DNA constructed was revealed.

5. *In vitro* reactivity of human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 against GM₂

The mouse-human chimeric anti-GM₂ antibody KM966 and the purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 were tested for reactivity against GM₂ by ELISA as described in Paragraph 1 (5) of Example 2. The results are shown in Fig. 46. GM₂ (N-acetyl-GM₂) used was purified from cultured cell line HPB-ALL [Oboshi *et al.*, *Tanpakushitsu, Kakusan & Koso (Protein, Nucleic Acid & Enzyme)*, **23**, 697 (1978)] in accordance with the known method [J. Biol. Chem., **263**, 10915 (1988)]. As shown, it was found that the purified human CDR-grafted anti-GM₂ antibody KM8966 exerted the binding activity comparable to that of the mouse-human chimeric anti-GM₂ antibody KM966. On the other hand, the binding activity of purified human CDR-grafted anti-GM₂ antibody KM8967 was about 1/4 to 1/5 of that of the mouse-human chimeric anti-GM₂ antibody KM966.

6. Reaction specificity of human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967

The mouse-human chimeric anti-GM₂ antibody KM966 and the human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 were tested for reactivity against the gangliosides GM₁, N-acetyl-GM₂, N-glycolyl-GM₂, N-acetyl-GM₃, N-glycolyl-GM₃, GD_{1a}, GD_{1b} (latron), GD₂, GD₃ (latron) and GQ_{1b} (latron) by ELISA as described in Paragraph 1 (5) of Example 2. The results are shown in Fig. 47. GM₁ and GD_{1a} were purified from bovine brain, N-acetyl-GM₂ from cultured cell line HPB-ALL [Oboshi *et al.*, *Tanpakushitsu, Kakusan & Koso (Protein, Nucleic acid & Enzyme)*, **23**, 697 (1978)], N-glycolyl-GM₂ and N-glycolyl-GM₃ from mouse liver, N-acetyl-GM₃ canine erythrocytes, and GD₂ from cultured cell line IMR32 (ATCC CCL127), respectively by the *per se* known method [J. Biol. Chem., **263**, 10915 (1988)]. Each antibody was used in a concentration of 10 µg/ml.

As shown in Fig. 47, it was confirmed that the human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 react specifically with GM₂ (N-acetyl-GM₂ and N-glycolyl-GM₂) like the mouse-human chimeric anti-GM₂ antibody KM966.

7. Reactivity of human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 against cancer cells

The human lung small cell carcinoma culture cell line SBC-3 (JCRB 0818) (1×10^6 cells) was suspended in PBS, the suspension was placed in a microtube (TREF) and centrifuged (1200 rpm, 2 minutes). To the thus-washed cells was added 50 µl (50 µg/ml) of the mouse-human chimeric anti-GM₂ antibody KM966 or the purified human CDR-grafted anti-GM₂ antibody KM8966 or KM8967, followed by stirring and 1 hour of standing at 4°C. After the above reaction step, the cells were washed three times with PBS, each time followed by centrifugation. Then, 20 µl of fluorescein isocyanate-labeled protein A (30-fold dilution, Boehringer Mannheim) was added and, after stirring, the reaction was allowed to proceed at 4°C for 1 hour. Thereafter, the cells were washed three times with PBS, each time followed by centrifugation, then further suspended in PBS and subjected to analysis using a flow cytometer, EPICS Elite (Coulter). In a control run, the above procedure was followed without addition of the human CDR-grafted anti-GM₂ antibody and analyzed. The results are shown in Fig. 48. It was found that the purified human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 strongly reacted with the human lung small cell carcinoma culture cell line SBC-3 like the mouse-human chimeric anti-GM₂ antibody KM966.

8. *In vitro* antitumor activity of human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967: CDC activity

(1) Preparation of target cells

The target cells SBC-3, cultured in RPMI1640-FCS (10) medium supplemented with 10% FCS, here adjusted to a cell concentration of 5×10^6 cells/500 µl, 3.7 MBq of Na₂⁵¹CrO₄ (Daiichi Pure Chemicals Co., Ltd.) was added thereto. Then, the reaction was allowed to proceed at 37°C for 1 hour, and the cells were washed three times with the medium. The cells were then allowed to stand in the medium at 4°C for 30 minutes and, after centrifugation, the medium was added to adjust the cell concentration to 1×10^6 cells/ml.

(2) Preparation of the complement

Sera from healthy subjects were combined and used as a complement source.

(3) CDC activity measurement

The mouse-human chimeric anti-GM₂ antibody KM966 or purified human CDR-grafted anti-GM₂ antibody KM8966 or KM8967 was added to wells of 96-well U-bottom plates within the final concentration range of 0.05 to 50 µg/ml and then 50 µl (5×10^4 cells/well) of the target cells prepared in (1) were added to each well. The reaction was allowed to proceed at room temperature for 1 hour. After centrifugation, the supernatants were discarded, the human complement obtained in (2) was added to each well to give a final concentration of 15% v/v, and the reaction was allowed to proceed at 37°C for 1 hour. After centrifugation, the amount of ⁵¹Cr in each supernatant was determined using a gamma counter. The amount of spontaneously dissociated ⁵¹Cr was determined by adding to the target cells the medium alone in stead of the antibody and complement solutions and measuring the amount of ⁵¹Cr in the supernatant in the same manner as mentioned above. The total amount of dissociated ⁵¹Cr was determined by adding to the target cells 1 N hydrochloric acid in stead of the antibody and complement solutions and measuring the amount of ⁵¹Cr in the supernatant in the same manner as mentioned above. The CDC activity was calculated as follows:

$$\text{CDC activity (\%)} = \frac{\text{Amount of } ^{51}\text{Cr in sample supernatant} - \text{Amount of } ^{51}\text{Cr spontaneously dissociated}}{\text{Total amount of } ^{51}\text{Cr dissociated} - \text{Amount of } ^{51}\text{Cr spontaneously dissociated}} \times 100$$

The results thus obtained are shown in Fig. 49. It was shown that CDC activity of the human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 was lower than that of the mouse-human chimeric anti-GM₂ antibody KM966.

9. *In vitro* antitumor activity of human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967: ADCC activity

(1) Preparation of target cells

The target cells SBC-3 cultured in RPMI1640-FCS (10) medium supplemented with 10% FCS were adjusted to a cell concentration of 1×10^6 cells/500 μ l. 3.7 MBq of Na₂⁵¹CrO₄ (Daiichi Pure Chemicals Co., Ltd.) was added thereto. Then, the reaction was allowed to proceed at 37°C for 1 hour and the cells were washed three times with the medium.

The cells were then allowed to stand in the medium at 4°C for 30 minutes and then, after centrifugation, the medium was added to adjust the cell concentration to 2×10^5 cells/ml.

(2) Preparation of effector cells

Human venous blood (50 ml) was collected, 0.5 ml of heparin sodium (Takeda Chemical Industries; 1,000 units/ml) was added, and the mixture was gently stirred. This mixture was overlaid on Polymorphprep (Nycomed) and centrifuged to separate the lymphocyte layer (PBMC). The resulting lymphocytes were washed three times by centrifugation with RPMI1640 medium supplemented with 10% FCS, and the cells were suspended in the medium (5×10^6 cells/ml) for use as effector cells.

(3) ADCC activity measurement

The mouse-human chimeric anti-GM₂ antibody KM966 or purified human CDR-grafted anti-GM₂ antibodies KM8966 or KM8967 was added to wells of 96-well U-bottom plates within the final concentration range of 0.05 to 50 μ g/ml and then 50 μ l (1×10^4 cells/well) of the target cell suspension prepared in (1) and 100 μ l (5×10^5 cells/well) of the effector cell suspension prepared in (2) were added to each well. The reaction was allowed to proceed at 37°C for 4 hours and, after centrifugation, the amount of ⁵¹Cr in each supernatant was measured using a gamma counter. The amount of spontaneously dissociated ⁵¹Cr was determined by adding to the target cells the medium alone in lieu of the antibody and effector cells and measuring the amount of ⁵¹Cr in the supernatant in the same manner as mentioned above. The total amount of dissociated ⁵¹Cr was determined by adding to the target cells 1 N hydrochloric acid in lieu of the antibody and effector cells and measuring the amount of ⁵¹Cr in the supernatant in the same manner as mentioned above. The ADCC activity was calculated as follows:

$$\text{ADCC activity (\%)} = \frac{\text{Amount of } ^{51}\text{Cr in sample supernatant} - \text{Amount of } ^{51}\text{Cr spontaneously dissociated}}{\text{Total amount of } ^{51}\text{Cr dissociated} - \text{Amount of } ^{51}\text{Cr spontaneously dissociated}} \times 100$$

The results thus obtained are shown in Fig. 50. The human CDR-grafted anti-GM₂ antibody KM8966 showed ADCC activity comparable to that of the mouse-human chimeric anti-GM₂ antibody KM966, whereas the human CDR-grafted anti-GM₂ antibody KM8967 showed ADCC activity slightly lower than that of the mouse-human chimeric anti-GM₂ antibody KM966.

EXAMPLE 4

Production of human CDR-grafted anti-GM₂ antibodies II

The human CDR-grafted anti-GM₂ antibodies KM8966 and KM8967 showed antigen binding activity (ELISA), binding specificity and ADCC activity comparable to those of the mouse-human chimeric anti-GM₂ antibody KM966, while its CDC activity was lower than that of the chimeric antibody. In order to improve the CDC activity, human CDR-grafted anti-GM₂ antibodies were produced in the following manner.

1. Modification of human CDR-grafted anti-GM₂ antibody KM8966 H chain V region

Among the human CDR-grafted anti-GM₂ antibodies prepared in Example 3, the antibody KM8966 showing higher CDC activity was subjected to amino acid residue replacements at the H chain V region (SEQ ID NO:7) in order to improve CDC activity. The amino acid residues to be replaced were selected at random with reference to the results of various replacement obtained in Example 3 and a computer model for the V region of mouse antibody KM796. Replacements were introduced by PCR method using as a template 1 ng of the plasmid pBSH10 containing the human CDR-grafted anti-GM₂ antibody H chain V region obtained in Paragraph 1 (2) of Example 3 and using as a primer antisense and sense synthetic DNA containing mutations described in Paragraph 1 (3) of Example 3.

The reaction was carried out in the same manner as described in Paragraph 1 (3) of Example 3 using the synthetic DNA of SEQ ID NO:62 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:63 as the mutant sense primer to obtain the plasmid pBSHM1 containing version HM1, shown in SEQ ID NO:64, of the human CDR-grafted anti-GM₂ antibody H chain V region. In the amino acid sequence of the version HM1, arginine in position 38, alanine in position 40, glutamine in position 43 and glycine in position 44 in the FR shown in SEQ ID NO:7 were replaced by lysine, serine, lysine and serine, respectively, that are found in the mouse antibody KM796 H chain V region.

The plasmid pBSHM2 containing version HM2, shown in SEQ ID NO:10, of the human CDR-grafted anti-GM₂ antibody H chain V region was obtained following the reaction described in Paragraph 1 (3) of Example 3 using the synthetic DNA of SEQ ID NO:65 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:66 as the mutant sense primer. In the amino acid sequence of the version HM2, arginine in position 38 and alanine in position 40 in the FR shown in SEQ ID NO:7 were replaced by lysine and serine, respectively, that are found in the mouse antibody KM796 H chain V region.

The plasmid pBSHM3 containing version BM3, shown in SEQ ID NO:69, of the human CDR-grafted anti-GM₂ antibody H chain V region was obtained following the reaction described in Paragraph 1 (3) of Example 3 using the synthetic DNA of SEQ ID NO:67 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:68 as the mutant sense primer. In the amino acid sequence of the version HM3, valine in position 68 and isoleucine in position 70 in the FR shown in SEQ ID NO:7 were replaced by alanine and leucine, respectively, that are found in the mouse antibody KM796 H chain V region.

The plasmid pBSHM31 containing version HM31, shown in SEQ ID NO:70, of the human CDR-grafted anti-GM₂ antibody H chain V region was obtained following the reaction described in Paragraph 1 (3) of Example 3 using 1 ng of the plasmid pBSHM3 as the template, the synthetic DNA of SEQ ID NO:62 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:63 as the mutant sense primer. In the amino acid sequence of the version HM31, arginine in position 38, alanine in position 40, glutamine in position 43 and glycine in position 44 in the FR of the version HM3 were replaced by lysine, serine, lysine and serine, respectively, that are found in the mouse antibody KM796 H chain V region.

Further, the plasmid pBSHM32 containing version HM32, shown in SEQ ID NO:71, of the human CDR-grafted anti-GM₂ antibody H chain V region was obtained following the reaction described in Paragraph 1 (3) of Example 3 using 1 ng of the plasmid pBSHM3 as the template, the synthetic DNA of SEQ ID NO:65 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:66 as the mutant sense primer. In the amino acid sequence of the version HM32, arginine in position 38 and alanine in position 40 in the FR of the version HM3 were replaced by lysine and serine, respectively, that are found in the mouse antibody KM796 H chain V region.

2. Evaluation of CDC activity of human CDR-grafted anti-GM₂ antibodies having various replacements in the human CDR-grafted anti-GM₂ antibody H chain V region

(1) Construction of expression vectors

Expression vectors for various human CDR-grafted anti-GM₂ antibodies containing the H chain V region of human CDR-grafted anti-GM₂ antibodies having various replacements obtained in Paragraph 1 of Example 4 and the L chain V region of KM8966 (SEQ ID NO:8) were prepared in the following manner.

Three µg each of the plasmids pBSHM1, pBSHM2, pBSHM3, pBSHM31 and pBSHM32 obtained in Paragraph 1 of Example 4 were dissolved in 10 µl of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of *Apal* (Takara Shuzo) were added thereto and the mixture was allowed to react at 37°C for 1 hour. The resulting mixture was subjected to ethanol precipitation and the thus-obtained precipitate was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 µg/ml BSA and 0.01% of Triton X-100. Ten units of *NotI* (Takara Shuzo) were further added thereto to allow the mixture to react at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis to recover about 0.2 µg of the *Apal*-*NotI* fragment of about 0.44 kb.

Then, 3 µg of the plasmid pKANTEX796HLCDR_LM-28 obtained in Paragraph 3 (3) of Example 3 was dissolved in

10 μ l of 10 mM Tris-hydrochloride buffer (pH 7.5) containing 10 mM magnesium chloride and 1 mM DTT, 10 units of *Apal* (Takara-Shuzo) were added thereto and the mixture was allowed to react at 37°C for 1 hour. The resulting mixture was subjected to ethanol precipitation and the thus-obtained precipitate was dissolved in 10 μ l of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT, 100 μ g/ml BSA and 0.01% of Triton X-100. 10 units of *NotI* (Takara Shuzo) were added thereto to allow the mixture to react at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis to recover about 1 μ g of the *Apal*-*NotI* fragment of about 13.14 kb.

About 0.1 μ g each of the thus-obtained *Apal*-*NotI* fragment of pBSHM1, pBSHM2, pBSHM3, pBSHM31 and pBSHM32 and 0.1 μ g of the *Apal*-*NotI* fragment of pKANTEX796HLCDR_LM-28 were added in a total of 20 μ l of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). Each of the resulting recombinant plasmid DNA solutions was used to transform *Escherichia coli* HB101 and plasmids, pKANTEX796HM1Lm-28, pKANTEX796HM2Lm-28, pKANTEX796HM3Lm-28, pKANTEX796HM31Lm-28 and pKANTEX796HM32Lm-28 shown in Fig. 51 were obtained.

(2) Expression of replacement versions of human CDR-grafted anti-GM₂ antibodies

Four μ g each of the plasmids pKANTEX796HM1Lm-28, pKANTEX796HM2Lm-28, pKANTEX796HM3Lm-28, pKANTEX796HM31Lm-28 and pKANTEX796HM32Lm-28 obtained in Paragraph 2 (1) of Example 4 were used to transform YB2/O cells (ATCC CRL 1581) in accordance with the method as described in Paragraph 1 (4) of Example 2. The cells were ultimately selected using G418 (0.5 mg/ml) and MTX (200 nM) to obtain about 2 to 5 μ g/ml of transformants capable of producing human CDR-grafted anti-GM₂ antibodies derived from the corresponding expression vectors.

(3) Purification of replacement versions of human CDR-grafted anti-GM₂ antibodies

Cells of each transformant obtained in Paragraph 2 (2) of Example 4 were suspended in GIT medium (Nihon Pharmaceutical) containing 0.5 mg/ml G418 and 200 nM MTX and about 1 to 3 mg of purified human CDR-grafted anti-GM₂ antibodies were obtained from about 0.6 liter of the culture broth in accordance with the method described in Paragraph 11 of Example 1 of JP-A-6-205694. The human CDR-grafted anti-GM₂ antibodies derived from the plasmids pKANTEX796HM1Lm-28, pKANTEX796HM2Lm-28, pKANTEX796HM3Lm-28, pKANTEX796HM31Lm-28 and pKANTEX796HM32Lm-28 are hereinafter referred to as "M1-28", "M2-28", "M3-28", "M31-28" and "M32-28", respectively. 4 μ g each of the purified human CDR-grafted anti-GM₂ antibodies, the human CDR-grafted anti-GM₂ antibody KM8966 and the mouse-human chimeric anti-GM₂ antibody KM966 were electrophoresed by the conventional method [Laemmli: *Nature*, 227, 680 (1970)] for molecular weight checking. The results are shown in Fig. 52. As shown in Fig. 52, under reducing conditions, the molecular weight of the antibody H chain was about 50 KDa and the molecular weight of the antibody L chain was about 25 KDa, thus confirming the expression of the H chain and L chain having the correct molecular weight. Under nonreducing conditions, the molecular weight of the human CDR-grafted anti-GM₂ antibodies was about 150 KDa, confirming that the antibody expressed was composed of two H chains and two L chains and was correct in size. The N-terminal amino acid sequence of the H and L chains of each purified human CDR-grafted anti-GM₂ antibodies was examined by automatic Edman degradation using a protein sequencer (Applied Biosystems model 470A). As a result, it was confirmed that the amino acid sequence was consistent with that deduced from the synthesized V region DNA sequence.

(4) CDC activity of replacement versions of human CDR-grafted anti-GM₂ antibodies

CDC activity of the replacement versions of the human CDR-grafted anti-GM₂ antibodies obtained in Paragraph 2 (3) of Example 4, the human CDR-grafted anti-GM₂ antibody KM8966 and the mouse-human chimeric anti-GM₂ antibody KM966 was measured in accordance with the method described in Paragraph 8 of Example 3. The results are shown in Fig. 53. As shown in Fig. 53, it was found that, among the replacement versions of the human CDR-grafted anti-GM₂ antibodies, the human CDR-grafted anti-GM₂ antibody M2-28 derived from the plasmid pKANTEX796HM2Lm-28 showed the highest CDC activity which was higher than that of the human CDR-grafted anti-GM₂ antibody KM8966 prepared in Example 3. This result indicates that the replaced amino acid residues of the version HM2 among the various replacement versions prepared in Paragraph 1 of Example 4 play an important role for improving CDC activity. It was assumed from the computer model for the V region of mouse antibody KM796 that the replacement of the amino acid residues of the version HM2 would influence on the entire structure of the V region since these amino acid residues are located at the site which interacts with the L chain V region. Recent study of the production of human CDR-grafted antibody reveals that the amino acid residues which affect the structure of the antibody varies in each antibody. No method for precisely predicting such amino acid residues has been established and the above results provide a significant finding for the production of the human CDR-grafted antibody.

The human CDR-grafted anti-GM₂ antibody M2-28 derived from the plasmid pKANTEX796HM2Lm-28 was designated as KM8970 and the antibody KM8970-producing transformant KM8970 has been deposited with National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology as of May 9, 1996 under the deposit number FERM BP-5528.

3. Modification of human CDR-grafted anti-GM₂ antibody KM8966 L chain V region

The human CDR-grafted anti-GM₂ antibody KM8966 prepared in Example 3 was subjected to amino acid residue replacements in the L chain V region (SEQ ID NO:8) to improve CDC activity. As an amino acid residue to be replaced, serine residue in position 59 was selected based on the results of various replacements obtained in Paragraph 1 (3) of Example 3 which suggested that it was important to support the structure of CDR2 for the human CDR-grafted anti-GM₂ antibody activity. Replacements were introduced by PCR method using as a template 1 ng of the plasmid pBSLm-28 containing the human CDR-grafted anti-GM₂ antibody L chain V region obtained in Paragraph 1 (3) of Example 3 and using as a primer antisense and sense synthetic DNA containing mutations described in Paragraph 1 (3) of Example 3.

The reaction was carried out in the same manner as described in Paragraph 1 (3) of Example 3 using the synthetic DNA of SEQ ID NO:72 as the mutant antisense primer and the synthetic DNA of SEQ ID NO:73 as the mutant sense primer to obtain the plasmid pBSLm-28 No.1, containing version Lm-28 No.1, shown in SEQ ID NO:11, of the human CDR-grafted anti-GM₂ antibody L chain V region. In the amino acid sequence of the version Lm-28 No.1, serine in position 59 in the FR shown in SEQ ID NO:83 was replaced by alanine that is found in the mouse antibody KM796 L chain V region.

4. Evaluation of CDC activity of human CDR-grafted anti-GM₂ antibody having new replacement in human CDR-grafted anti-GM₂ antibody L chain V region

(1) Construction of expression vectors

Expression vectors for the human CDR-grafted anti-GM₂ antibody containing the human CDR-grafted anti-GM₂ antibody L chain V region having the replacement obtained in Paragraph 3 of Example 4 and the human CDR-grafted anti-GM₂ antibody H chain V region were obtained in the following manner.

Six µg of the plasmid pBSLm-28 No.1 obtained in Paragraph 3 of Example 4 was dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride, 1 mM DTT and 100 µg/ml BSA. 10 units each of *Eco*RI (Takara Shuzo) and *Sp*I (Takara Shuzo) were added thereto to allow the mixture to react at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis to recover about 0.4 µg of the *Eco*RI-*Sp*I fragment of about 0.39 kb.

Then, 3 µg each of the plasmid pKANTEX796HLCDRLm-28 obtained in Paragraph 3 of Example 3 and the plasmids pKANTEX796HM1Lm-28, pKANTEX796HM2Lm-28 and pKANTEX796HM3Lm-28 obtained in Paragraph 2 (1) of Example 4 were dissolved in 10 µl of 50 mM Tris-hydrochloride buffer (pH 7.5) containing 100 mM sodium chloride, 10 mM magnesium chloride and 1 mM DTT and 100 µg/ml BSA, 10 units each of *Eco*RI (Takara Shuzo) and *Sp*I were added thereto and the mixture was allowed to react at 37°C for 1 hour. The reaction mixture was fractionated by agarose gel electrophoresis to recover about 1 µg of the *Eco*RI-*Sp*I fragment of about 13.19 kb.

A 0.1 µg portion each of the thus-obtained *Eco*RI-*Sp*I fragment of pBSLm-28 No.1 and 0.1 µg of the *Eco*RI-*Sp*I of pKANTEX796HLCDRLm-28, pKANTEX796HM1Lm-28, pKANTEX796HM2Lm-28 and pKANTEX796HM3Lm-28 were added in a total of 20 µl of sterilized water and ligated to each other using Ready-To-Go T4 DNA Ligase (Pharmacia Biotech). Each of the resulting recombinant plasmid DNA solutions was used to transform *Escherichia coli* HB101 and the plasmids pKANTEX796HLm-28 No.1, pKANTEX796HM1Lm-28 No.1, pKANTEX796HM2Lm-28 No.1 and pKANTEX796HM3Lm-28 No.1 shown in Fig. 54 were obtained.

(2) Expression of human CDR-grafted anti-GM₂ antibodies having replacements in the L chain V region

Four µg each of the plasmids pKANTEX796HLm-28 No.1, pKANTEX796HM1Lm-28 No.1, pKANTEX796HM2Lm-28 No.1 and pKANTEX796HM3Lm-28 No.1 obtained in Paragraph 4 (1) of Example 4 was used to transform YB2/O cells (ATCC CRL 1581) in accordance with the method as described in Paragraph 11 of Example 1. The cells were ultimately selected using G418 (0.5 mg/ml) and MTX (200 nM) to obtain about 2 to 5 µg/ml of transformants capable of producing human CDR-grafted anti-GM₂ antibodies derived from the corresponding expression vectors.

(3) Purification of human CDR-grafted anti-GM₂ antibodies having replacements in the L chain V region

Cells of each transformant obtained in Paragraph 4 (2) of Example 4 were suspended in GIT medium (Nihon Pharmaceutical) containing 0.5 mg/ml G418 and 200 nM MTX and about 1 to 3 mg of purified human CDR-grafted anti-GM₂ antibodies were obtained from about 0.6 liter of the culture broth in accordance with the method described in Paragraph 11 of Example 1 of JP-A-6-205694. The human CDR-grafted anti-GM₂ antibodies derived from the plasmids pKANTEX796HLM-28 No.1, pKANTEX796HM1Lm-28 No.1, pKANTEX796HM2Lm-28 No.1 and pKANTEX796HM3Lm-28 No.1 are hereinafter referred to as "h796H-No.1", "M1-No.1", "M2-No.1" and "M3-No.1", respectively. Four µg each of the purified human CDR-grafted anti-GM₂ antibodies and the mouse-human chimeric anti-GM₂ antibody KM966 was electrophoresed by the conventional method [Laemmli: *Nature*, 227, 680 (1970)] for molecular weight checking. The results are shown in Fig. 55. As shown in Fig. 55, under reducing conditions, the molecular weight of the antibody H chain was about 50 KDa and the molecular weight of the antibody L chain was about 25 KDa, thus confirming the expression of the H chain and L chain having the correct molecular weight. Under nonreducing conditions, the molecular weight of the human CDR-grafted anti-GM₂ antibodies was about 150 KDa, confirming that the antibody expressed was composed of two H chains and two L chains and was correct in size. The N-terminal amino acid sequence of the H and L chains of each purified human CDR-grafted anti-GM₂ antibodies was examined by automatic Edman degradation using a protein sequencer (Applied Biosystems model 470A). As a result, it was confirmed that the amino acid sequence was consistent with that deduced from the synthesized V region DNA sequence.

(4) CDC activity of human CDR-grafted anti-GM₂ antibodies having replacements in the L chain V region

CDC activity of the human CDR-grafted anti-GM₂ antibodies having replacements in the L chain V region obtained in Paragraph 4 (3) of Example 4, the human CDR-grafted anti-GM₂ antibody KM8970, the human CDR-grafted anti-GM₂ antibody KM8966 and the mouse-human chimeric anti-GM₂ antibody KM966 was measured in accordance with the method described in Paragraph 8 of Example 3. The results are shown in Fig. 56. Comparing CDC activity of KM8966 with that of h796H-No.1, it was found that the replacement introduced into only the L chain V region showed improved CDC activity. Among the replaced antibodies having replacements in both of the L chain V region and the H chain V region, M2-No.1 having replacement in the human CDR-grafted anti-GM₂ antibody KM8970 H and L chain V region obtained in Paragraph 2 of Example 4 showed the highest CDC activity, which was comparable to or higher than that of KM8970. These results indicate that the replaced amino acid residue in position 59 in the FR of the L chain V region prepared in Paragraph 3 of Example 4 played an important role for improving its CDC activity and it interacted with the replaced amino acid residue in the H chain V region of KM8970 for improving its CDC activity cooperatively. It was not assumed from the computer model for the V region of mouse antibody KM796 that the replacement of the amino acid residue in position 59 in the FR of the version Lm-28 No.1 would be involved in direct action with antigen GM₂ and interaction with each CDR residue. However, the above results suggested that they were quite important for maintaining the entire structure of the whole V region. This knowledge cannot be predicted from the known production method of a humanized antibody, and the above findings will provide an important indication for the production of human CDR-grafted antibody.

The human CDR-grafted anti-GM₂ antibody M2-No.1 derived from the plasmid pKANTEX796HM2Lm-28 No.1 was designated as KM8969 and the antibody KM8969-producing transformant KM8969 has been deposited with National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology as of May 9, 1996 under the deposit number FERM BP-5527.

5. *In vitro* reactivity of human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 with GM₂

Reactivities of the mouse-human chimeric anti-GM₂ antibody KM966 and the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 with GM₂ were measured in accordance with the method described in Paragraph 1 (5) of Example 2. The results are shown in Fig. 57. As shown in Fig. 57, the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 showed binding activity comparable to that of the mouse-human chimeric anti-GM₂ antibody KM966.

6. Reaction specificity of human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970

The mouse-human chimeric anti-GM₂ antibody KM966 and the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 were examined for reactivity with various gangliosides in accordance with the method described in Paragraph 6 of Example 3. The results are shown in Fig. 58. As shown in Fig. 58, it was found that the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 specifically reacted with GM₂ (N-acetyl GM₂ and N-glycolyl GM₂) like the mouse-human chimeric anti-GM₂ antibody KM966.

7. Reactivity of human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 with cancer cells

The mouse-human chimeric anti-GM₂ antibody KM966 and the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 were examined for reactivity with the human lung small cell carcinoma cell line SBC-3 (JCRB 0818) using fluorescein isocyanate-labeled rabbit anti-human IgG antibody (Dako) as a second antibody in accordance with the method described in Paragraph 7 of Example 3. The results are shown in Fig. 59. As shown in Fig. 59, the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 strongly reacted with the human lung small cell carcinoma cell line SBC-3 like the mouse-human chimeric anti-GM₂ antibody KM966.

10 8. In vitro antitumor effect of human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970: antibody dependent cell mediated cytotoxicity (ADCC)

The mouse-human chimeric anti-GM₂ antibody KM966 and the human CDR-grafted anti-GM₂ antibodies KM8966, KM8969 and KM8970 were examined for ADCC activity against the human lung small cell carcinoma cell line SBC-3 (JCRB 0818) in accordance with the method described in Paragraph 9 of Example 3. The results are shown in Fig. 123. As shown in Fig. 123, the human CDR-grafted anti-GM₂ antibodies KM8969 and KM8970 showed ADCC activity comparable to that of the mouse-human chimeric anti-GM₂ antibody KM966.

20 9. Comparison of *in vitro* anti-tumor activities of humanized anti-GM₂ antibodies: comparison of CDC activity

CDC activities of various humanized anti-GM₂ antibodies (KM966, KM8966, KM8969 and KM8970) established in the aforementioned Inventive Examples 3 and 4 were compared by prolonging the reaction time. Illustratively, the reaction time of the method described in the item 8 of Inventive Example 3 after addition of the human complement was set to 4 hours. The results are shown in Fig. 61. As shown in Fig. 61, it was revealed that the CDC activity of each of these humanized antibodies increases by the 4 hours of reaction and, at an antibody concentration of 5 µg/ml or more, the mouse-human chimeric anti-GM₂ antibody KM966 and the human CDR-grafted anti-GM₂ antibodies KM8966, KM8969 and KM8970 show almost the same level of CDC activity. Particularly, KM8969 showed the highest CDC activity which was about 1/2 of that of the mouse-human chimeric anti-GM₂ antibody KM966, so that it was revealed that a human CDR-grafted anti-GM₂ antibody having further high CDC activity was able to be produced by the examination of Inventive Example 4.

Thus, production method of human CDR-grafted anti-GM₂ antibodies and evaluation of their various activities have been described, and these results show that the established human CDR-grafted anti-GM₂ antibodies are useful for the treatment of human cancers.

By the present invention, human CDR-grafted antibodies to ganglioside GM₂, whose binding activity and binding specificity for GM₂ and anti-tumor effect upon ganglioside GM₂-positive cells are comparable to the levels of chimeric human antibodies, and the production method thereof are provided.

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50

55

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Kyowa Hakko Kogyo Co., Ltd

(ii) TITLE OF INVENTION: Human complementarity determining Region
(CDR)-grafted antibody to ganglioside
GM2

(iii) NUMBER OF SEQUENCES: 73

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Kyowa Hakko Kogyo Co., Ltd.
(B) STREET: 6-1, Ohtemachi 1-chome, Chiyoda-ku
(C) CITY: Tokyo
(E) COUNTRY: Japan

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: 98105047.9
(B) FILING DATE: 10.03.1998

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Kinzebach, Werner, Dr.
(C) REFERENCE/DOCKET NUMBER: M/39063

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (089) 998397-0
(B) TELEFAX: (089) 987304

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 5 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Asp Tyr Asn Met Asp

1

5

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

5 Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn Gln Lys Phe Lys
 1 5 10 15
 Ser
 17

10 (2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 11 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

20 Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr
 1 5 10 11

(2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 10 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ser Ala Ser Ser Ser Val Ser Tyr Met His
 1 5 10

35 (2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 7 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

40 (ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

45 Ser Thr Ser Asn Leu Ala Ser
 1 5 7

(2) INFORMATION FOR SEQ ID NO:6:

50 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 9 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Gln Gln Arg Ser Ser Tyr Pro Tyr Thr
1 5 9

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 433 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
(B) LOCATION: -19...-1
(C) IDENTIFICATION METHOD: S
(A) NAME/KEY: domain
(B) LOCATION: 31..35
(C) IDENTIFICATION METHOD: S
(D) OTHER INFORMATION: hypervariable region 1
(A) NAME/KEY: domain
(B) LOCATION: 50..66
(C) IDENTIFICATION METHOD: S
(D) OTHER INFORMATION: hypervariable region 2
(A) NAME/KEY: domain
(B) LOCATION: 99..109
(C) IDENTIFICATION METHOD: S
(D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT 48
Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly
1 5 10 15

GTC CTC TCT GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG 96
Val Leu Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys
20 25 30

CCT GGG GCC TCA GTG AAG GTC TCC TGC AAG GCT TCC GGA TAC ACC TTC 144
Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe
35 40 45

ACT GAC TAC AAC ATG GAC TGG GTG CGA CAG GCC CCT GGA CAA GGG CTC 192
Thr Asp Tyr Asn Met Asp Trp Val Arg Gln Ala Pro Gly Gln Gly Leu
50 55 60

GAG TGG ATG GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC TAC AAC 240
Glu Trp Met Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn
65 70 75 80

CAG AAG TTC AAG AGC AAG GTC ACC ATT ACC GTA GAC ACA TCC ACG AGC 288
Gln Lys Phe Lys Ser Lys Val Thr Ile Thr Val Asp Thr Ser Thr Ser
85 90 95

ACA GCC TAC ATG GAG CTG CAC AGC CTG AGA TCT GAG GAC ACG GCC GTG 336
Thr Ala Tyr Met Glu Leu His Ser Leu Arg Ser Glu Asp Thr Ala Val
100 105 110

TAT TAC TGT GCG ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT GCT TAC 384
 Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr
 115 120 125

TGG GGC CAG GGA ACC CTG GTC ACC GTC TCC TCA GCC TCC ACC AAG GGC 442
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly
 130 135 140 144

C 443

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
- (B) LOCATION: -22..-1
- (C) IDENTIFICATION METHOD: S
- (A) NAME/KEY: domain
- (B) LOCATION: 24..33
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 1
- (A) NAME/KEY: domain
- (B) LOCATION: 49..55
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 2
- (A) NAME/KEY: domain
- (B) LOCATION: 86..96
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
 Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
 1 5 10 15

GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
 Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
 20 25 30

CTG AGC GCT AGC CCA GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
 Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45

TCA AGT GTA AGT TAC ATG CAC TGG TTC CAG CAG AAA CCA GGT AAG GCT 192
 Ser Ser Val Ser Tyr Met His Trp Phe Gln Gln Lys Pro Gly Lys Ala
 50 55 60

CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
 Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80

TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT CTC ACC ATC 288
 Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Leu Thr Ile
 85 90 95

EP 0 882 794 A2

AGC CGA CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
 Ser Arg Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110

5 AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
 115 120 125

CGT ACG 390
 Arg Thr
 130

10

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
- (B) LOCATION: -22..-1
- (C) IDENTIFICATION METHOD: S
- (A) NAME/KEY: domain
- (B) LOCATION: 24..33
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 1
- (A) NAME/KEY: domain
- (B) LOCATION: 49..55
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 2
- (A) NAME/KEY: domain
- (B) LOCATION: 86..96
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
 Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
 1 5 10 15

40 GTC ATA ATG TCC AGA GGA GAC ATC CAG CTG ACC CAG TCT CCA TCC TCC 96
 Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
 20 25 30

45 ATG TCT GCA TCT CCA GGA GAC AGA GTC ACC ATC ACT TGT AGT GCA AGT 144
 Met Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45

TCA AGT GTA AGT TAC ATG CAC TGG TTT CAG CAG AAA CCA GGG AAA TCA 192
 Ser Ser Val Ser Tyr Met His Trp Phe Gln Gln Lys Pro Gly Lys Ser
 50 55 60

50 CCT AAG CTC TGG ATC TAC TCA ACT TCA AAT TTA GCT TCT GGT GTG CCA 240
 Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80

55

EP 0 882 794 A2

TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT CTC ACC ATC 288
 Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Leu Thr Ile
 85 90 95

5 AGC AGC ATG CAG CCT GAA GAT TTT GCA ACT TAT TAC TGT CAG CAA AGG 336
 Ser Ser Met Gln Pro Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110

10 AGT AGT TAC CCG TAC ACG TTC GGC CAG GGG ACC AAG CTG GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gln Gly Thr Lys Leu Glu Ile Lys
 115 120 125

CGT ACG 390
 Arg Thr
 130

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 433 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
- (B) LOCATION: -19..-1
- (C) IDENTIFICATION METHOD: S
- (A) NAME/KEY: domain
- (B) LOCATION: 31..35
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 1
- (A) NAME/KEY: domain
- (B) LOCATION: 50..66
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 2
- (A) NAME/KEY: domain
- (B) LOCATION: 99..109
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

40 ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT 48
 Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly
 1 5 10 15

45 GTC CTC TCT GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG 96
 Val Leu Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys
 20 25 30

CCT GGG GCC TCA GTG AAG GTC TCC TGC AAG GCT TCC GGA TAC ACC TTC 144
 Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe
 35 40 45

50 ACT GAC TAC AAC ATG GAC TGG GTG AAG CAG AGC CCT GGA CAA GGG CTC 192
 Thr Asp Tyr Asn Met Asp Trp Val Lys Gln Ser Pro Gly Gln Gly Leu
 50 55 60

5 GAG TGG ATG GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC TAC AAC 240
 Glu Trp Met Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn
 65 70 75 80
 CAG AAG TTC AAG AGC AAG GTC ACC ATT ACC GTA GAC ACA TCC ACG AGC 288
 Gln Lys Phe Lys Ser Lys Val Thr Ile Thr Val Asp Thr Ser Thr Ser
 85 90 95
 10 ACA GCC TAC ATG GAG CTG CAC AGC CTG AGA TCT GAG GAC ACG GCC GTG 336
 Thr Ala Tyr Met Glu Leu His Ser Leu Arg Ser Glu Asp Thr Ala Val
 100 105 110
 TAT TAC TGT GCG ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT GCT TAC 384
 Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr
 115 120 125
 15 TGG GGC CAG GGA ACC CTG GTC ACC GTC TCC TCA GCC TCC ACC AAG GGC 432
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly
 130 135 140 144
 C 433

20 (2) INFORMATION FOR SEQ ID NO:11:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 25 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
 (ix) FEATURES:
 30 (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 35 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 40 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
 45 ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
 Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
 1 5 10 15
 GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
 Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Glu Ser Pro Ser Ser
 20 25 30
 50 CTG AGC GCT AGC CCA GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
 Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45

55

TCA AGT GTA AGT TAC ATG CAC TGG TTC CAG CAG AAA CCA GGT AAG GCT 192
 Ser Ser Val Ser Tyr Met His Trp Phe Gln Gln Lys Pro Gly Lys Ala
 50 55 60
 5 CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
 Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80
 GCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT CTC ACC ATC 288
 Ala Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Leu Thr Ile
 10 85 90 95
 AGC CGA CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGT CAG CAA AGG 336
 Ser Arg Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110
 15 AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTS GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
 115 120 125
 CGT ACG 390
 Arg Thr
 20 130

(2) INFORMATION FOR SEQ ID NO:12:

- 25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 32 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
 30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CACTCAGTGT TAACTGAGGA GCAGGTGAAT TC 32

35 (2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 40 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 40 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

AGCTGAATTC ACCTGCTCCT CAGTTAACAC TGAGTGGTAC 40

45 (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 50 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

55

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

5

AATTCGTACG GTGGCTGCAC C

21

(2) INFORMATION FOR SEQ ID NO:15:

10

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

15

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GGTGCAGCCA CCGTACG

17

20

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 26 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

30

CTCGCGACTA GTGGGCCCGC GGCCGC

26

(2) INFORMATION FOR SEQ ID NO:17:

35

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 34 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

AGCTGCGGCC GCGGGCCAC TAGTCGCGAG GTAC

34

45

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

50

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GTGGCGGCCG CTTGGGCCCC

20

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CGGGCCCAAG CGGCCGCCAC

20

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

CATGAATTCT TCGTACGGTT CGATAATCG ATACCG

36

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

CGGTATCGAT TTATCGAACC GTACGAAGAA TTCATGAGCT

40

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 35 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

CACGTTCCGA GGGGGGACCA AGCTGGAAAT AAAAC

35

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 35 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GTACGTTTTA TTTCCAGCTT GGTCCCCCT CCGAA

35

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 61 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

TCGACACCAG CAAGAACACA GCCTACCTGA GACTCAGCAG CGTGACAGCC GCCGACACCG 60

C

61

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 60 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

CCGGATACAC ATTCACTGAC TACAACATGG ACTGGGTGAG ACAGAGCCAT GGACGAGGTC 60

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 442 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -19..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 31..35
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 50..66
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 99..109
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

GGCCGCACC ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT 51
 Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr
 1 5 10

GCT GGT GTC CTC TCT CAG GTC CAA CTG CAG GAG AGC GGT CCA GGT CTT 99
 Ala Gly Val Leu Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu
 15 20 25 30

GTG AGG CCT AGC CAG ACC CTG AGC CTG ACC TGC ACC GTG TCC GGA TTC 147
 Val Arg Pro Ser Gln Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Phe
 35 40 45

ACC TTC AGC GAC TAC AAC ATG GAC TGG GTG AGA CAG CCA CCT GGA CGA 195
 Thr Phe Ser Asp Tyr Asn Met Asp Trp Val Arg Gln Pro Pro Gly Arg
 50 55 60

GGT CTC GAG TGG ATT GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC 243
 Gly Leu Glu Trp Ile Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly
 65 70 75

TAC AAC CAG AAG TTC AAG AGC AGA GTG ACA ATG CTG GTC GAC ACC AGC 291
 Tyr Asn Gln Lys Phe Lys Ser Arg Val Thr Met Leu Val Asp Thr Ser
 80 85 90

AAG AAC ACA GCC TAC CTG AGA CTC AGC AGC GTG ACA GCC GCC GAC ACC 339
 Lys Asn Thr Ala Tyr Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr
 95 100 105 110

GCG GTC TAT TAT TGT GCA ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT 387
 Ala Val Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe
 115 120 125

GCT TAC TGG GGT CAA GGT ACC ACC GTC ACA GTC TCC TCA GCC TCC ACC 435
 Ala Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Ala Ser Thr
 130 135 140

AAG GGC C 442
 Lys Gly
 144

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 442 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -19..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 31..35
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 50..66
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 99..109
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

```

GGCCGCACC ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT 51
Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr
   1             5             10

GCT GGT GTC CTC TCT CAG GTC CAA CTG CAG GAG AGC GGT CCA GGT CTT 99
Ala Gly Val Leu Ser Gln Val Gln Leu Gln Glu Ser Gly Pro Gly Leu
  15             20             25             30

GTG AGG CCT AGC CAG ACC CTG AGC CTG ACC TGC ACC GTG TCC GGA TAC 147
Val Arg Pro Ser Gln Thr Leu Ser Leu Thr Cys Thr Val Ser Gly Tyr
           35             40             45

ACC TTC ACT GAC TAC AAC ATG GAC TGG GTG AGA CAG AGC CAT GGA CGA 195
Thr Phe Thr Asp Tyr Asn Met Asp Trp Val Arg Gln Ser His Gly Arg
           50             55             60

GGT CTC GAG TGG ATT GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC 243
Gly Leu Glu Trp Ile Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly
           65             70             75

TAC AAC CAG AAG TTC AAG AGC AGA GTG ACA ATG CTG GTC GAC ACC AGC 291
Tyr Asn Gln Lys Phe Lys Ser Arg Val Thr Met Leu Val Asp Thr Ser
           80             85             90

AAG AAC CAG TTC AGC CTG AGA CTC AGC AGC GTG ACA GCC GCC GAC ACC 339
Lys Asn Gln Phe Ser Leu Arg Leu Ser Ser Val Thr Ala Ala Asp Thr
           95             100            105            110

GCG GTC TAT TAT TGT GCA ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT 387
Ala Val Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe
           115            120            125

GCT TAC TGG GGT CAA GGT ACC ACC GTC ACA GTC TCC TCA GCC TCC ACC 435
Ala Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Ala Ser Thr
           130            135            140

```


AAG GGC C
Lys Gly
144

442

5

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

10

- (A) LENGTH: 100 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

15

CAGGAAACAG CTATGACGCG GCCGCCACCA TGGGATGGAG CTGGATCTTT CTCTTCCTCC 60

TGTCAGGAAC TGCAGGTGTC CTCTCTGAGG TGCAGCTGGT 100

20

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:

25

- (A) LENGTH: 100 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

30

AGTCAGTGAA GGTGTATCCG GAAGCCTTGC AGGAGACCTT CACTGAGGCC CCAGGCTTCT 60

TCACCTCTGC TCCAGACTGC ACCAGCTGCA CCTCAGAGAG 100

35

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 100 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

CGGATACACC TTCACTGACT ACAACATGGA CTGGGTGCCA CAGGCCCTG GACAAGGGCT 60

45

CGAGTGGATG GGATATATTT ATCCTAACAA TGGTGGTACT 100

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

50

- (A) LENGTH: 94 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

55

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

AGCTCCATGT AGGCTGTGCT CGTGGATGTG TCTACGGTAA TGGTGACCTT GCTCTTGAAC 60
TTCTGGTTGT AGCCAGTACC ACCATTGTTA GGAT 94

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 96 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

AGCACAGCCT ACATGGAGCT GCACAGCCTG AGATCTGAGG ACACGGCCGT GTATTACTGT 60
GCGACCTACG GTCATTACTA CGGCTACATG TTTGCT 96

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 90 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GTTTTCCAG TCACGACGGG CCCTTGGTGG AGGCTGAGGA GACGGTGACC AGGGTTCCTT 60
GGCCCCAGTA AGCAAACATG TAGCCGTAGT 90

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 68 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

GTACTACTGC CAGCAAAGGA GTAGTTACCC GTACACGTTT GGCGGGGGGA CCAAGGTGGA 60
AATCAAAC 68

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ACTCTGTCAC CTGGGCTAGC GCTCA

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

TGAGCGCTAG CCCAGGTGAC AGAGT

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA	48
Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser	
1 5 10 15	
GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC	96
Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser	
20 25 30	

EP 0 882 794 A2

CTG AGC GCT AGC CCA GGT GAG AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
 Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45

5 TCA AGT GTA AGT TAC ATG CAC TGG TAT CAG CAG AAA CCA GGT AAG GCT 192
 Ser Ser Val Ser Tyr Met His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
 50 55 60

10 CCA AAG CTT CTG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
 Pro Lys Leu Leu Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80

TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA GAC TTC ACC TTC ACC ATC 288
 Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Phe Thr Ile
 85 90 95

15 AGC AGC CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
 Ser Ser Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110

20 AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
 115 120 125

CGT ACG 390
 Arg Thr
 130

25

(2) INFORMATION FOR SEQ ID NO:38:

- 30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- 35 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

GTGCTGTAGA TCCAAAGCTT TGGAG 25

40 (2) INFORMATION FOR SEQ ID NO:39:

- 45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

50 CTCCAAAGCT TTGGATCTAC AGCAC 25

55

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

```

25  ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
    Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
      1             5             10             15

    GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
    Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
      20             25             30

    CTG AGC GCT AGC GTG GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
    Leu Ser Ala Ser Val Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
      35             40             45

    TCA AGT GTA AGT TAC ATG CAC TGG TAT CAG CAG AAA CCA GGT AAG GCT 192
    Ser Ser Val Ser Tyr Met His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
      50             55             60

    CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
    Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
      65             70             75             80

    TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA GAC TTC ACC TTC ACC ATC 288
    Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Phe Thr Ile
      85             90             95

    AGC AGC CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
    Ser Ser Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
      100            105            110

    AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
    Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
      115            120            125

50  CGT ACG
    Arg Thr
      130

55

```

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

ACGTAGCAGC ATCTTCAGCC TGGAG

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

CTCCAGGCTG AAGATGCTGC TACGT

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
- (B) LOCATION: -22...-1
- (C) IDENTIFICATION METHOD: S
- (A) NAME/KEY: domain
- (B) LOCATION: 24..33
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 1
- (A) NAME/KEY: domain
- (B) LOCATION: 49..55
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 2
- (A) NAME/KEY: domain
- (B) LOCATION: 86..96
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
 Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
 1 5 10 15

5 GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
 Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
 20 25 30
 CTG AGC GCT AGC GTG GGT GAC AGA GTG ACC ATC ACG TGC AGT GCT AGC 144
 Leu Ser Ala Ser Val Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45
 TCA AGT GTA AGT TAC ATG CAC TGG TAT CAG CAG AAA CCA GGT AAG GCT 192
 Ser Ser Val Ser Tyr Met His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
 50 55 60
 CCA AAG CTT CTG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTC CCA 240
 Pro Lys Leu Leu Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80
 TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA GAC TTC ACC TTC AAT ATC 288
 Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Phe Thr Ile
 85 90 95
 AGC AGC CTC CAG GCT GAA GAT GCT GCT ACA TAC TAC TGC CAG CAA AGG 336
 Ser Ser Leu Gln Ala Glu Asp Ala Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110
 AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
 115 120 125
 25 CGT ACG 390
 Arg Thr
 130

(2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

ATGGTGAAAG AGTAAGATGT ACCGC

25

(2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

GCGGTACATC TTACTCTTTC ACCAT

25

(2) INFORMATION FOR SEQ ID NO:46:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

```

25  ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
    Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
      1             5             10             15

30  GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
    Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
      20             25             30

    CTG AGC GCT AGC CCA GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
    Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
      35             40             45

35  TCA AGT GTA AGT TAC ATG CAC TGG TAT CAG CAG AAA CCA GGT AAG GCT 192
    Ser Ser Val Ser Tyr Met His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
      50             55             60

    CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
    Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
      65             70             75             80

40  TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT TTC ACC ATC 288
    Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Phe Thr Ile
      85             90             95

45  AGC AGC CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
    Ser Ser Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
      100            105            110

    AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
    Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
      115            120            125

50  CGT ACG
    Arg Thr
      130

55

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(2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 40 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

TCTGGCTCCA TTCGGCTGAT GGTGAAAGAG TAAGATGTAC

40

(2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 40 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

GTACATCTTA CTCTTTCACC ATCAGCCGAA TGGAGCCAGA

40

(2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
 Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
 1 5 10 15

5 GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
 Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
 20 25 30
 10 CTG AGC GCT AGC CCA GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
 Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45
 15 TCA AGT GTA AGT TAC ATG CAC TGG TAT CAG CAG AAA CCA GGT AAG GCT 192
 Ser Ser Val Ser Tyr Met His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
 50 55 60
 20 CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
 Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80
 25 TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT TTC ACC ATC 288
 Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Phe Thr Ile
 85 90 95
 30 AGC CGA ATG GAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
 Ser Arg Met Glu Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110
 35 AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
 115 120 125
 40 CGT ACG 390
 Arg Thr
 130

(2) INFORMATION FOR SEQ ID NO:50:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

TTCTGCTGGA ACCAGTGCAT

20

(2) INFORMATION FOR SEQ ID NO:51:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

ATGCACTGGT TCCAGCAGAA

20

(2) INFORMATION FOR SEQ ID NO:52:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

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25  ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
    Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
      1           5           10           15

    GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
    Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
      20           25           30

    CTG AGC GCT AGC CCA GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
    Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
      35           40           45

    TCA AGT GTA AGT TAC ATG CAC TGG TTC CAG CAG AAA CCA GGT AAG GCT 192
    Ser Ser Val Ser Tyr Met His Trp Phe Gln Gln Lys Pro Gly Lys Ala
      50           55           60

    CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
    Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
      65           70           75           80

    TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT TTC ACC ATC 288
    Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Phe Thr Ile
      85           90           95

    AGC AGC CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
    Ser Ser Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
      100          105          110

    AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
    Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
      115          120          125

50  CGT ACG
    Arg Thr
      130

55

```

(2) INFORMATION FOR SEQ ID NO:53:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

TGGAGTCGGC TGATGGTGAG AGAGT

25

(2) INFORMATION FOR SEQ ID NO:54:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

ACTCTCTCAC CATCAGCCGA CTCCA

25

(2) INFORMATION FOR SEQ ID NO:55:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 390 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -22..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 24..33
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 49..55
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 86..96
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

ATG CAT TTT CAA GTG CAG ATT TTC AGC TTC CTG CTA ATC AGT GCC TCA 48
 Met His Phe Gln Val Gln Ile Phe Ser Phe Leu Leu Ile Ser Ala Ser
 1 5 10 15

5 GTC ATA ATG TCC AGA GGA GAT ATC CAG CTG ACC CAG AGC CCA AGC AGC 96
 Val Ile Met Ser Arg Gly Asp Ile Gln Leu Thr Gln Ser Pro Ser Ser
 20 25 30
 CTG AGC GCT AGC CCA GGT GAC AGA GTG ACC ATC ACG TGC AGT GCC AGC 144
 Leu Ser Ala Ser Pro Gly Asp Arg Val Thr Ile Thr Cys Ser Ala Ser
 35 40 45
 TCA AGT GTA AGT TAC ATG CAC TGG TAT CAG CAG AAA CCA GGT AAG GCT 192
 Ser Ser Val Ser Tyr Met His Trp Tyr Gln Gln Lys Pro Gly Lys Ala
 50 55 60
 CCA AAG CTT TGG ATC TAC AGC ACA TCC AAC CTG GCT TCT GGT GTG CCA 240
 Pro Lys Leu Trp Ile Tyr Ser Thr Ser Asn Leu Ala Ser Gly Val Pro
 65 70 75 80
 TCT AGA TTC AGC GGT AGC GGT AGC GGT ACA TCT TAC TCT CTC ACC ATC 288
 Ser Arg Phe Ser Gly Ser Gly Ser Gly Thr Ser Tyr Ser Leu Thr Ile
 85 90 95
 AGC CGA CTC CAG CCA GAG GAC ATC GCT ACA TAC TAC TGC CAG CAA AGG 336
 Ser Arg Leu Gln Pro Glu Asp Ile Ala Thr Tyr Tyr Cys Gln Gln Arg
 100 105 110
 AGT AGT TAC CCG TAC ACG TTC GGC GGG GGG ACC AAG GTG GAA ATC AAA 384
 Ser Ser Tyr Pro Tyr Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys
 115 120 125
 25 CGT ACG 390
 Arg Thr
 130

30 (2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 94 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

40 CAGGAAACAG CTATGACGAA TTCCACCATG CATTTTCAAG TGCAGATTTT CAGCTTCCTG 60
 CTAATCAGTG CCTCAGTCAT AATGTCCAGA GGAG 94

(2) INFORMATION FOR SEQ ID NO:57:

- 45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 88 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

5 ACAAGTGATG GTGACTCTGT CTCCTGGAGA TGCAGACATG GAGGATGGAG ACTGGGTCAG 60
CTGGATGTCT CCTCTGGACA TTATGACT 88

(2) INFORMATION FOR SEQ ID NO:58:

10 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 92 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

20 ACAGAGTCAC CATCACTTGT AGTGCAAGTT CAAGTGTAAG TTACATGCAC TGGTTTCAGC 60
AGAAACCAGG GAAATCACCT AAGCTCTGGA TC 92

(2) INFORMATION FOR SEQ ID NO:59:

25 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 87 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

30 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

35 AAGATGTACC GCTACCGCTA CCGCTGAATC TAGATGGCAC ACCAGAAGCT AAATTTGAAG 60
TTGAGTAGAT CCAGAGCTTA GGTGATT 87

(2) INFORMATION FOR SEQ ID NO:60:

40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 89 base pairs
(B) TYPE: nucleic acid
(D) TOPOLOGY: linear

45 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

50 TAGCGGTAGC GGTACATCTT ACTCTCTCAC CATCAGCAGC ATGCAGCCTG AAGATTTTGC 60
AACTTATTAC TGTCAGCAAA GGAGTAGTT 89

(2) INFORMATION FOR SEQ ID NO:61:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 84 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:

GTTTCCCAG TCACGACCGT ACGTTTGATT TCCAGCTTGG TCCCCTGGCC GAACGTGTAC 60
 GGGTAACTAC TCCTTTGCTG ACAG 84

(2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 35 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

ACTCGAGGCT CTTTCCAGGG CTCTGCTTCA CCCAG 35

(2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 35 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

CTGGGTGAAG CAGAGCCCTG GAAAGAGCCT CGAGT 35

(2) INFORMATION FOR SEQ ID NO:64:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 433 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -19..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 31..35
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 50..66
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 99..109
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT	48
Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly	
1 5 10 15	
GTC CTC TCT GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG	96
Val Leu Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys	
20 25 30	
CCT GGG GCC TCA GTG AAG GTC TCC TGC AAG GCT TCC GGA TAC ACC TTC	144
Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe	
35 40 45	
ACT GAC TAC AAC ATG GAC TGG GTG AAG CAG AGC CCT GGA AAG AGC CTC	192
Thr Asp Tyr Asn Met Asp Trp Val Lys Gln Ser Pro Gly Lys Ser Leu	
50 55 60	
GAG TGG ATG GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC TAC AAC	240
Glu Trp Met Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn	
65 70 75 80	
CAG AAG TTC AAG AGC AAG GTC ACC ATT ACC GTA GAC ACA TCC ACG AGC	288
Gln Lys Phe Lys Ser Lys Val Thr Ile Thr Val Asp Thr Ser Thr Ser	
85 90 95	
ACA GCC TAC ATG GAG CTG CAC AGC CTG AGA TCT GAG GAC ACG GCC GTG	336
Thr Ala Tyr Met Glu Leu His Ser Leu Arg Ser Glu Asp Thr Ala Val	
100 105 110	
TAT TAC TGT GCG ACC TAC GGT CAT TAC TAC GGC TAC ATG TTI GCT TAC	384
Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr	
115 120 125	
TGG GGC CAG GGA ACC CTG GTC ACC GTC TCC TCA GCC TCC ACC AAG GGC	432
Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly	
130 135 140 144	
C	433

(2) INFORMATION FOR SEQ ID NO:65:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

TGTCCAGGGC TCTGCTTCAC CCAG 24

(2) INFORMATION FOR SEQ ID NO:66:

10 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:

CTGGGTGAAG CAGAGCCCTG GACA 24

(2) INFORMATION FOR SEQ ID NO:67:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

25 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

30 TCTACGGTCA AGGTGGCCTT GCTCT 25

(2) INFORMATION FOR SEQ ID NO:68:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

40 (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:

AGAGCAAGGC CACCTTGACC GTAGA 25

(2) INFORMATION FOR SEQ ID NO:69:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 433 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

50 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

55

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -19..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 31..35
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 50..66
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 99..109
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

20	ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT	48
	Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly	
	1 5 10 15	
25	GTC CTC TCT GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG	96
	Val Leu Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys	
	20 25 30	
30	CCT GGG GCC TCA GTG AAG GTC TCC TGC AAG GCT TCC GGA TAC ACC TTC	144
	Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe	
	35 40 45	
35	ACT GAC TAC AAC ATG GAC TGG GTG CGA CAG GCC CCT GGA CAA GGG CTC	192
	Thr Asp Tyr Asn Met Asp Trp Val Arg Gln Ala Pro Gly Gln Gly Leu	
	50 55 60	
40	GAG TGG ATG GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC TAC AAC	240
	Glu Trp Met Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn	
	65 70 75 80	
45	CAG AAG TTC AAG AGC AAG GCC ACC TTG ACC GTA GAC ACA TCC ACG AGC	288
	Gln Lys Phe Lys Ser Lys Ala Thr Leu Thr Val Asp Thr Ser Thr Ser	
	85 90 95	
50	ACA GCC TAC ATG GAG CTG CAC AGC CTG AGA TCT GAG GAC ACG GCC GTG	336
	Thr Ala Tyr Met Glu Leu His Ser Leu Arg Ser Glu Asp Thr Ala Val	
	100 105 110	
55	TAT TAC TGT GCG ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT GCT TAC	384
	Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr	
	115 120 125	
	TGG GGC CAG GGA ACC CTG GTC ACC GTC TCC TCA GCC TCC ACC AAG GGC	432
	Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly	
	130 135 140 144	
	C	433

(2) INFORMATION FOR SEQ ID NO:70:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 433 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

- (A) NAME/KEY: sig peptide
 (B) LOCATION: -19..-1
 (C) IDENTIFICATION METHOD: S
 (A) NAME/KEY: domain
 (B) LOCATION: 31..35
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 1
 (A) NAME/KEY: domain
 (B) LOCATION: 50..66
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 2
 (A) NAME/KEY: domain
 (B) LOCATION: 99..109
 (C) IDENTIFICATION METHOD: S
 (D) OTHER INFORMATION: hypervariable region 3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:

ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT	48
Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly	
1 5 10 15	
GTC CTC TCT GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG	96
Val Leu Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys	
20 25 30	
CCT GGG GCC TCA GTG AAG GTC TCC TGC AAG GCT TCC GGA TAC ACC TTC	144
Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe	
35 40 45	
ACT GAC TAC AAC ATG GAC TGG GTG AAG CAG AGC CCT GGA AAG AGC CTC	192
Thr Asp Tyr Asn Met Asp Trp Val Lys Gln Ser Pro Gly Lys Ser Leu	
50 55 60	
GAG TGG ATG GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC TAC AAC	240
Glu Trp Met Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn	
65 70 75 80	
CAG AAG TTC AAG AGC AAG GCC ACC TTG ACC GTA GAC ACA TCC ACG AGC	288
Gln Lys Phe Lys Ser Lys Ala Thr Leu Thr Val Asp Thr Ser Thr Ser	
85 90 95	
ACA GCC TAC ATG GAG CTG CAC AGC CTG AGA TCT GAG GAC ACG GCC GTG	336
Thr Ala Tyr Met Glu Leu His Ser Leu Arg Ser Glu Asp Thr Ala Val	
100 105 110	
TAT TAC TGT GCG ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT GCT TAC	384
Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr	
115 120 125	

TGG GGC CAG GGA ACC CTG GTC ACC GTC TCC TCA GCC TCC ACC AAG GGC 432
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly
 130 135 140 144

5

C

433

(2) INFORMATION FOR SEQ ID NO:71:

10

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 433 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

15

(ii) MOLECULE TYPE: other nucleic acid, synthetic DNA

(ix) FEATURES:

20

- (A) NAME/KEY: sig peptide
- (B) LOCATION: -19..-1
- (C) IDENTIFICATION METHOD: S
- (A) NAME/KEY: domain
- (B) LOCATION: 31..35
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 1
- (A) NAME/KEY: domain
- (B) LOCATION: 50..66
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 2
- (A) NAME/KEY: domain
- (B) LOCATION: 99..109
- (C) IDENTIFICATION METHOD: S
- (D) OTHER INFORMATION: hypervariable region 3

25

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:

ATG GGA TGG AGC TGG ATC TTT CTC TTC CTC CTG TCA GGA ACT GCA GGT 48
 Met Gly Trp Ser Trp Ile Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly
 1 5 10 15

35

GTC CTC TCT GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAG AAG 96
 Val Leu Ser Glu Val Gln Leu Val Gln Ser Gly Ala Glu Val Lys Lys
 20 25 30

40

CCT GGG GCC TCA GTG AAG GTC TCC TGC AAG GCT TCC GGA TAC ACC TTC 144
 Pro Gly Ala Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Thr Phe
 35 40 45

45

ACT GAC TAC AAC ATG GAC TGG GTG AAG CAG AGC CCT GGA CAA GGG CTC 192
 Thr Asp Tyr Asn Met Asp Trp Val Lys Gln Ser Pro Gly Gln Gly Leu
 50 55 60

GAG TGG ATG GGA TAT ATT TAT CCT AAC AAT GGT GGT ACT GGC TAC AAC 240
 Glu Trp Met Gly Tyr Ile Tyr Pro Asn Asn Gly Gly Thr Gly Tyr Asn
 65 70 75 80

CAG AAG TTC AAG AGC AAG GCC ACC TTG ACC GTA GAC ACA TCC ACG AGC 288
 Gln Lys Phe Lys Ser Lys Ala Thr Leu Thr Val Asp Thr Ser Thr Ser
 85 90 95

50

ACA GCC TAC ATG GAG CTG CAC AGC CTG AGA TCT GAG GAC ACG GCC GTG 336
 Thr Ala Tyr Met Glu Leu His Ser Leu Arg Ser Glu Asp Thr Ala Val
 100 105 110

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5 TAT TAC TGT GCG ACC TAC GGT CAT TAC TAC GGC TAC ATG TTT GCT TAC 384
 Tyr Tyr Cys Ala Thr Tyr Gly His Tyr Tyr Gly Tyr Met Phe Ala Tyr
 115 120 125

10 TGG GGC CAG GGA ACC CTG GTC ACC GTC TCC TCA GCC TCC ACC AAG GGC 432
 Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser Ala Ser Thr Lys Gly
 130 135 140 144

C 433

(2) INFORMATION FOR SEQ ID NO:72:

- 15 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
- 20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

TGAATCTAGC TGGCACACCA 20

25 (2) INFORMATION FOR SEQ ID NO:73:

- 30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid, synthetic DNA
- 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

TGGTGTGCCA GCTAGATTCA 20

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Claims

- 45 1. A human CDR-grafted antibody which specifically reacts with ganglioside GM₂, wherein said antibody comprises CDR 1, CDR 2 and CDR 3 of heavy chain (H chain) variable region (V region) comprising amino acid sequences of SEQ ID NO:1, SEQ ID NO:2 and SEQ ID NO:3 or functional equivalents thereof, and CDR 1, CDR 2 and CDR 3 of light chain (L chain) V region comprising amino acid sequences of SEQ ID NO:4, SEQ ID NO:5 and SEQ ID NO:6 or functional equivalents thereof, and wherein at least one of the frameworks (FR) of said H chain and L chain V regions comprises an amino acid sequence selected from common sequences (HMHCS; human most homologous consensus sequence) derived from human antibody subgroups.
- 50
2. The human CDR-grafted antibody according to claim 1, wherein said FR is an amino acid sequence of an FR of an HMHCS having a high homology with an FR of a monoclonal antibody originated from nonhuman animal which specifically reacts with ganglioside GM₂.
- 55
3. The human CDR-grafted antibody according to claim 1 or 2, wherein said FR of H chain or L chain V region of the human CDR-grafted antibody comprises an amino acid sequence in which at least one amino acid is replaced by

an other amino acid, and wherein said antibody has antigen-binding activity and binding specificity comparable to those of a human chimeric antibody having a V region of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂.

- 5 4. The human CDR-grafted antibody according to any one of claims 1 to 3, wherein said FR of H chain or L chain V region of the human CDR-grafted antibody comprises an amino acid sequence in which at least one amino acid is replaced by an other amino acid, and wherein said antibody has antibody dependent cell mediated cytotoxicity (ADCC) comparable to that of a human chimeric antibody having a V region of a monoclonal antibody derived from nonhuman animal which specifically reacts with ganglioside GM₂.
- 10 5. The human CDR-grafted antibody according to any one of claims 1 to 4, wherein said FR of H chain or L chain V region of the human CDR-grafted antibody comprises an amino acid sequence in which at least one amino acid is replaced by an other amino acid, and wherein said antibody has complement dependent cytotoxicity (CDC) comparable to that of a human chimeric antibody having a V region of a monoclonal antibody originated from nonhuman animal which specifically reacts with ganglioside GM₂.
- 15 6. The human CDR-grafted antibody according to any one of claims 3 to 5, wherein said other amino acid is selected from amino acids in a position corresponding to the FR of the monoclonal antibody derived from nonhuman animal.
- 20 7. The human CDR-grafted antibody according to any one of claims 3 to 6, wherein at least one amino acid of positions 38, 40, 67, 72, 84 and 98 in the FR of H chain V region and positions 4, 11, 15, 35, 42, 46, 59, 69, 70, 71, 72, 76, 77 and 103 in the FR of L chain V region is replaced by an other amino acid.
- 25 8. The human CDR-grafted antibody according to any one of claims 1 to 7, wherein said H chain C region of the antibody is derived from an antibody belonging to the human antibody IgG class.
9. The human CDR-grafted antibody according to any one of claims 1 to 4 and 6 to 8, which is KM8966 comprising the H chain V region of the antibody having an amino acid sequence of SEQ ID NO:7 and the L chain V region of the antibody having an amino acid sequence of SEQ ID NO:8.
- 30 10. The human CDR-grafted antibody according to any one of claims 1 to 4 and 6 to 8, which is KM8967 comprising the H chain V region of the antibody having an amino acid sequence of SEQ ID NO:7 and the L chain V region of the antibody having an amino acid sequence of SEQ ID NO:9.
- 35 11. The human CDR-grafted antibody and according to any one of claims 1 to 4 and 6 to 8, which is KM8970 comprising the H chain V region of the antibody having an amino acid sequence of SEQ ID NO:10 and the L chain V region of the antibody having an amino acid sequence of SEQ ID NO:8.
- 40 12. The human CDR-grafted antibody according to any one of claims 1 to 8, which is KM8969 comprising the H chain V region of the antibody having an amino acid sequence of SEQ ID NO:10 and the L chain V region of the antibody having an amino acid sequence of SEQ ID NO:11.
- 45 13. A DNA fragment encoding an amino acid sequence of the H chain V region and L chain V region of the antibody according to any one of claims 1 to 12.
14. A recombinant vector comprising the DNA fragment according to claim 13 or a part thereof.
15. The recombinant vector according to claim 14, which is derived from a tandem cassette vector, pKANTEX 93, for expressing a human chimeric antibody and a human CDR-grafted antibody.
- 50 16. A transformant comprising the recombinant vector according to claim 14 or 15.
17. A transformant cell line KM8966 (FERM BP-5105), which produces the antibody KM8966 according to claim 9.
- 55 18. A transformant cell line KM8967 (FERM BP-5106), which produces the antibody KM8967 according to claim 10.
19. A transformant cell line KM8970 (FERM BP-5528), which produces the antibody KM8970 according to claim 11.

20. A transformant cell line KM8969 (FERM BP-5527), which produces the antibody KM8969 according to claim 12.

21. A method for producing the antibodies according to any one of claims 1 to 12 using said transformant according to any one of claims 17 to 20.

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22. An anti-tumor agent comprising the antibody of any one of claims 1 to 12 as an active ingredient.

23. A diagnostic agent for cancer comprising the antibody of any one of claims 1 to 12 as an active ingredient.

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FIG. 1

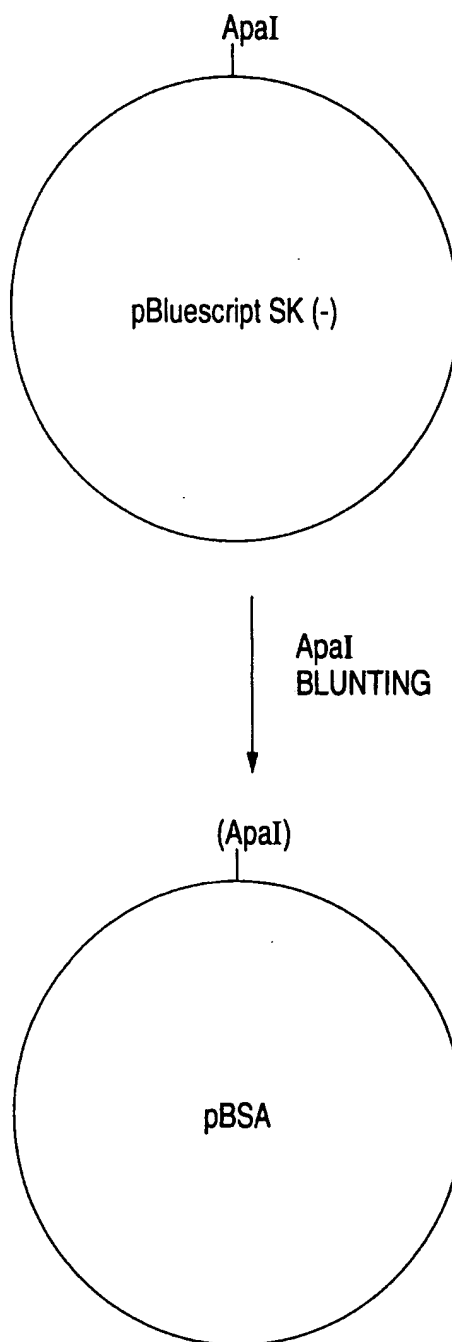


FIG. 2

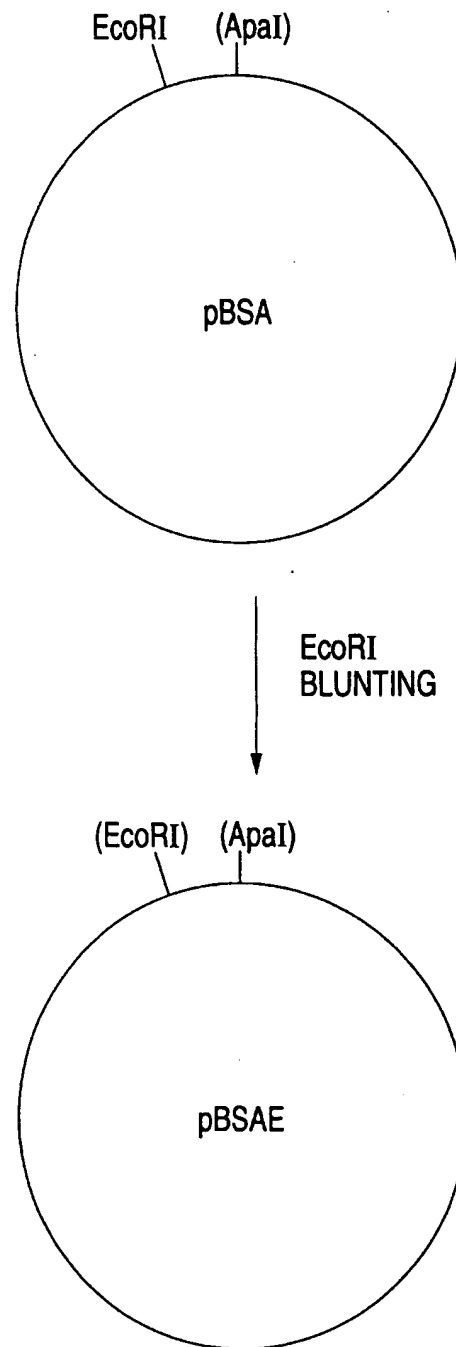


FIG. 3

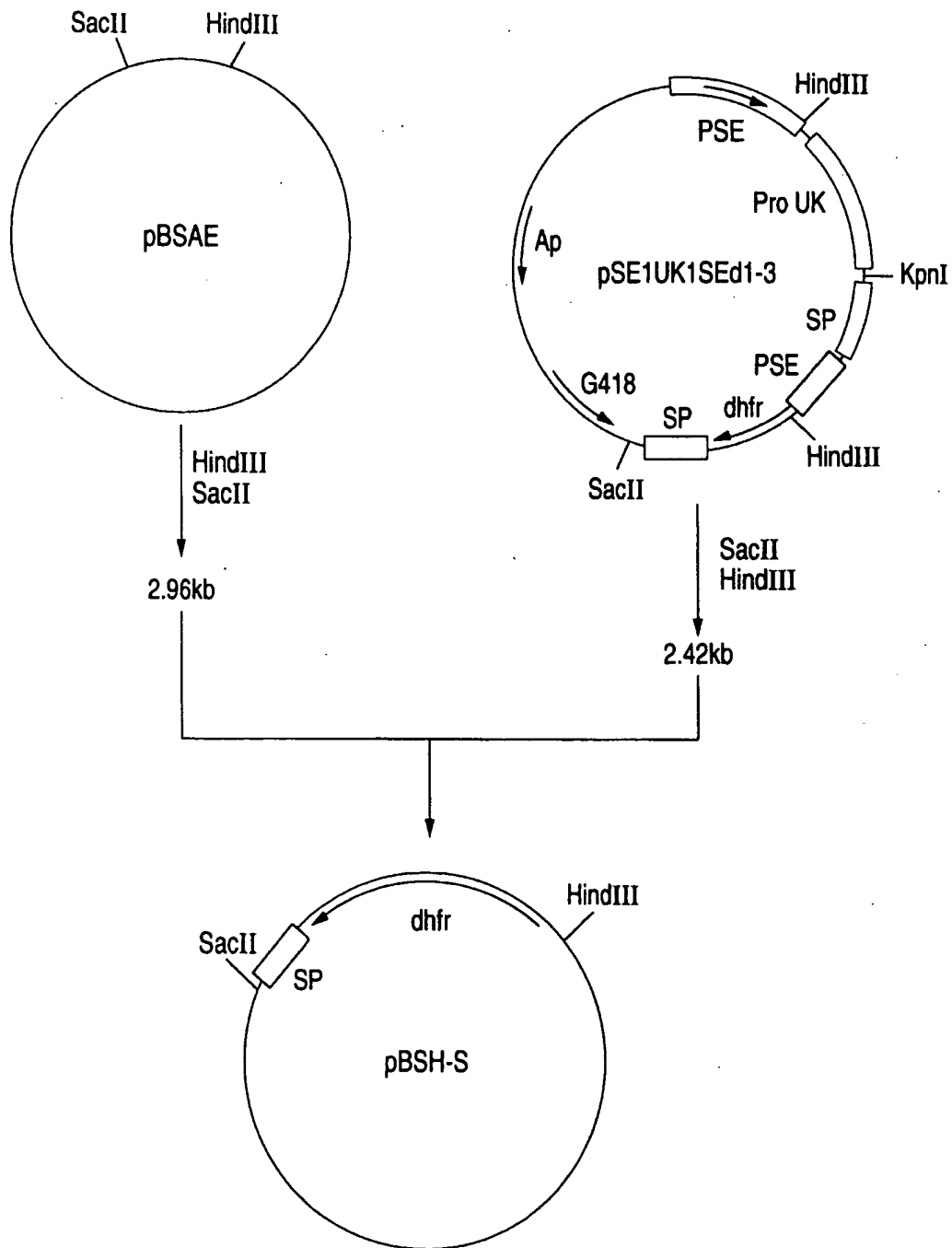


FIG. 4

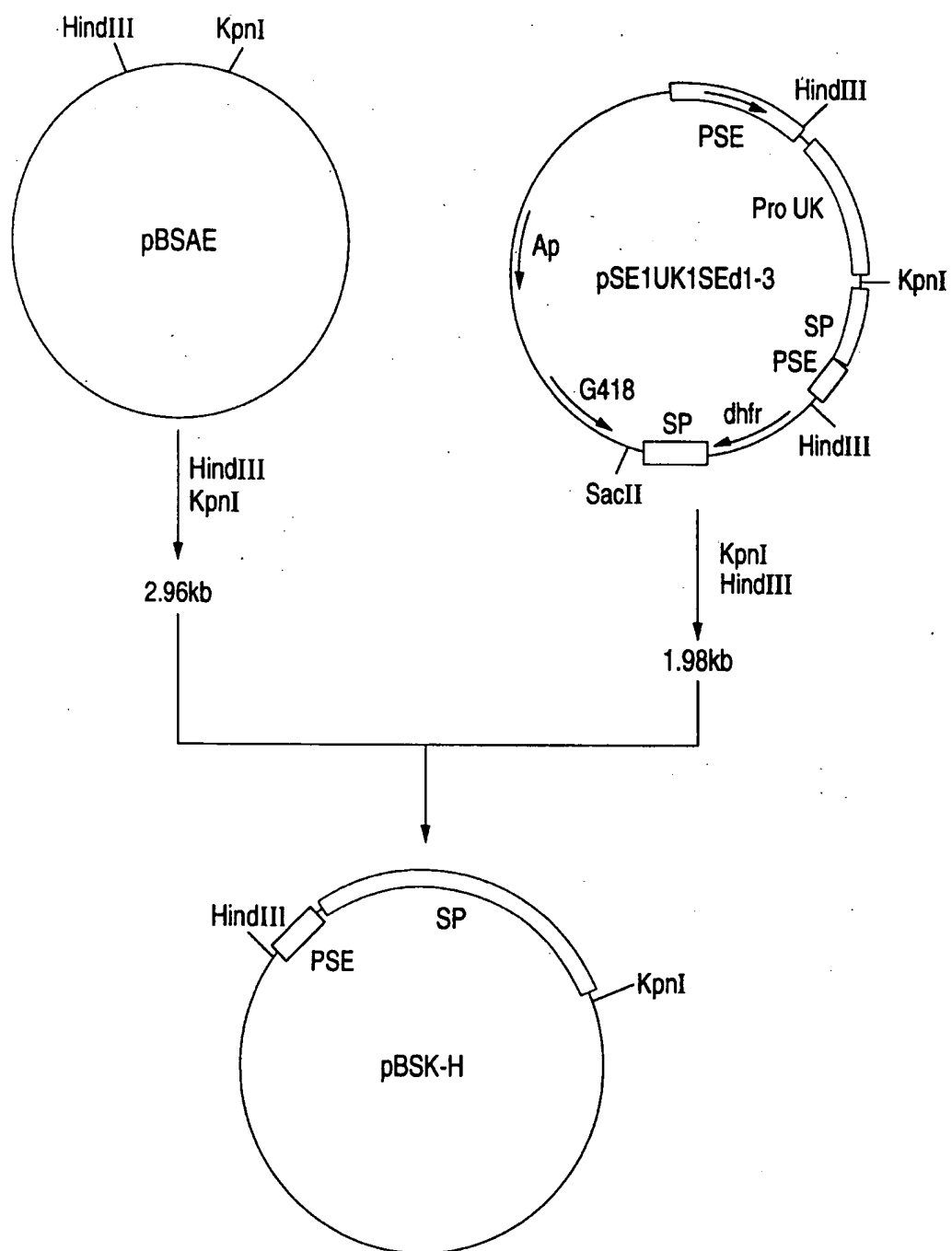


FIG. 5

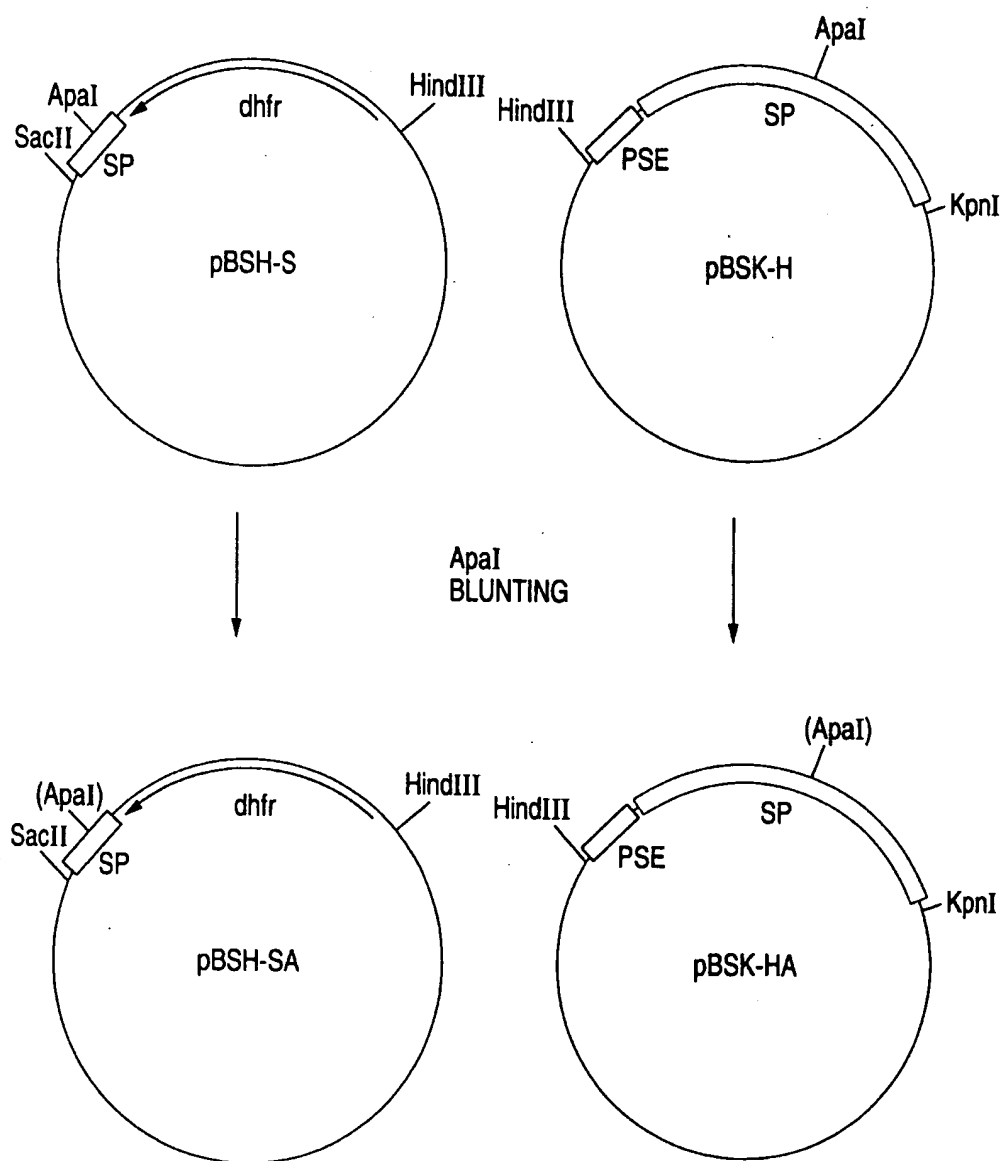


FIG. 6

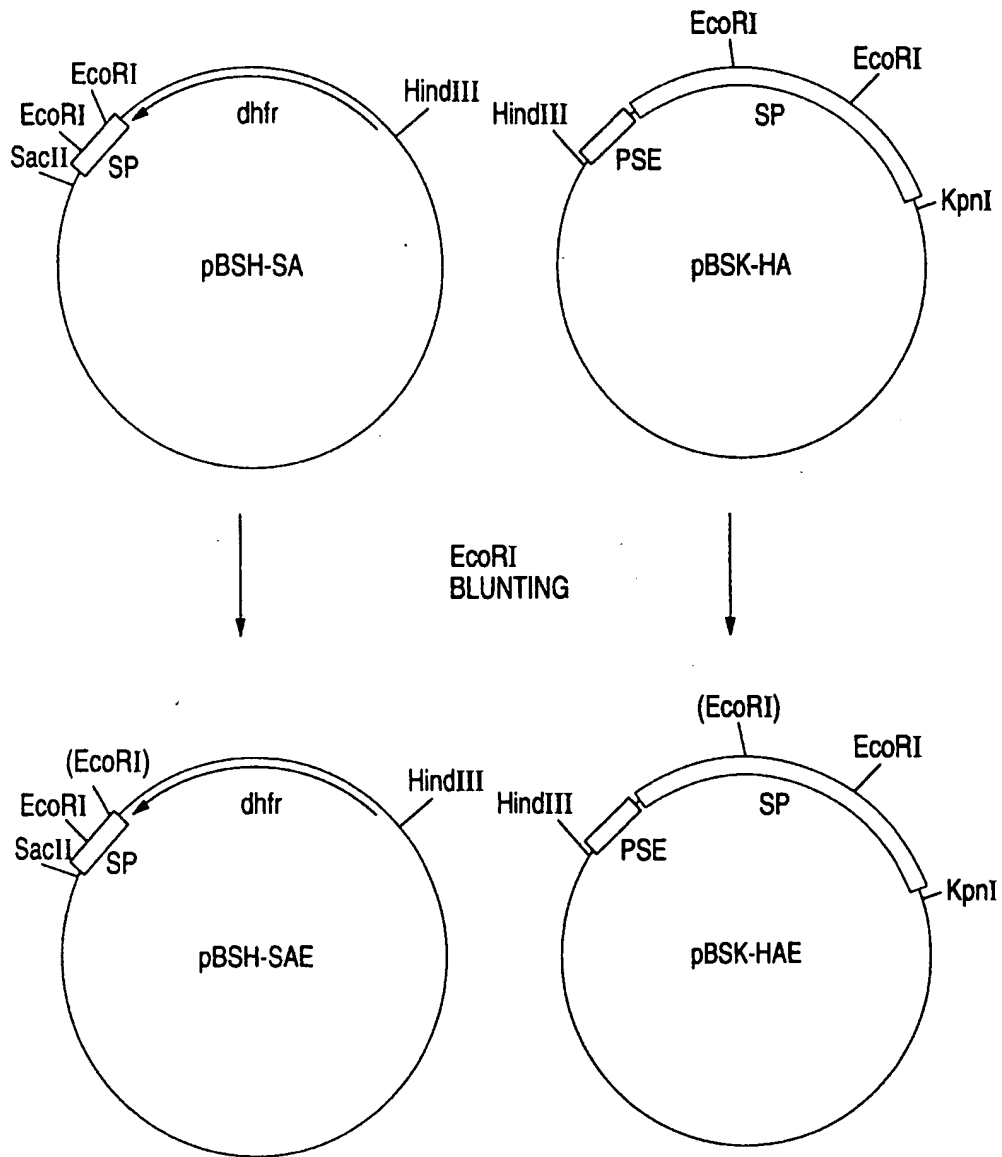


FIG. 7

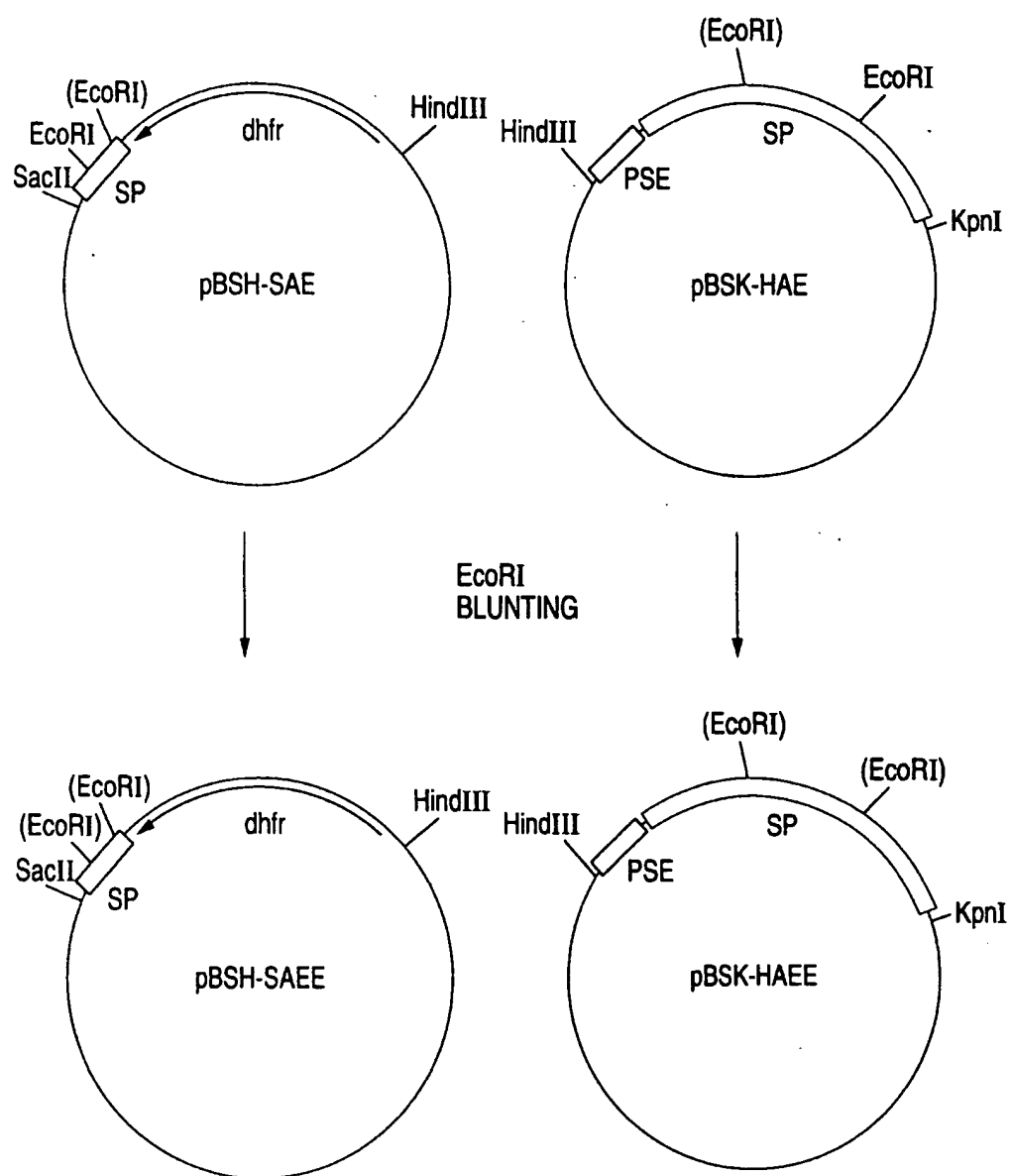


FIG. 8

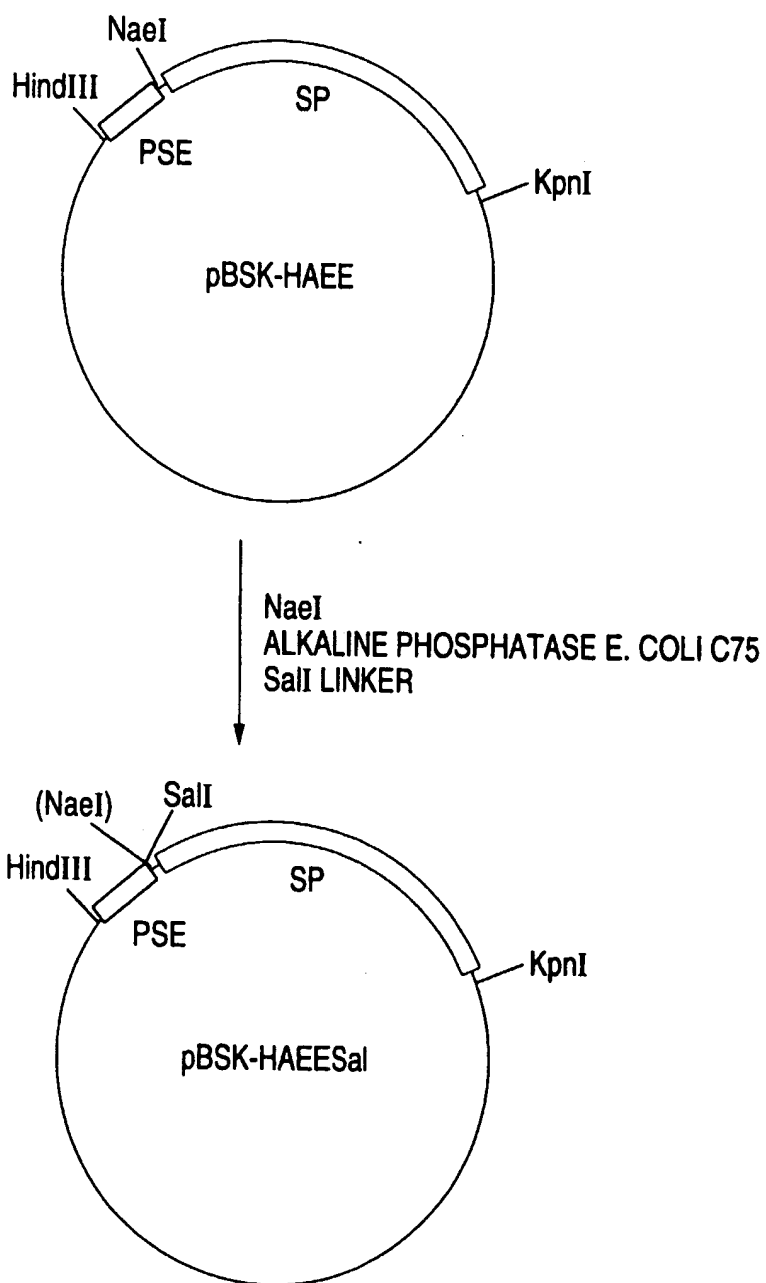


FIG. 9

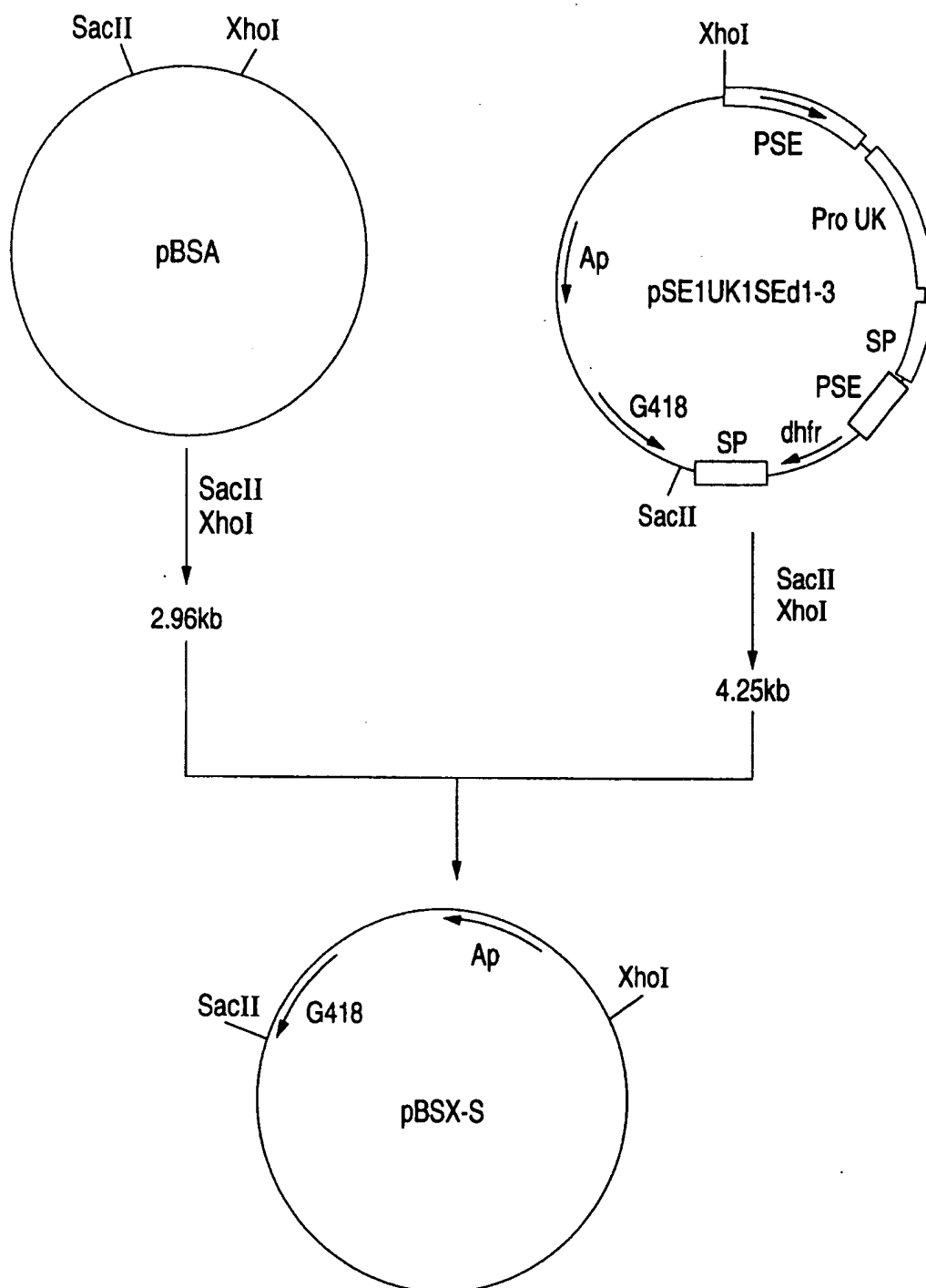


FIG. 10

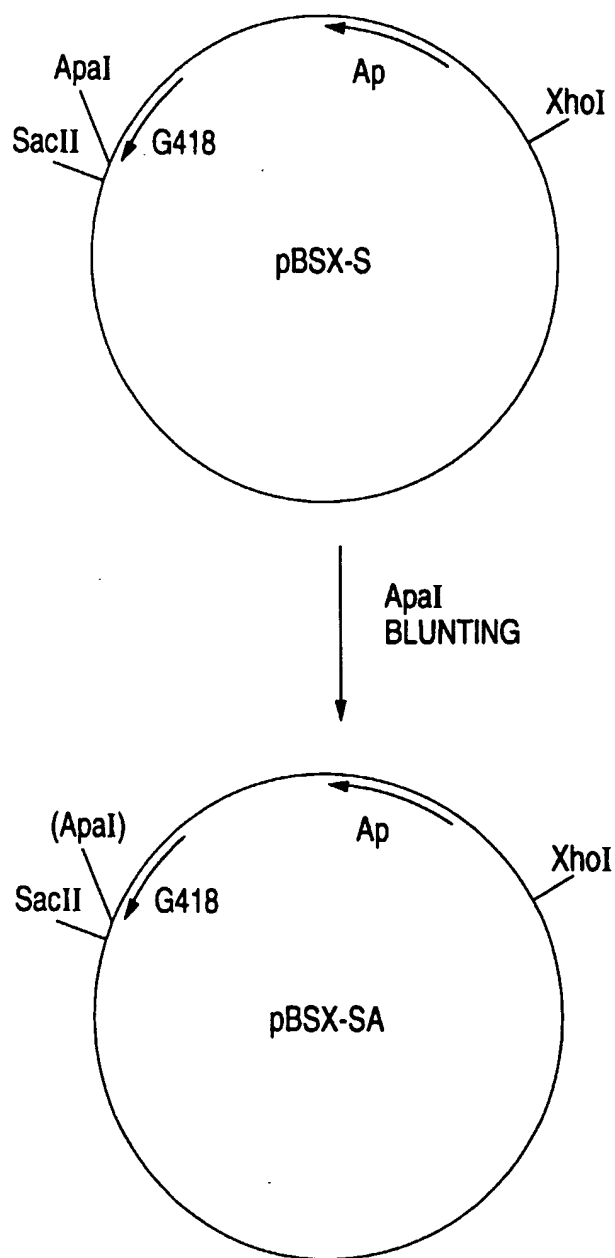


FIG. 11

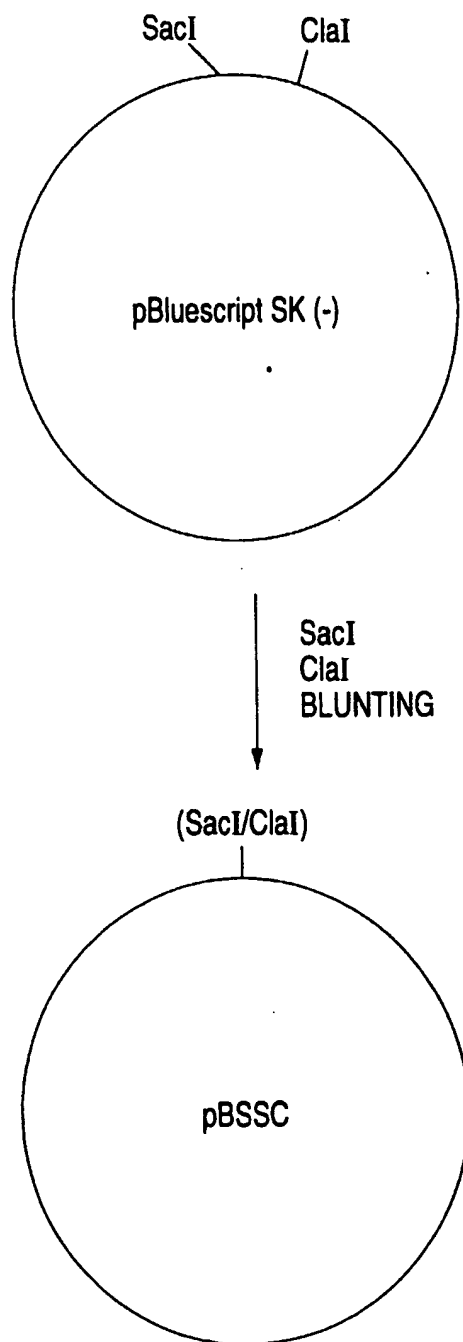


FIG. 12

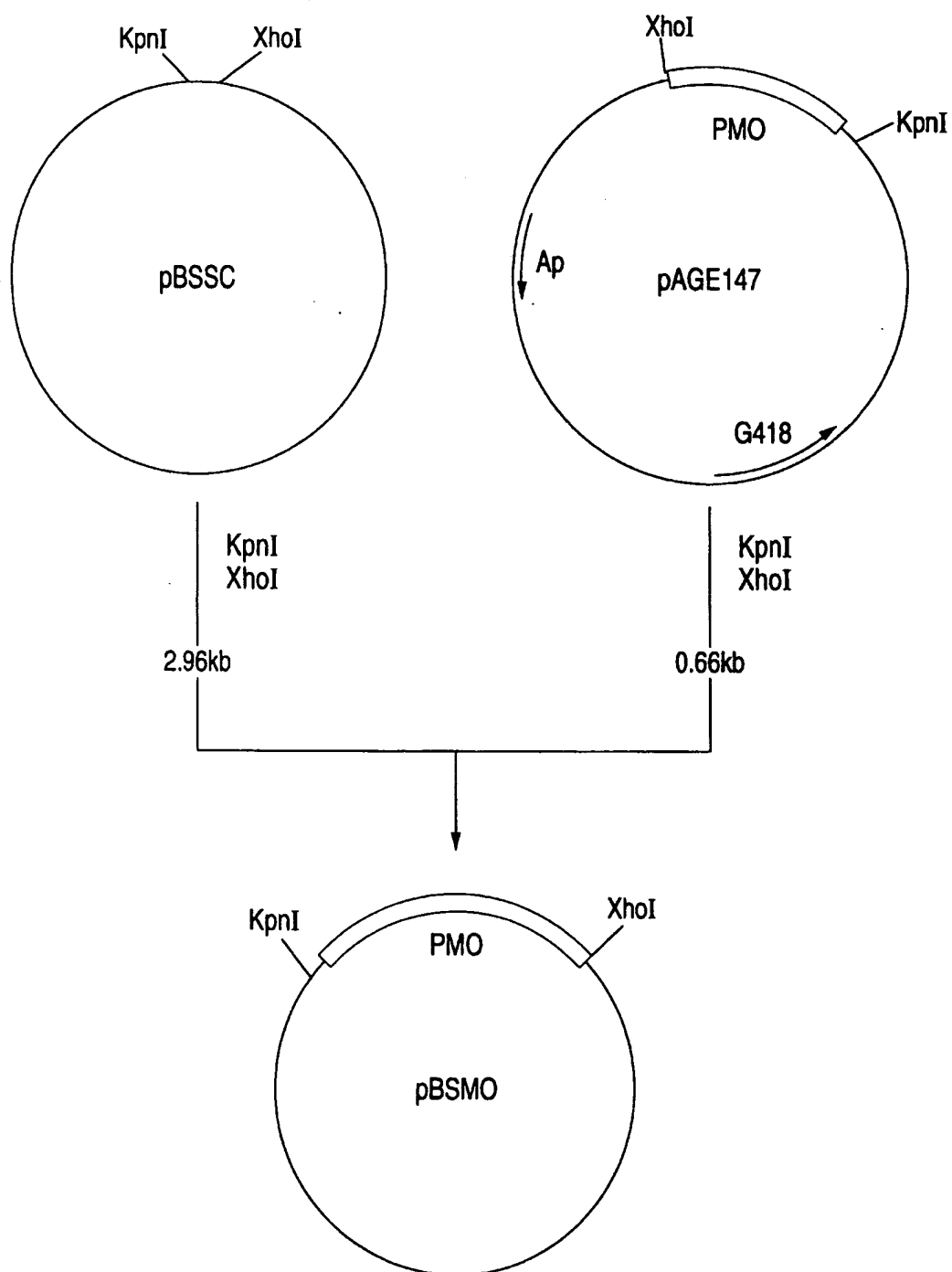


FIG. 13

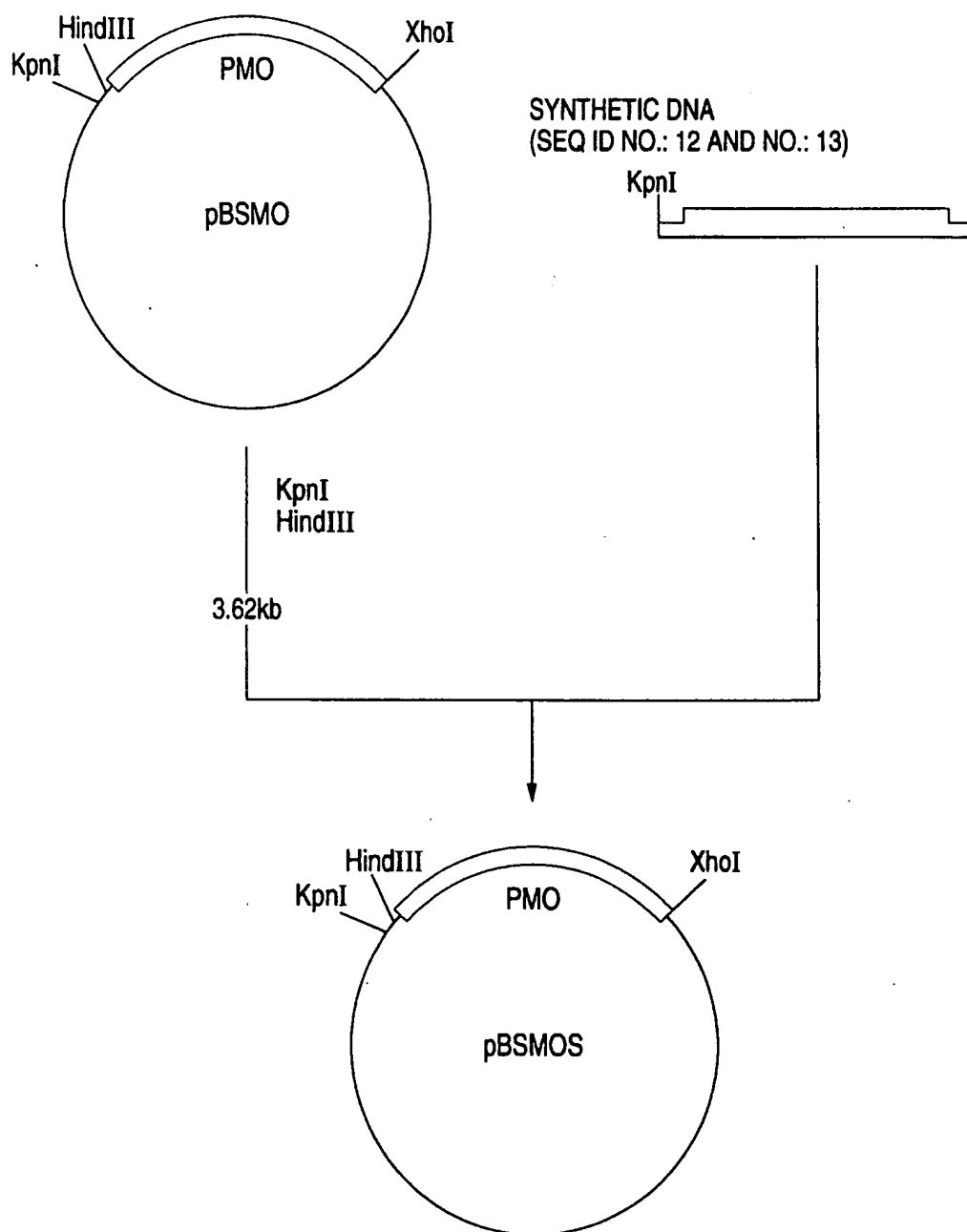


FIG. 14

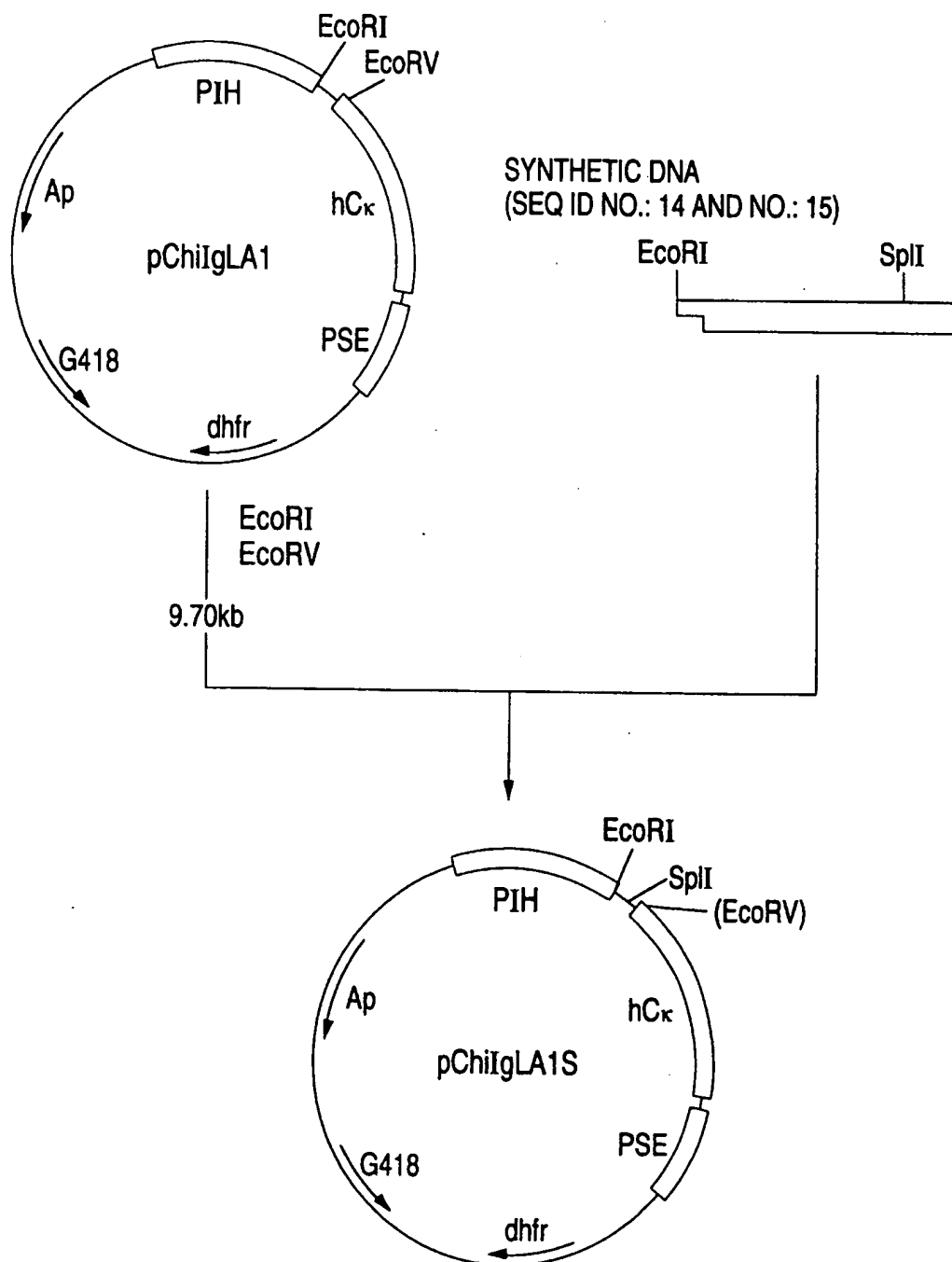


FIG. 15

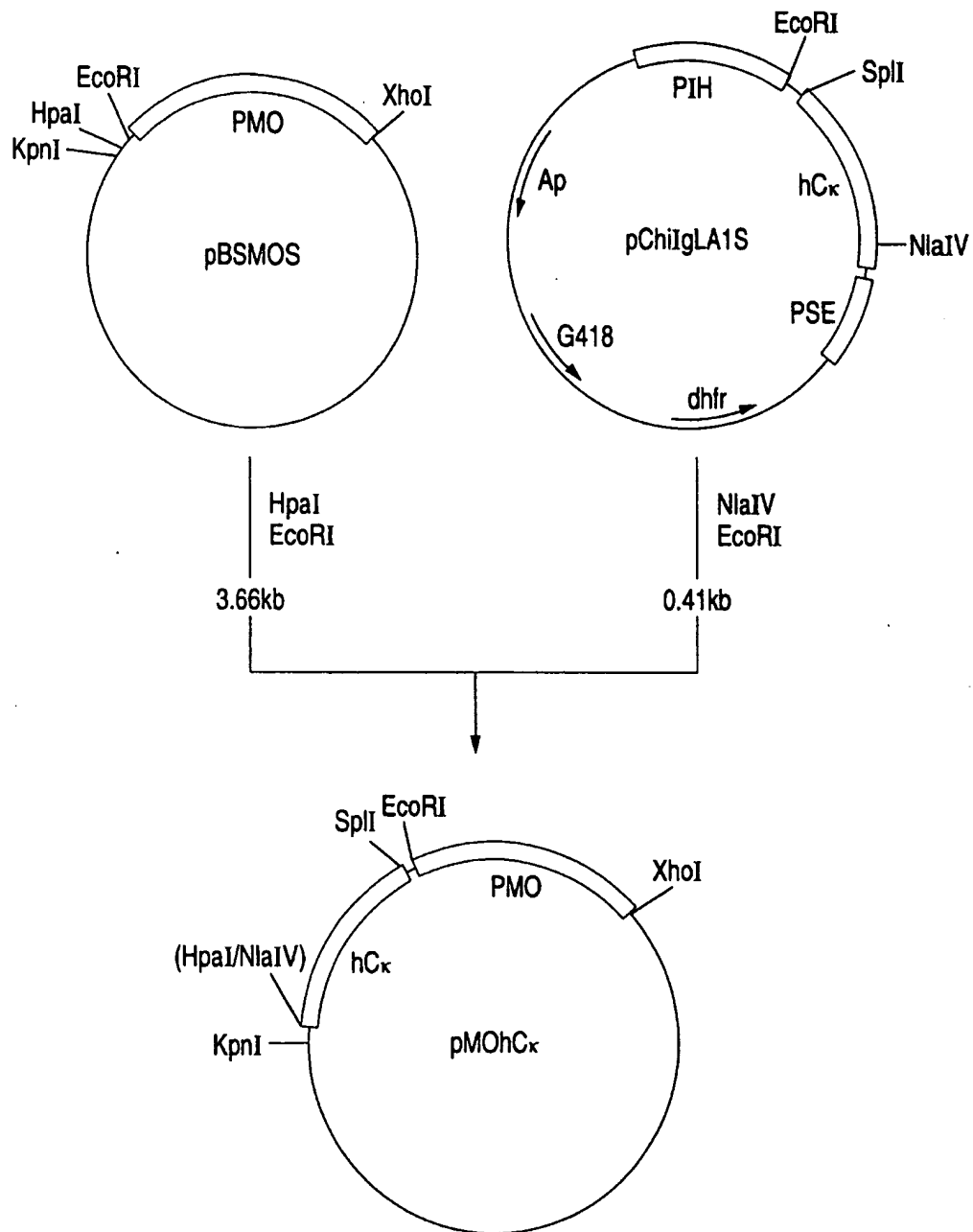


FIG. 16

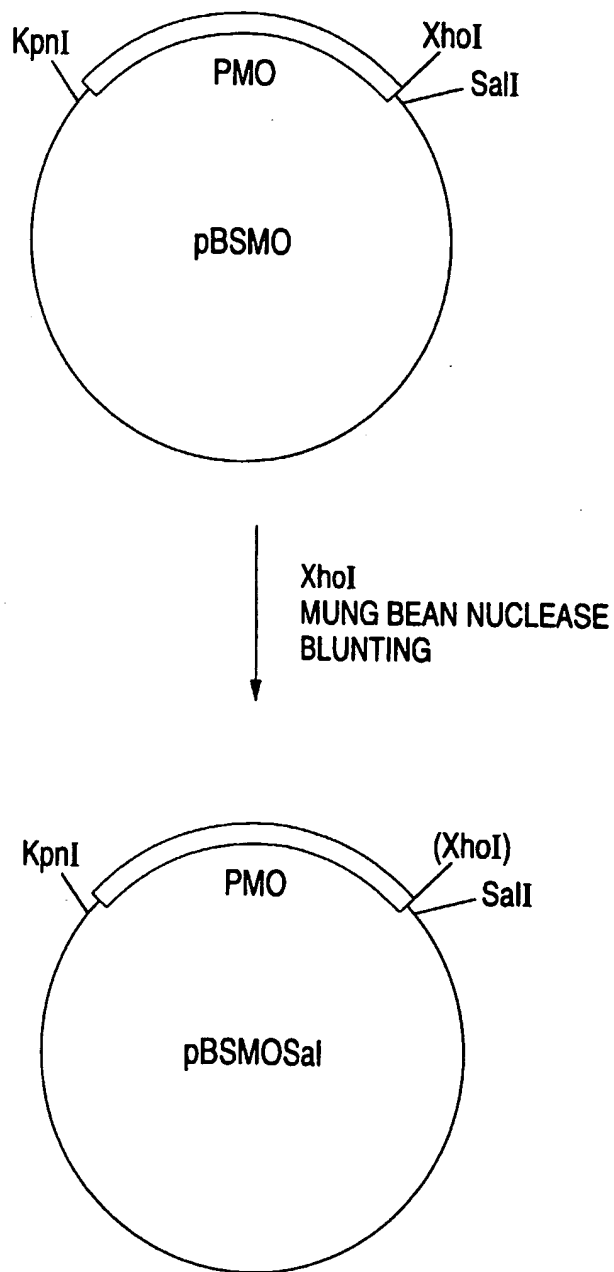


FIG. 17

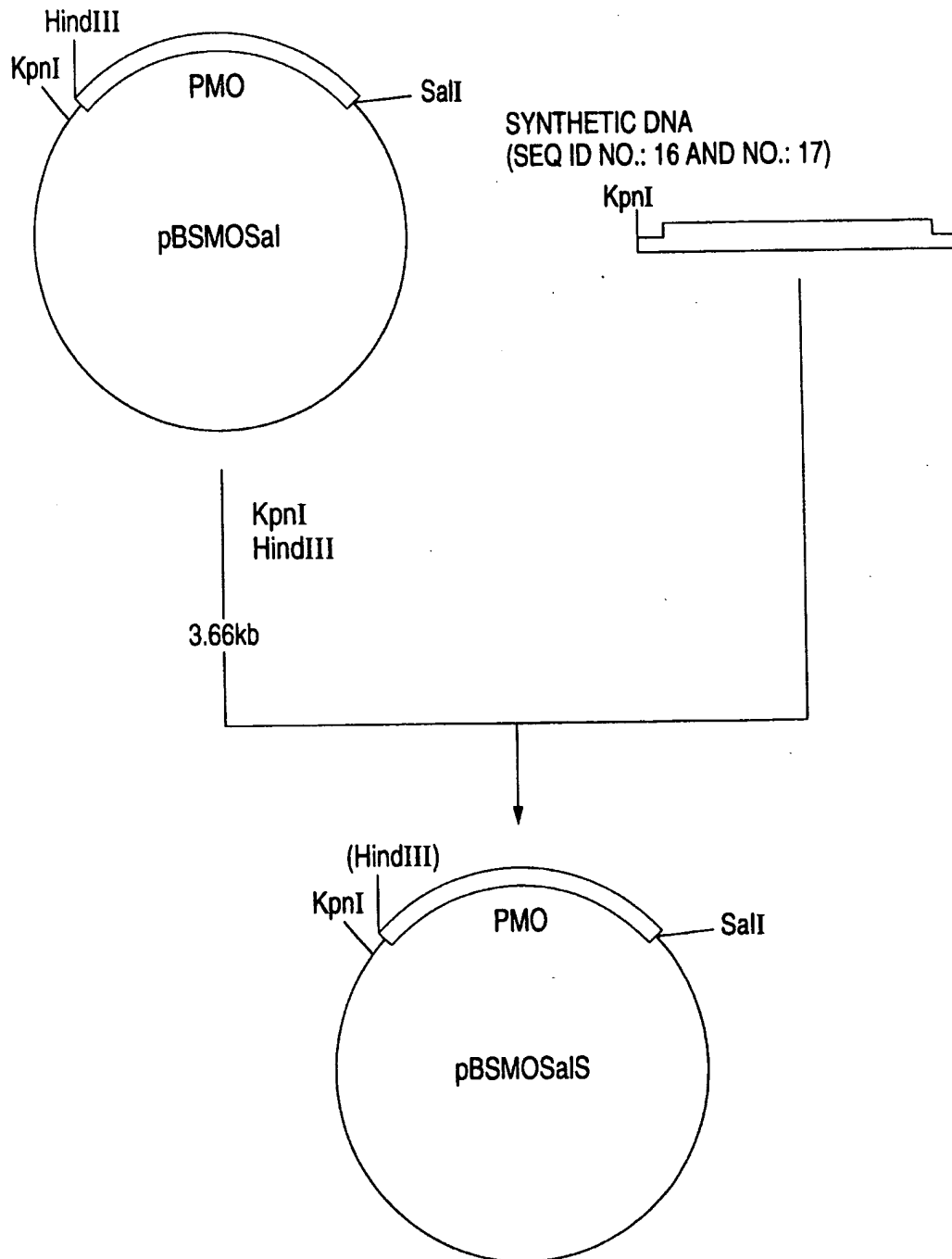


FIG. 18

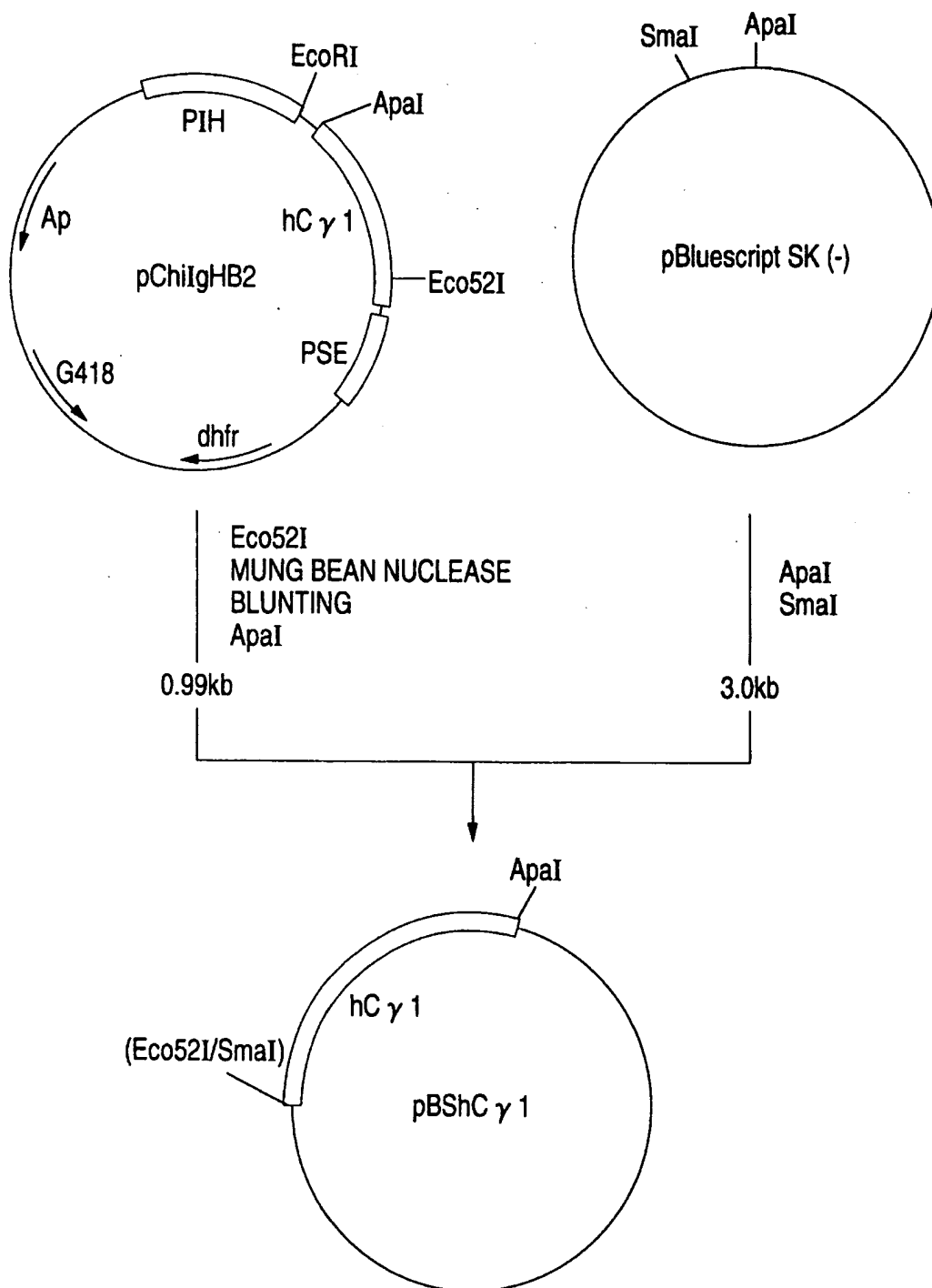


FIG. 19

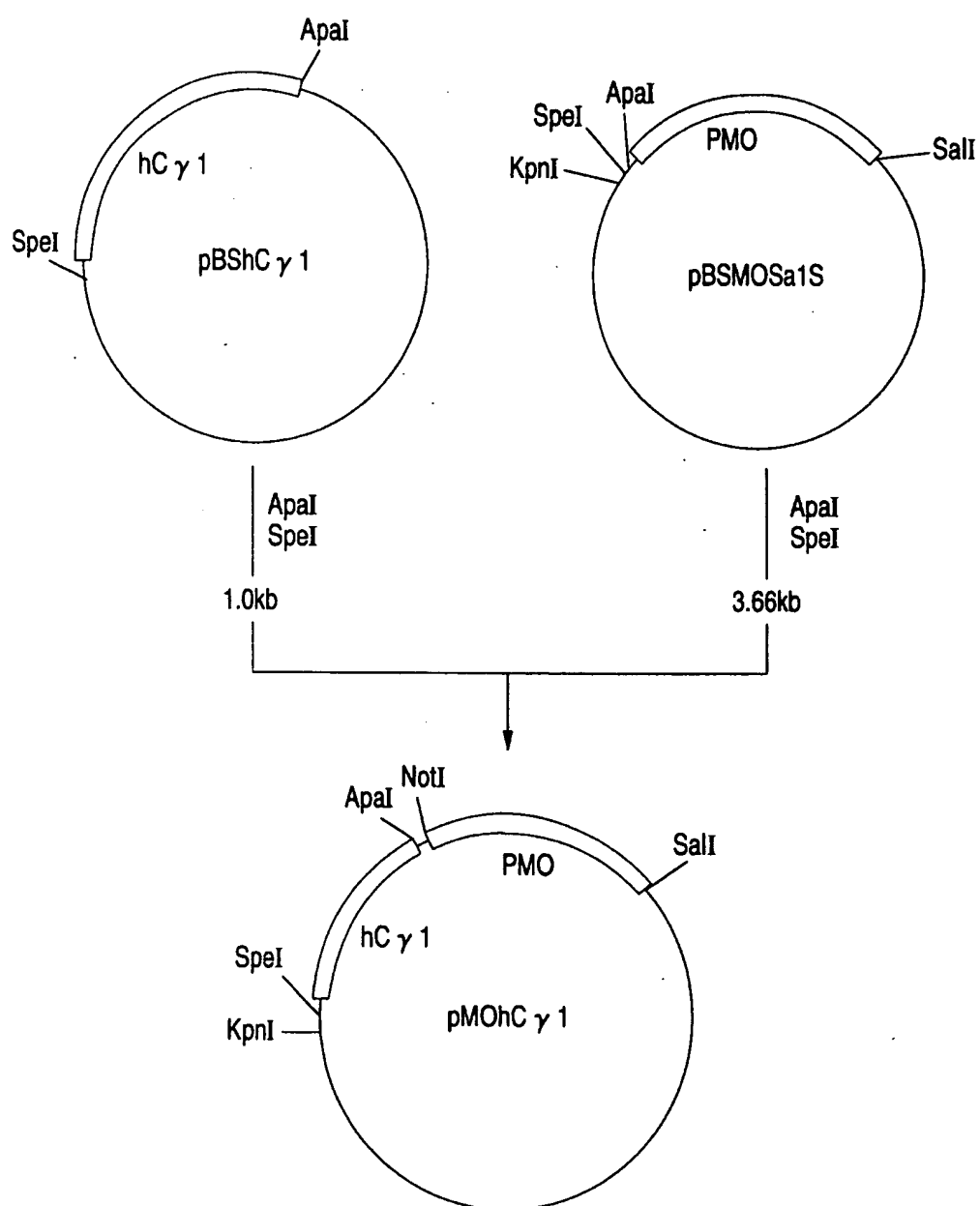


FIG. 20

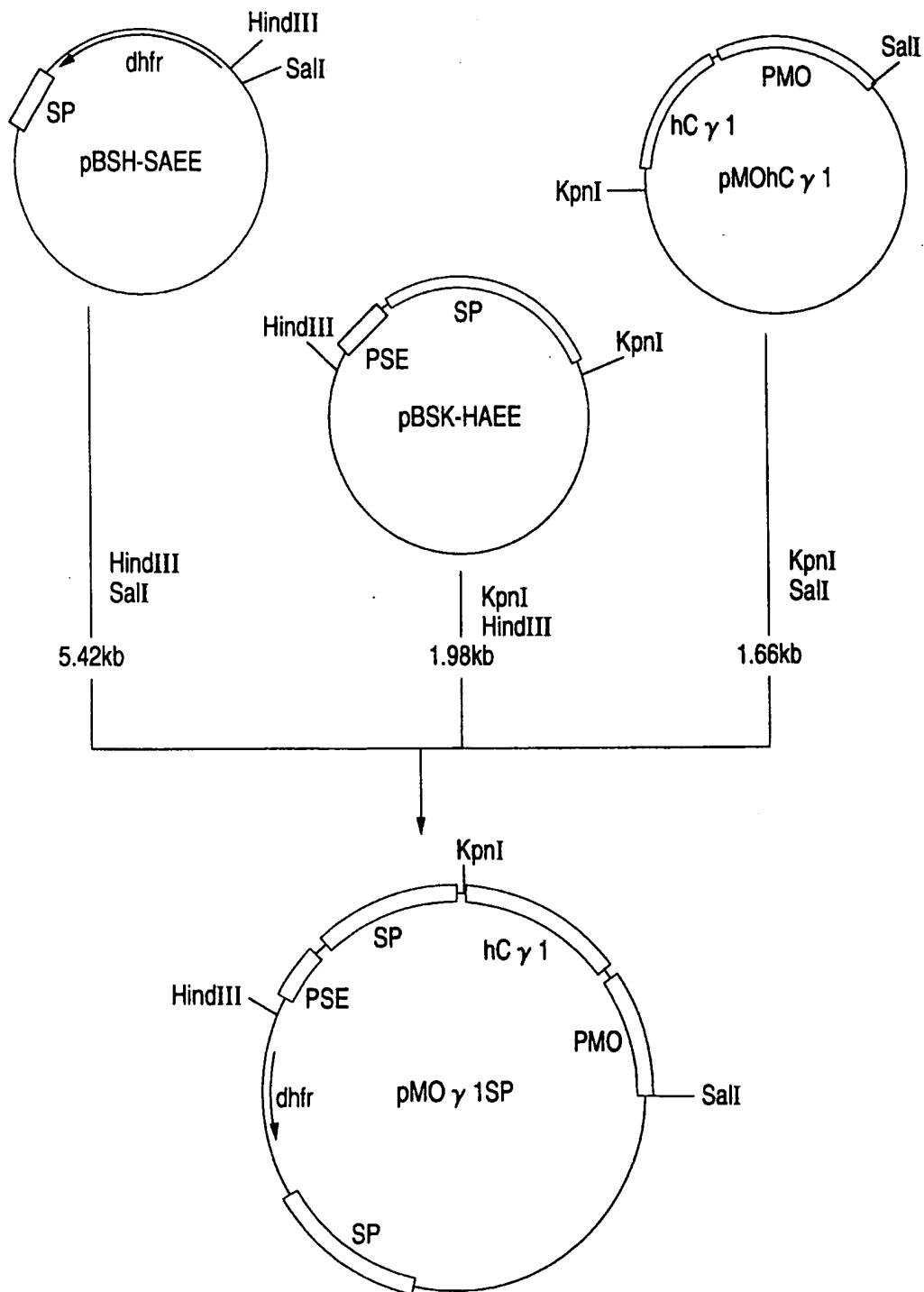


FIG. 21

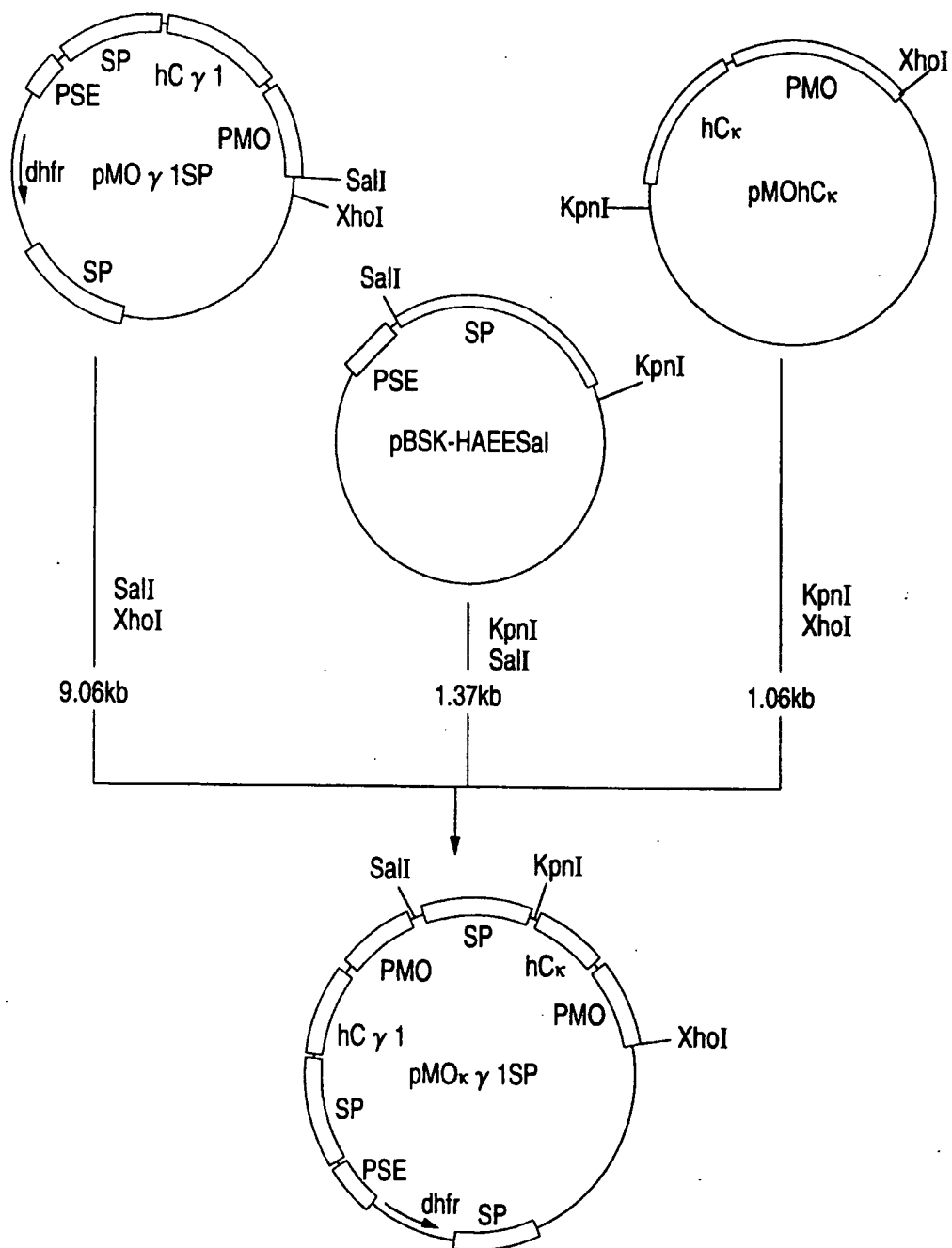


FIG. 22

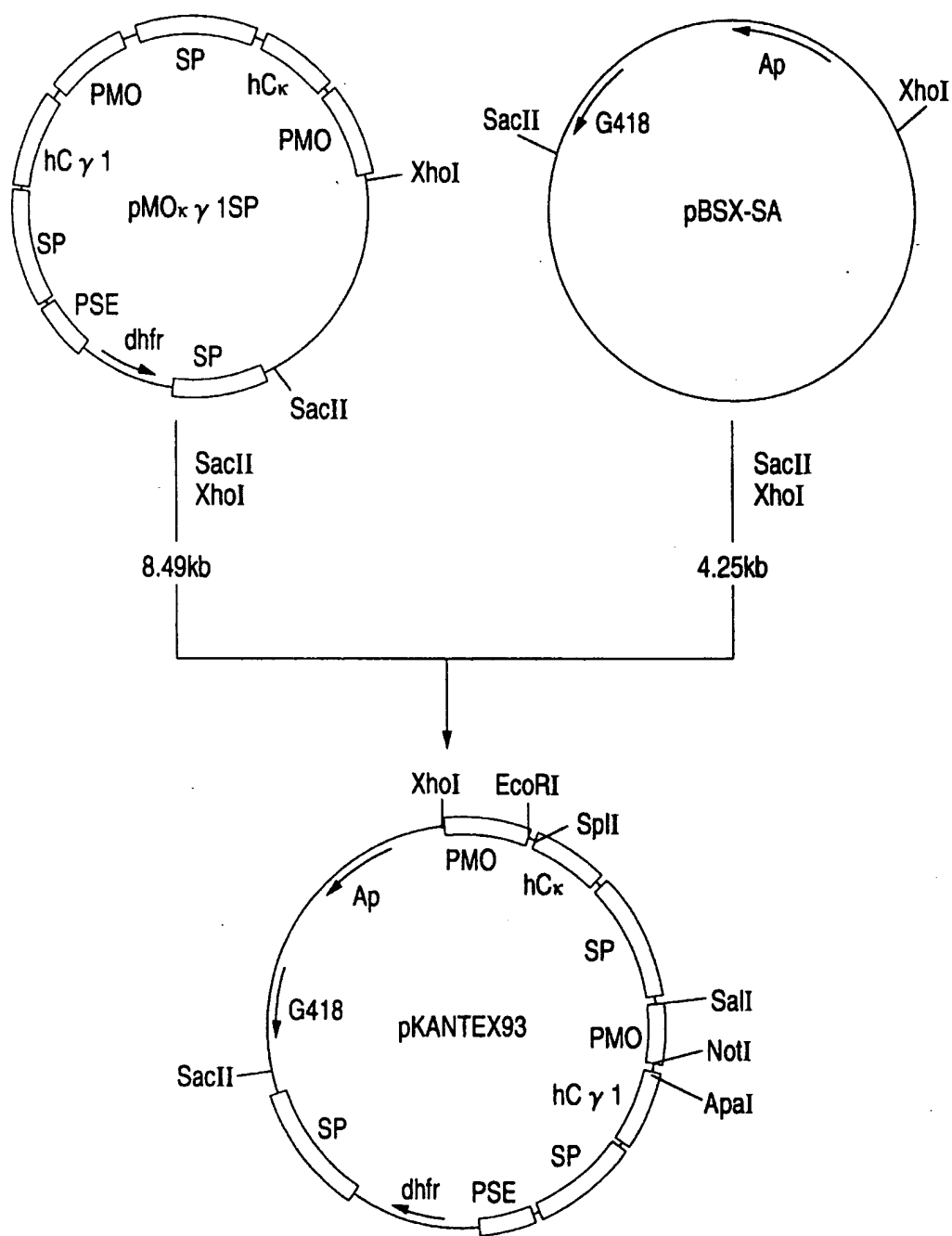


FIG. 23

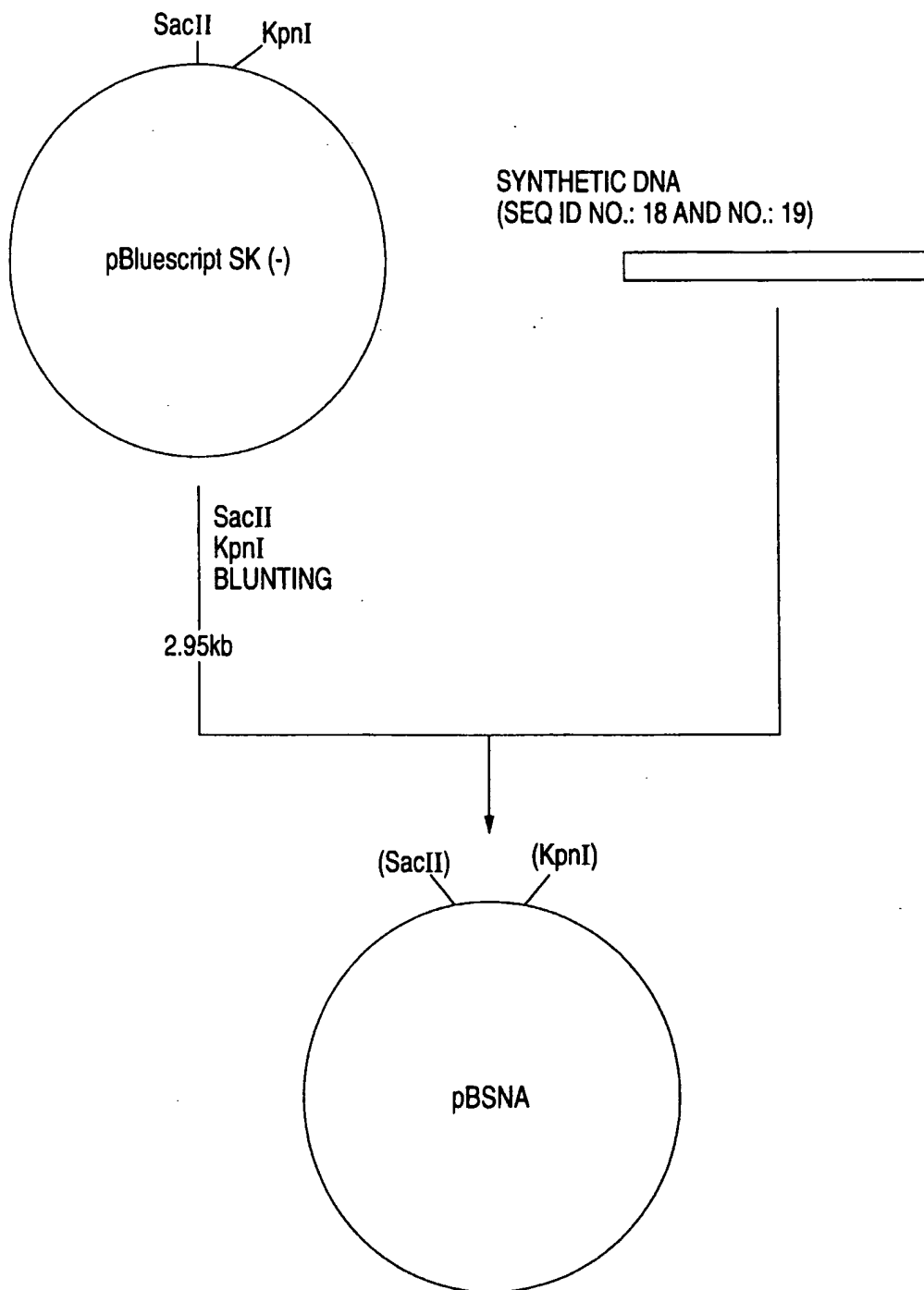


FIG. 24

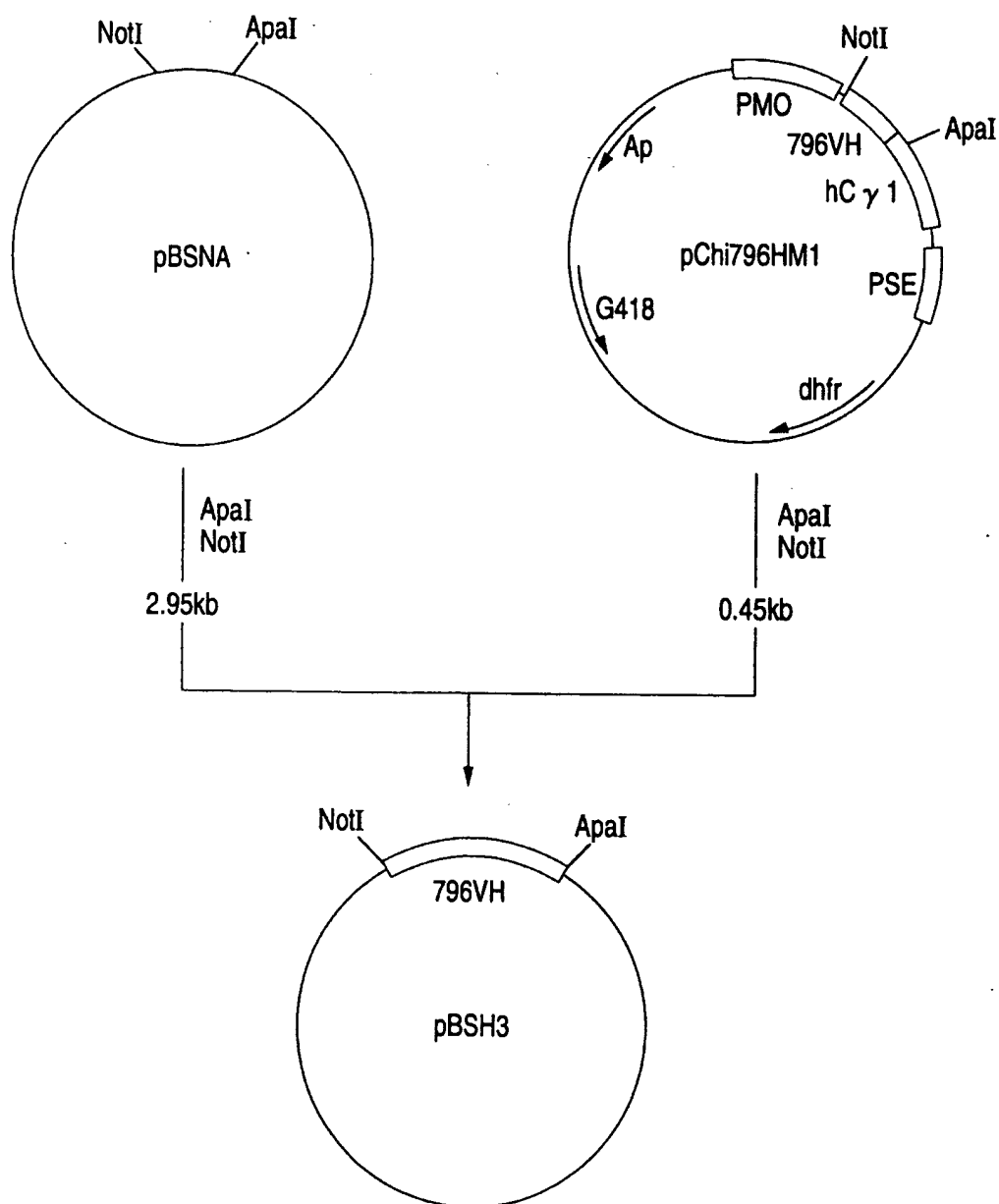


FIG. 25

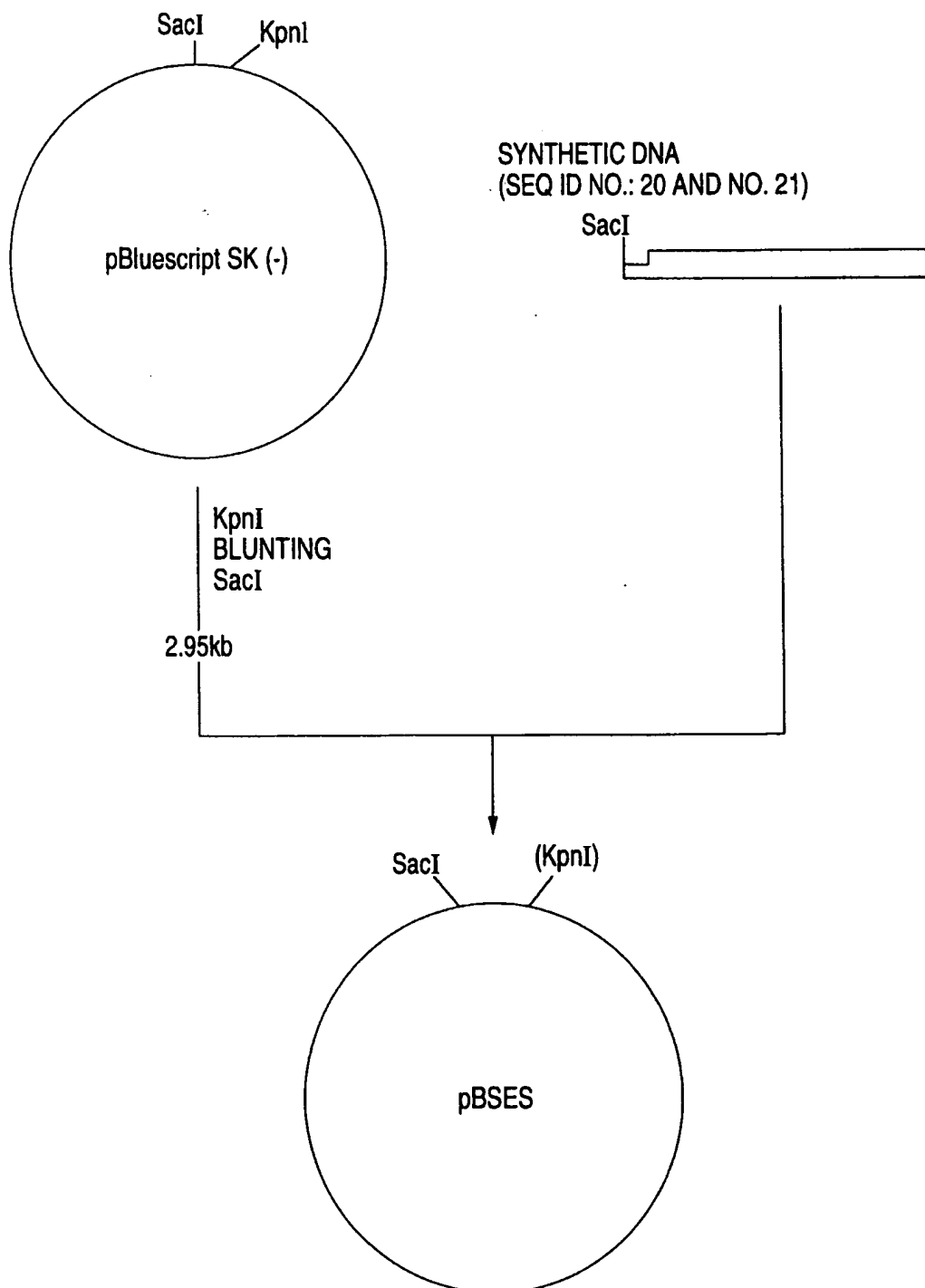


FIG. 26

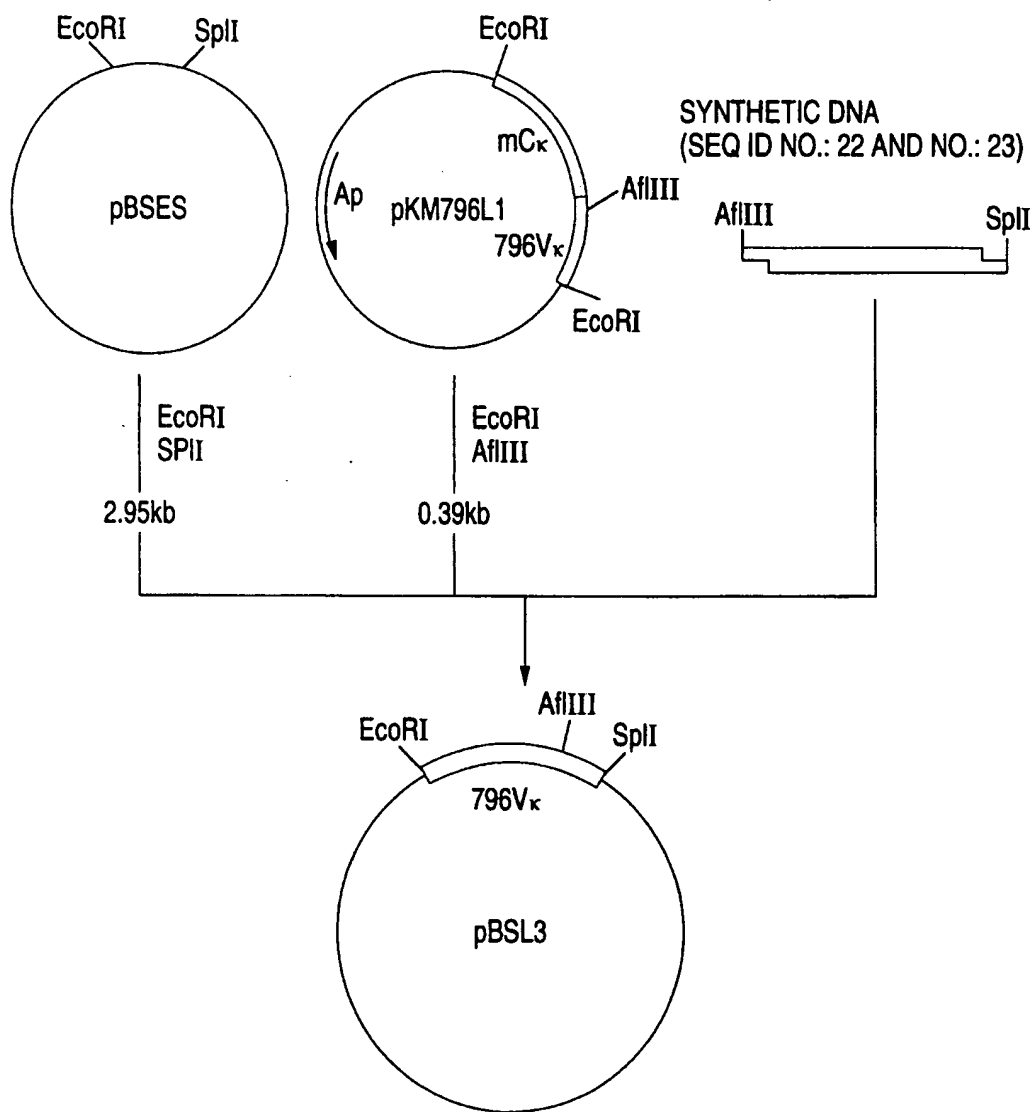


FIG. 27

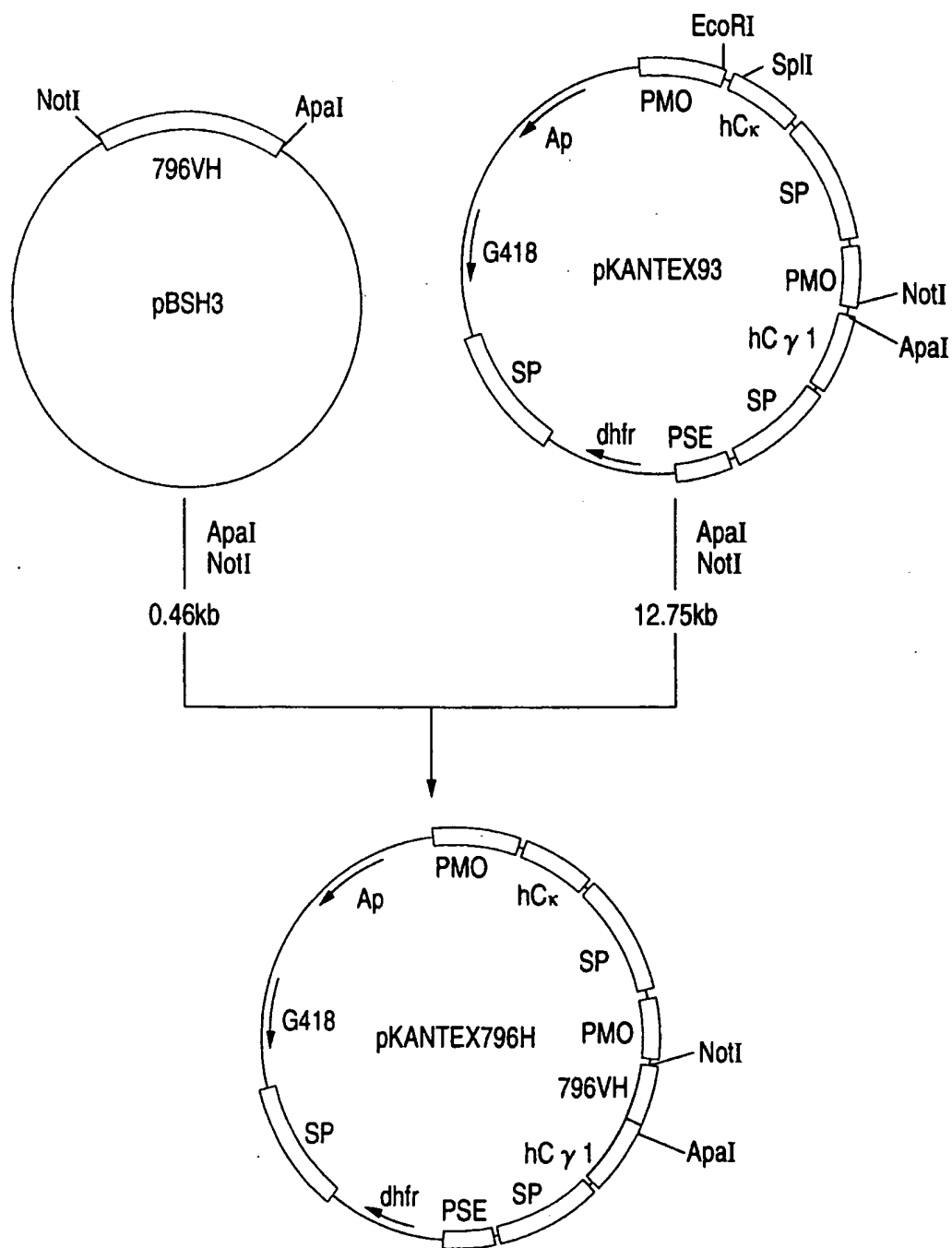


FIG. 28

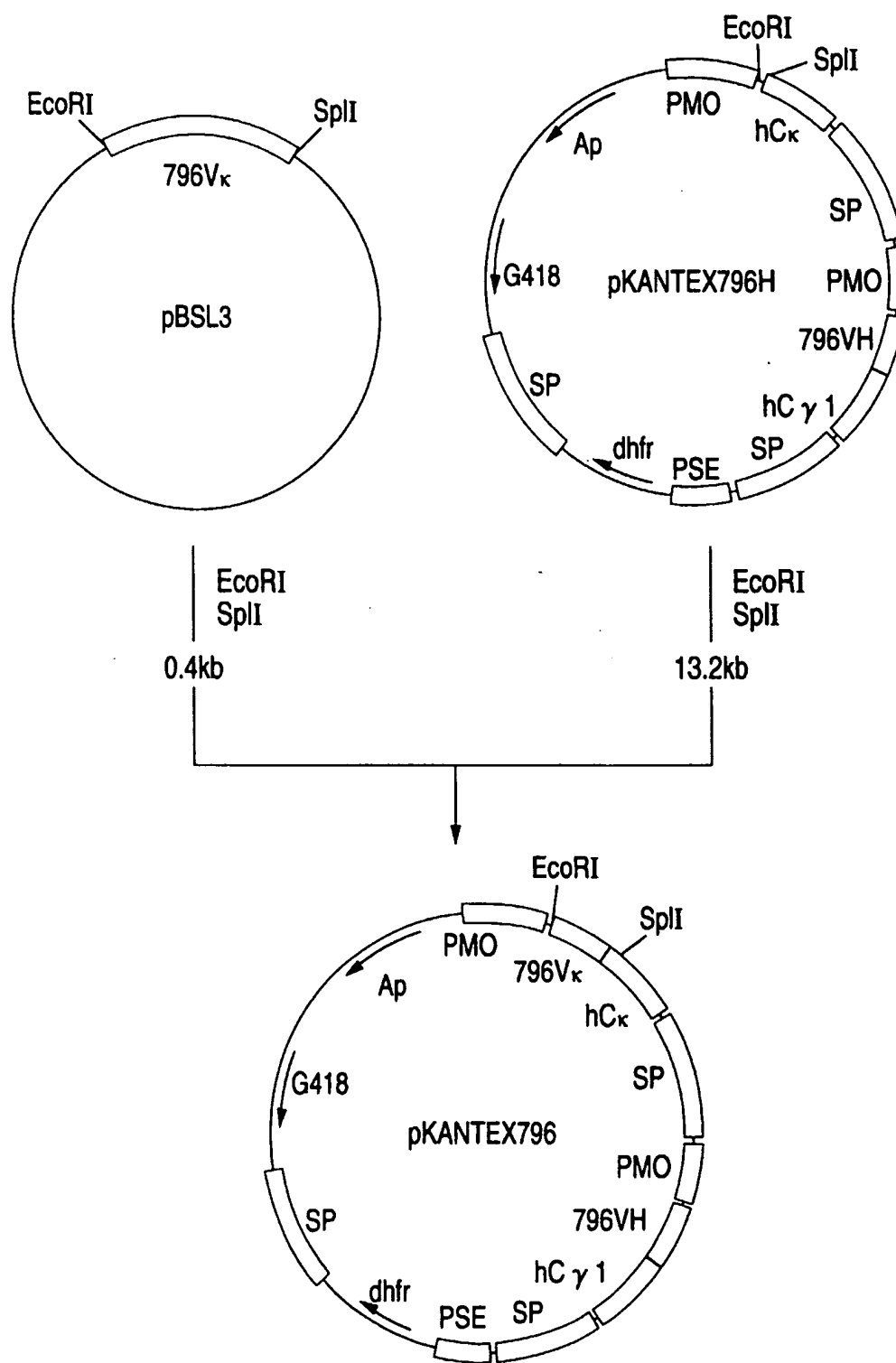


FIG. 29

